

Washington University School of Medicine

Digital Commons@Becker

Open Access Publications

2010

Target enzyme mutations are the molecular basis for resistance towards pharmacological inhibition of nicotinamide phosphoribosyltransferase

Uffe H. Olesen
Rigshospitalet

Jakob G. Petersen
Rigshospitalet

Antje Garten
University of Leipzig

Wieland Kiess
University of Leipzig

Jun Yoshino
Washington University School of Medicine

Follow this and additional works at: https://digitalcommons.wustl.edu/open_access_pubs
See next page for additional authors



Part of the [Medicine and Health Sciences Commons](#)

Please let us know how this document benefits you.

Recommended Citation

Olesen, Uffe H.; Petersen, Jakob G.; Garten, Antje; Kiess, Wieland; Yoshino, Jun; Imai, Shin-Ichiro; Christensen, Mette K.; Fristrup, Peter; Thougard, Annemette V.; Björklind, Fredrik; Jensen, Peter B.; Nielsen, Søren J.; and Sehested, Maxwell, "Target enzyme mutations are the molecular basis for resistance towards pharmacological inhibition of nicotinamide phosphoribosyltransferase." *BMC Cancer*. 10, 677. (2010).

https://digitalcommons.wustl.edu/open_access_pubs/53

This Open Access Publication is brought to you for free and open access by Digital Commons@Becker. It has been accepted for inclusion in Open Access Publications by an authorized administrator of Digital Commons@Becker. For more information, please contact vanam@wustl.edu.

Authors

Uffe H. Olesen, Jakob G. Petersen, Antje Garten, Wieland Kiess, Jun Yoshino, Shin-Ichiro Imai, Mette K. Christensen, Peter Fristrup, Annemette V. Thougard, Fredrik Björkling, Peter B. Jensen, Søren J. Nielsen, and Maxwell Sehested

RESEARCH ARTICLE

Open Access

Target enzyme mutations are the molecular basis for resistance towards pharmacological inhibition of nicotinamide phosphoribosyltransferase

Uffe H Olesen^{1,2*}, Jakob G Petersen^{1,2}, Antje Garten³, Wieland Kiess³, Jun Yoshino⁴, Shin-Ichiro Imai⁴, Mette K Christensen², Peter Fristrup⁵, Annemette V Thougard², Fredrik Björkling^{2,6}, Peter B Jensen², Søren J Nielsen², Maxwell Sehested^{1,2}

Abstract

Background: Inhibitors of nicotinamide phosphoribosyltransferase (NAMPT) are promising cancer drugs currently in clinical trials in oncology, including APO866, CHS-828 and the CHS-828 prodrug EB1627/GMX1777, but cancer cell resistance to these drugs has not been studied in detail.

Methods: Here, we introduce an analogue of CHS-828 called TP201565 with increased potency in cellular assays. Further, we describe and characterize a panel of cell lines with acquired stable resistance towards several NAMPT inhibitors of 18 to 20,000 fold compared to their parental cell lines.

Results: We find that 4 out of 5 of the resistant sublines display mutations of NAMPT located in the vicinity of the active site or in the dimer interface of NAMPT. Furthermore, we show that these mutations are responsible for the resistance observed. All the resistant cell lines formed xenograft tumours *in vivo*. Also, we confirm CHS-828 and TP201565 as competitive inhibitors of NAMPT through docking studies and by NAMPT precipitation from cellular lysate by an analogue of TP201565 linked to sepharose. The NAMPT precipitation could be inhibited by addition of APO866.

Conclusion: We found that CHS-828 and TP201565 are competitive inhibitors of NAMPT and that acquired resistance towards NAMPT inhibitors can be expected primarily to be caused by mutations in NAMPT.

Background

Drug resistance is a serious concern in the treatment of cancer [1]. It can occur as either *de novo* or acquired resistance following therapy. Besides multi-drug resistance (MDR) caused by ABC efflux pumps, several targeted therapies have described the development of target-specific drug resistance. Thus, up to 90% of the cases of acquired resistance to tyrosine kinase inhibitors are due to over-expression of, or mutations in, the target kinase [2-4]. Acquired resistance can be studied by inducing resistance *in vitro* by growing cells in the presence of increasing concentrations of drug [1].

