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**Brief Definitive Report**

**Type I Interferon Sensitizes Lymphocytes to Apoptosis and Reduces Resistance to *Listeria* Infection**

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**Abstract**

Infection with *Listeria monocytogenes* causes lymphocyte apoptosis that is mediated by the actions of the pore-forming virulence factor listeriolysin O (LLO). Previous work showed that activated lymphocytes were highly sensitive to LLO-induced apoptosis, whereas resting lymphocytes were less susceptible. We now show that mice deficient in the type I interferon (IFN) receptor were more resistant to *Listeria* infection and had less apoptotic lesions than wild-type counterparts. Furthermore, treatment of resting splenic lymphocytes with recombinant IFN-αA enhanced their susceptibility to LLO-induced apoptosis. Together, these data suggest that type I IFN signaling is detrimental to handling of a bacterial pathogen and may enhance the susceptibility of lymphocytes undergoing apoptosis in response to bacterial pore-forming toxins.

Key words: apoptosis • cytokines • *Listeria* • inflammation • T lymphocytes

**Introduction**

Previous studies with *Listeria monocytogenes* have exposed the role of multiple cytokines at different stages of the infection using gene-ablated mice or neutralizing monoclonal antibodies (for review see references 1 and 2). IL-12, TNF, and the IL-1 family of proteins participate in activating different cells and controlling *Listeria* growth. These early cytokines contribute to the induction of IFN-γ, a product of NK and T cells, and activation of macrophages.

Much is known about the role of type I IFNs (IFN-α and IFN-β) in the antiviral response and type II IFN (IFN-γ) in the antiviral and antibacterial response (3–6). However, there exists a paucity of data on the role of type I IFNs during bacterial infection. Here we report that the absence of the shared IFN-α/β receptor (IFN-αβR−/−) provides mice with an advantage during *Listeria* infection. This resistance was accompanied by an amelioration of the lymphocyte apoptotic process that took place during the early exponential growth of *Listeria* in tissues.

We and others have described an early transitory phase of lymphocyte apoptosis in infective foci that peaks at 48 h after infection (7, 8). Experiments ex vivo indicated that listeriolysin O (LLO), a pore-forming molecule and major virulence factor, caused lymphocyte apoptosis particularly in cells that were replicating (9). Moreover, injection of pure LLO subcutaneously led to lymphocyte apoptosis in the T cell–rich zones of draining lymph nodes. Of note is that during the in vivo infection the apoptotic lesions were either not affected or were increased in mice in which IL-1, IL-12, or IFN-γ were either neutralized or not produced (7). The lesions were unrelated to activation-induced cell death, to Fas–Fas ligand interactions, or to the production of reactive oxygen or nitrogen intermediates. In toto the evidence points to the release of soluble LLO during the early robust growth of *Listeria* as the mechanism leading to the apoptotic death of lymphocytes in infective foci. In fact, neutralization of LLO by injection of monoclonal antibody to LLO controlled the infection as well as the intensity of the apoptotic lesions (10). The data presented herein suggests that type I IFN sensitizes lymphocytes to undergo apoptosis during *Listeria* infection, and that this has a negative effect on bacterial handling in the mouse.

**Materials and Methods**

*Mice and Listeria Infection.* Wild-type and IFN-αβR−/− mice on the 129Sv/Ev genetic background were provided by H.W. Virgin IV (Washington University School of Medicine, St. Louis, MO; reference 3). *L. monocytogenes* strain EGD was used as before for intraperitoneal infection (7). Histological examination of hematoxylin and eosin– and TdT-mediated dUTP nick-end labeling (TUNEL)-stained sections was performed on the spleens from infected mice (7). Cytometric bead array (CBA) analysis was performed with the Mouse Inflammation Kit and used according to the manufacturer’s instructions with 50 μL serum per assay (BD Biosciences). CBA data was analyzed on a FACSCalibur™ with CELLQuest™ software and the CBA analysis soft-
ware package (BD Biosciences). For flow cytometric staining, portions of spleens from infected mice were disrupted to generate single cell suspensions. Cells were then stained for CD69 and either CD4 or CD8 using conventional techniques. All antibodies were obtained from BD Biosciences.

Cell Cultures. A CD4 T cell line reactive to ovalbumin was generated from normal 129Sv/J mice (The Jackson Laboratory) immunized with an ovalbumin emulsion in complete Freund’s adjuvant using previously described techniques (9). The line was passaged every 10–12 d by the addition of irradiated 129Sv/J splenocytes (3,000 rads), 10 μM ovalbumin, and 50 U/ml IL-2. The usual behavior was that of proliferation that slowly stopped by about day 8. On day 10 after stimulation, the T cells were harvested and were not in cell cycle. Assays with the T cell line were performed as described previously (9). Whole splenocytes were isolated from 129Sv/Ev or IFN-αβR−/− mice. Single cell suspensions were made and cells were plated at a density of 5 × 10^6 cells/ml in DMEM and 10% FCS, and treated with recombinant mouse IFN-α at 1, 10, or 100 U/ml (specific activity: 4.8 × 10^7 U/mg; PBL Biomedical Laboratories). Cells were incubated for 24 h, and then the nonadherent cells were removed and purified. Both the T cell line and the splenocytes were purified over a Histopaque 1119 gradient (Sigma-Aldrich). After Histopaque, cells were resuspended in DMEM containing 1% FCS, and then treated with 250 ng/ml of purified recombinant LLO (4.4 nM) for 6 h. Cells were stained with annexin V-PE and 7-AAD (BD Biosciences), and analyzed by flow cytometry. For assays involving splenocytes, cells were also stained with anti-CD3-APC (BD Biosciences) to identify T cells.