NAD is an essential cofactor in cell energy production and metabolism as well as the substrate for mono-

ADP-ribosyltransferases [5], poly-(ADP-ribose) polymerases (PARPs) [6] and sirtuins [7], all of these converting NAD to nicotinamide. PARPs are involved in DNA repair whereas sirtuins can increase cancer cell survival. To survive under stress and supply metabolites for cell growth malignant cells depend heavily on aerobic glycolysis for generation of ATP [8]. Glycolysis requires relatively more NAD to generate ATP compared to the oxidative phosphorylation normally occurring in non-malignant tissues. Also, cancer cells may display increased expression or activity of PARPs [9-11] and sirtuins [7] for increased DNA repair and cell survival. The first, rate-limiting step in the resynthesis pathway of NAD from nicotinamide is catalyzed by nicotinamide phosphoribosyltransferase (NAMPT) [12]. Nicotinamide is converted to nicotinamide mononucleotide (NMN) using 5-phosphoribosyl-1-pyrophosphate

* Correspondence: olesen.uh@gmail.com

¹Experimental Pathology Unit, Rigshospitalet, Copenhagen, Denmark
Full list of author information is available at the end of the article

and ATP as substrates. NMN is then converted to NAD by NMN adenylyltransferase (NMNAT) [13]. The crystal structure of NAMPT has been resolved and it has been identified as a dimer belonging to the family of type II phosphoribosyltransferases [14-16] - each monomer containing two domains. The dimer contains two binding sites for nicotinamide located in the vicinity of the dimer interface and residues of both monomers may be part of the binding site. Inhibition of NAMPT leads to depletion of NAD [17], secondarily leading to reduction of ATP and later, cell death. Also, it leads to substrate depletion of PARPs and sirtuins and furthermore, both PARPs and sirtuins are inhibited by nicotinamide [18-20]. Tumour cells are more sensitive to NAMPT inhibition and NAD depletion due to increased ATP and NAD consumption [17]. NAMPT inhibition shows high efficacy in haematological malignancies in preclinical studies [21]. APO866 is a specific, competitive, potent inhibitor of NAMPT that displays cytotoxicity in a broad panel of cell lines (Figure 1) [17,22]. APO866 has completed a phase I trial in oncology [22] and is currently undergoing several phase II trials for advanced melanoma and cutaneous T-cell lymphoma as well as a phase I/II trial for refractory and relapsed B-chronic lymphocytic leukaemia.

CHS-828 (Figure 1), a pyridyl cyanoguanidine, is a small molecule inhibitor displaying cytotoxicity in a broad panel

of cell lines [23]. We previously identified CHS-828 as an inhibitor of NAD synthesis [24]. We found CHS-828 to function similarly to APO866 in a number of assays although the two compounds are chemically distinct. Therefore, we suggested CHS-828 as an inhibitor of NAMPT. Furthermore, we compared a cell line, NYH/CHS, with acquired, specific resistance towards CHS-828 with its wild type counterpart without identifying the molecular basis for the resistance observed towards CHS-828 and APO866. CHS-828 has completed several phase I trials in oncology [25,26] and a prodrug EB1627/GMX1777 is currently also in phase I trials [27].

Here, we present a novel, potent analogue of CHS-828, namely TP201565 (Figure 1). This compound was found as part of a screen for NAMPT inhibitors and has shown activity in xenograft models. We developed a number of cell lines resistant to APO866 and TP201565. In most (4/5) of these we find mutations in the NAMPT gene that confer resistance when transfected into sensitive cells. The resistant cell lines show tumourigenicity in xenograft mouse models and *in vivo* resistance. Furthermore, through computer modelling and *in vitro* biochemistry we find that APO866, CHS-828 and TP201565 share a binding site in the active site of NAMPT, thus conclusively identifying CHS-828 and TP201565 as competitive inhibitors of NAMPT.