Results and Discussion

Listeria Infection. Infection of 129Sv/Ev mice with Listeria showed the expected increase in the number of organisms in the spleen and liver as a function of time (Fig. 1, A and B). IFN-αβR−/− mice had the same number of Listeria as wild-type mice 24 h after infection. Therefore, there was no difference between the two strains during the initial innate resistance to infection, which was mediated by neutrophils and macrophages (1). However, at 48 h after infection, there was about a 20-fold increase in CFUs in both the spleens and livers of wild-type versus IFN-αβR−/− mice. By 96 h after infection, IFN-αβR−/− mice had no further Listeria growth and showed ~1,200-fold fewer CFUs per organ (Fig. 1, A and B).

At 48 h after infection, the peak time at which lymphocyte apoptosis was evident, histological examination showed a marked difference between both sets of mice (Fig. 2). The difference in lymphocyte apoptosis was seen at three different infectious doses (Table I). Wild-type mice showed infective foci around the periarteriolar lymphoid sheath, characterized by a paucity of lymphocytes and the presence of...
In pathology, the percentage refers to the number of white pulp profiles showing the lesions. Severe means extensive depletion of lymphocytes, mostly in the periarteriolar lymphoid sheath but extending to peripheral areas of the pulp. Mild refers to a slight reduction localized to the periarteriolar lymphoid sheath. TUNEL analysis shows varying numbers of positive nuclei. Slides were scored blindly by one of us.

Table I. Spleens Examined 48 h after the Indicated Dose of Listeria

<table>
<thead>
<tr>
<th>L. monocytogenes dose</th>
<th>Mice</th>
<th>Pathology (48 h)</th>
<th>Apoptosis (48 h)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 × 10^3</td>
<td>WT</td>
<td>4 30–50% – severe</td>
<td>3+</td>
</tr>
<tr>
<td>5 × 10^3</td>
<td>IFN-αβR−/−</td>
<td>5 10% – mild</td>
<td>1+</td>
</tr>
<tr>
<td>2.5 × 10^4</td>
<td>WT</td>
<td>4 50–100% – severe</td>
<td>4+</td>
</tr>
<tr>
<td>2.5 × 10^4</td>
<td>IFN-αβR−/−</td>
<td>5 10% – mild</td>
<td>1+</td>
</tr>
<tr>
<td>5 × 10^4</td>
<td>WT</td>
<td>5 75–100% – severe</td>
<td>4+</td>
</tr>
<tr>
<td>5 × 10^4</td>
<td>IFN-αβR−/−</td>
<td>6 20% – mild</td>
<td>1–2+</td>
</tr>
</tbody>
</table>

In pathology, the percentage refers to the number of white pulp profiles showing the lesions. Severe means extensive depletion of lymphocytes, mostly in the periarteriolar lymphoid sheath but extending to peripheral areas of the pulp. Mild refers to a slight reduction localized to the periarteriolar lymphoid sheath. TUNEL analysis shows varying numbers of positive nuclei. Slides were scored blindly by one of us.
We first tested CD4+ T lymphocytes from ex vivo culture 10 d after antigen stimulation by incubating them in the presence or absence of IFN-αA for 24 h, and then exposing them to 250 ng/ml LLO for 6 h. We selected this time point on the basis of previous kinetic studies on cultured T cells treated with LLO (9). In the absence of LLO, 8–10% of cells underwent apoptosis (i.e., became annexin V+/7-AAD-). 6 h of treatment with LLO increased this number to 22%, in accordance with our previous study (Fig. 3, A, D, and J; reference 9). The set of cells staining with both annexin V and 7-AAD also increased upon LLO treatment. Our previous study demonstrated that LLO-treated cells shifted over time from the single positive (annexin V+/7-AAD-) to the double positive (annexin V+/7-AAD+), the latter of which contains late apoptotic as well as necrotic cells (9).

Treatment with IFN-αA alone did not significantly increase the number of apoptotic cells. Annexin V+/7-AAD- cells constituted, on average, 8, 8, and 10% of the cells after treatment with 1, 10, and 100 U/ml IFN-αA, respectively (Fig. 3 J). However, treatment with IFN-αA augmented the number of cells undergoing LLO-induced apoptosis, in a dose-dependent manner. Cells treated with 1, 10, and 100 U/ml IFN-αA and then exposed to 250 ng/ml LLO had annexin V+/7-AAD- populations of 29, 38, and 43%, respectively (Fig. 3, G and J).

To extend our results to primary cells, we isolated splenocytes, cultured them for 24 h in increasing doses of IFN-αA, treated them with LLO, and assayed them for apoptosis by annexin V/7-AAD staining. As an average, 13% of untreated CD3+ cells were annexin V+/7-AAD- (Fig. 3, B and K). This number increased to 24% upon LLO treatment (Fig. 3, E and K). The addition of type I IFN increased the response of T cells to LLO-induced apoptosis in a dose-dependent manner. At the highest IFN-αA dose tested (100 U/ml), 37% of cells CD3+ became annexin V+/7-AAD- (Fig. 3, H and K). This increase, albeit modest, was highly reproducible in three experiments. In contrast, type I IFN did not increase LLO-induced apoptosis in splenocytes isolated from IFN-αR-/- mice. The percentage increase in annexin V+/7-AAD- cells taken from IFN-αR-/- spleen after treatment with LLO was the same in the absence of IFN-αA (Fig. 3, F and L) and at all doses of IFN-αA tested (Fig. 3, I and L).
increased the number of cells at the same time points to 35, 42, and 37%. Spleen cells from the IFN-αR−/− mice cultured for 2, 4, and 6 h with LLO resulted in 30, 32, and 30%, respectively. However, this number did not increase after culture in IFN-αA: 29, 31, and 30%. Both untreated spleen cell populations were 16% annexin V+/7-AAD− cells after 6 h of culture. Thus, there were no major changes in kinetics during the 2–6-h time period. Fig. 3 shows the 6 h results.

Comments. Our experiments with IFN-αβR−/− mice indicate a deleterious effect on early resistance to Listeria infection by type I IFNs. Fehr et al. (16) reported that IFN-αβR−/− mice were not impaired in their resistance to Listeria infection. Moreover, their results suggested an increased resistance to Listeria at day 5 after infection when compared with wild-type controls. They hypothesized that this was a function of type I and II IFN receptor competition for components of signaling pathways. However, that study, which focused on the role of transcription factors and nitric oxide in resistance to Listeria, did not fully explain the increased resistance of IFN-αβR−/− mice.

Our report expands on previous findings in the following ways. In the absence of a response to type I IFNs (i.e., in the receptor null mice), the infective foci contained fewer TUNEL+ lymphocytes as well as fewer activated T cells. Type I IFN unresponsive mice were also more resistant to infection. We believe that early type I IFN production during Listeria infection results in nonspecific activation of T cells. The provocative finding is that this early activation also sensitizes these cells to the apoptogenic actions of LLO. Furthermore, macrophages treated with type I IFN were shown to be susceptible to Listeria killing in vitro (17). These results suggest that type I IFN could enhance the susceptibility to death of multiple leukocyte subsets during Listeria infection.

The immunological consequences of cellular death during Listeria infection of the mouse must be further examined, but could involve the development of responses that are inhibitory for bacterial clearance. Clearance of apoptotic bodies by macrophages is generally thought to down-regulate inflammation through macrophage release of TGF-β, prostaglandin E2, and platelet-activating factor (18–20). Apoptotic lymphocytes themselves may also down-regulate inflammation through the release of preformed TGF-β upon apoptosis induction (21). We suggest an inverse relationship between death by apoptosis, in part enhanced by type I IFNs and the growth of Listeria. In the absence of type I IFN signaling, apoptosis is limited, and less inhibition of phagocyte antimicrobial processes takes place. In support of these hypotheses, we have observed that SCID mice, lacking T and B lymphocytes, do not show apoptotic lesions after Listeria infection, and are more resistant than wild-type mice during the first 2–4 d after infection (22 and unpublished data).

The cellular source of type I IFN during Listeria infection and the Listeria component(s) inducing its production remain unknown at this time. Candidate ligands would include Toll-like receptor (TLR)4 or TLR9 agonists such as lipoteichoic acid or CpG DNA motifs, which have been shown to induce type I IFN expression by dendritic cells (23). Analysis of TLR4−/−, TLR9−/−, and MyD88−/− mice or cells may yield insight into the generation of type I IFN in response to Listeria infection. Macrophages recognizing cytosolic Listeria also generate IFN-β (15), suggesting this cell type as a source for the cytokine. Alternatively, Listeria could generate proteins that promote expression of type I IFN to enhance its virulence and dissemination.

A classical role ascribed to type I IFN signaling is the enhancement of cancer cell and virus-infected cell apoptosis. One possible mechanism for type I IFN’s sensitization of T cells to undergo apoptosis would be the accumulation of the p53 tumor suppressor gene (24). After p53 expression, cellular stress caused by LLO disruption of the plasma membrane might then lead to apoptosis induction. Further studies will examine the role of type I IFNs in LLO-induced apoptosis.

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