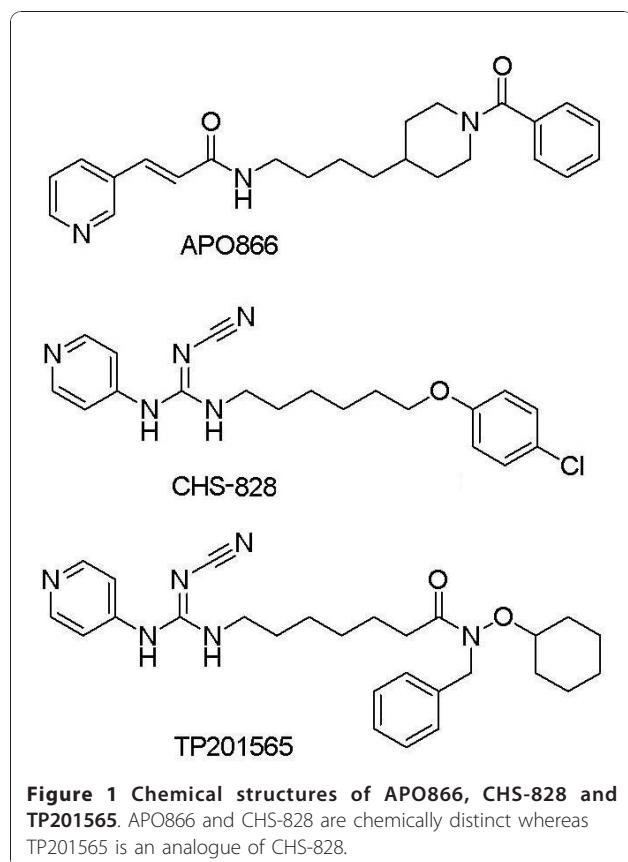


Figure 1 Chemical structures of APO866, CHS-828 and TP201565. APO866 and CHS-828 are chemically distinct whereas TP201565 is an analogue of CHS-828.

Methods

Drugs and chemicals

CHS-828 was synthesized as previously described [28]. APO866 and TP201565 and its linkable analogue were obtained from TopoTarget A/S (Copenhagen, Denmark). All other chemicals used in this paper were obtained from Sigma (St. Louis, MO, USA).

Cell lines

NYH and NYH/CHS have been described previously [24]. HCT-116, PC-3 and HEK293T were obtained from ATCC. To obtain resistance towards NAMPT inhibitors, the cell lines were cultured in a medium containing the drug in gradually increasing doses. This process was continued over an extended period of time until a level of stable, high-grade resistance was reached.

Clonogenic assay

Cells were incubated with the drug at different concentrations, in triplicates with a 3-week incubation period. Afterwards, the colonies were counted as described previously [29]. The colonies were counted in triplicate.

Western blotting

Western blotting of NAMPT and GAPDH was performed as described before [24]. The anti-rabbit PBEF antibody (Bethyl Laboratories, Montgomery, TX, USA) was used for

the detection of NAMPT at a dilution of 1:500. Densitometry measurements were done using Quantity One software (Bio-Rad Laboratories, Hercules, CA, USA) and results were averaged from 2-3 separate blots.

Cloning of NAMPT

Total RNA was purified from NYH/CHS, NYH/APO866, HCT-116/APO866, HCT-116/TP201565 and PC-3/TP201565 using Trizol reagent (Invitrogen, Carlsbad, CA, USA). mRNA was converted to cDNA using TaqMan[®] Reverse Transcription Kit (Applied Biosystem, Foster City, CA, USA) which was used as template for full length PCR of NAMPT by RED Extract-N-Amp[™]PCR ReadyMix[™] (Sigma). PCR products of all cell lines were sequenced twice and compared to NAMPT (GenBank accession # U02020). NAMPT flanked by BamHI/XhoI restriction sites was cloned into the TOPO TA cloning vector pCR4-TOPO[®] (Invitrogen) and clones containing identified mutations were selected. The mutated sequences were enzymatically cut with ScaI and BsaI (New England Biolabs, Ipswich, MA, USA) and ligated into wild type NAMPT in a pET28a(+) expression plasmid backbone (a kind gift from Dr. Shin-Ichiro Imai from Washington University School of Medicine, St. Louis USA) using T4 DNA ligase (New England Biolabs). Rosetta[™]2 (DE3) competent cells (Novagen, Merck, Darmstadt, Germany) were transformed with the resulting vectors. Sequencing was used to confirm correct insertion of the mutations.

Transfection

NAMPTwt, NAMPTH191R, NAMPTD93del and NAMPTQ388R/K342R with BamHI/XhoI restriction sites were ligated into the mammalian expression vector TRex pcDNA-4/TO/Myc.His (Invitrogen). 100 μ l sterile water containing 20 μ g/ml vector was mixed with 4 μ l FuGENE HD transfection agent (Roche, Basel, Switzerland). After 20 min the mix was added to 2×10^5 HEK-293T cells plated in 2 ml DMEM medium (Invitrogen) in 6-well plates 24 hours before transfection. After 24 hours, the cells were replated for experiments or for establishment of stable cell lines (HEK/CTR and HEK/NAMPTwt) in medium containing 300 ng/ml zeocin (Invitrogen). Cell proliferation was examined using Cell-TiterGlo (Promega, Madison, WI, USA) as described previously [24] using 72 hours incubation.

Recombinant protein expression and determination of enzymatic activity of NAMPT

The proteins were expressed as previously described [12] using 5 hour induction at 37°C with 1.5 mM IPTG (Sigma) and lysis was done in 20 mM Tris-HCl pH 8.0, 300 mM NaCl, 0.1% Triton X-100 and protease inhibitor cocktail (Sigma) using sonication. His-Tagged NAMPT proteins were purified using nickel-

nitriilotriacetic acid resin (Qiagen, Hilden, Germany) and imidazole elution. Purity and yield was evaluated by SDS gel electrophoresis and A280 measurements. Purified His-Tagged NAMPT proteins (WT, H191R and D93del) were incubated in reaction buffer (50 mM Tris-HCl [pH 8.5], 100 mM NaCl, 0.25 mM nicotinamide, 10 mM MgSO₄, 1.5% ethanol, 0.5 mM phosphoribosyl pyrophosphate, 2.0 mM ATP) for 55 min at 37°C. The reaction was then coupled with NMNAT1 (cloned and expressed similarly to NAMPT - as above) and alcohol dehydrogenase (Sigma) 10 μ g/ml each and incubated for further 5 minutes at 37°C. NMN production by NAMPT was evaluated indirectly by measuring the fluorescence (340 nm excitation, 465 nm emission). Fluorescence created by NADH was converted to NMN amounts using a standard curve with known amounts of NMN. k_{cat} values were averaged from calculations of k_{cat} for 500, 1000 and 2500 ng NAMPT enzyme. IC₅₀ values for APO866 were obtained using the same assay setup as described above. For each enzymatic reaction 2000 ng purified NAMPT was used. The concentrations of inhibitor used to generate the inhibition curves were in the range from 0 nM - 0.5 μ M for the wild type NAMPT and 0 nM - 100 μ M for H191R.

Xenograft studies

All mouse experiments were performed in accordance with national and EU legislation, and a permit to perform the specific type of experiments was obtained from the Experimental Animal Inspectorate, Danish Ministry of Justice. 10^7 cells of the paternal or resistant cell line of each of the four cell lines were injected s.c. in a mixture of PBS/matrigel in the right and left flank of female NMRI athymic mice (Taconic, Ry; Denmark), respectively. The mice were observed daily and tumour volumes were calculated from caliper measurements to estimate the radius and using the formula for a sphere. Tumours were allowed to reach a maximum volume of 1000 mm³, where after the mouse was sacrificed. Tissue samples of the xenograft tumours were excised and placed in phosphate buffered formaldehyde (4%, pH 7) for at least a week, and were then embedded in paraffin for sectioning and H&E staining. In xenograft studies with either the wild type or APO866-resistant variant of the HCT-116 tumour cell line mice were treated with a standard two daily i.p. injections of 15-20 mg/kg APO866 in PBS with 1% hydroxypropyl- β -cyclodextrin/12% propylene glycol on day 0-13, starting when tumour volumes were around 100 mm³, as previously described [30].

Statistical analyses

Statistical analysis and the graphical presentation were performed using the software GraphPad Prism[®] v. 4.0 (GraphPad Software, Inc., San Diego, CA, USA). Tumour doubling

times were calculated using the line-fitting function for exponential growth in Graph Pad Prism, and were compared using Student's *t*-test. The outcome of xenografts in the treatment experiment was quantified as the number of days until the tumour of each individual mouse reached a size of 800 mm³, expressed as survival days. The survival in each treatment group was compared using log-rank analysis. The level of significance was set to a *p* value of 0.05.

Measurement of NAMPT activity in lysates

Activity of NAMPT was examined in PC-3 and PC-3/TP201565 as well as HEK293T cells and HEK/NAMPTwt. NAMPT activity was measured by the conversion of ¹⁴C-labelled nicotinamide to ¹⁴C-NMN using a method previously described [31,32] using 50 µg total protein from cell lysate. Briefly, lysate was incubated with ATP, 5-phosphoribosyl-1-pyrophosphate and ¹⁴C-nicotinamide in reaction buffer. The mix was transferred to glass fibre filter and washed in acetone to remove excess ¹⁴C-nicotinamide. Counts per minute (cpm) determined by a liquid scintillation counter.

Computer modelling of NAMPT

The molecular modelling was based on the published X-ray structure of NAMPT in complex with the inhibitor APO866 (2.10 Å resolution, PDB id: 2GVJ) [14]. The structure was imported into Maestro v. 9.0 release 111 and prepared for docking in the following way using the protein preparation wizard (Schrodinger Suite 2009, Schrodinger LLC, Oregon, USA). First, bond orders were assigned and hydrogen atoms were added. The four selenocysteines (Cys368 and Cys372 in both monomers) which had been incorporated to facilitate X-ray crystallographic analysis were mutated back to cysteine. Water molecules further than 5Å away from the binding site were removed, and finally the hydrogen bonding network was optimized. The Glide docking module was used for the docking of the ligands. Initially, the receptor grid was generated (no vdW scaling of the receptor atoms was applied). The enclosing box for the docking of TP201565 and CHS-828 was based on the position of APO866 in the X-ray crystal structure. The hydroxyl groups in the receptor were allowed to rotate. For studies on the H191R mutation, APO866 was removed in the 2GVJ X-ray structure which had already been prepared for molecular modelling. The Prime module allowed for introduction of the mutation, which was followed by sampling of Arg191 dihedral angles to obtain the most favorable orientation of the new side chain.

Precipitation assays with sepharose linked analogue of TP201565

10 mg of the linkable analogue of TP201565 were coupled to NHS activated sepharose (GE-Healthcare)

following the manufacturer's instructions using Na₂CO₃-containing reaction buffer and ethanolamine for blocking. HCT-116 cells were lysed in MMENGM buffer (25 mM MOPS pH[7,4], 2 mM EDTA, 2 mM sodium orthovanadate, 1 mM NaN₃, 10% glycerol and 0,1% NP-40) with (1:200) protease inhibitor cocktail (Sigma) by sonication. Binding of NAMPT was conducted by mixing 30 µg of cellular protein with 20 µl coupled TP202648 beads and incubation for 60 min at 5°C with end over end rotation. After incubation, the beads were spun down and the supernatant removed followed by 3× washes with PBS (Invitrogen). Samples were boiled for 5 min at 95°C and run on a SDS gel followed by evaluation of the precipitated NAMPT protein by western blotting. For competitive studies, lysates were pre-incubated with APO866 (250 µM-2.5 nM) or TP201565 (250 µM-2.5 nM) for 15 min at 5°C before adding inhibitor linked beads and incubating for 60 min at 5°C.

Results

Development of resistant cell lines and identification of NAMPT mutations

We previously used the cell line NYH/CHS with resistance towards CHS-828 to determine the mechanism of action of CHS-828 as depletion of NAD, likely through inhibition of NAMPT [24]. From sensitive cell lines we developed a number of resistant cell lines to the NAMPT inhibitor APO866: NYH/APO866 and HCT-116/APO866, and to the CHS-828 analogue TP201565 (figure 1): HCT-116/TP201565 and PC-3/TP201565. Furthermore, we attempted to induce resistance in several other combinations of compounds and cell lines including NYH and HepG2 (hepatoma) cells with TP201565 and RPMI-8226 (myeloma) cells with APO866. No high-grade, stable resistance developed during 4 months of culture with drug (data not shown). Further, we have previously suggested that the resistance of NYH/CHS could be caused by a mutation in NAMPT [24]. Thus, we sequenced the NAMPT gene in the resistant cell lines and their parental lines. We found acquired mutations in NYH/CHS, NYH/APO866, HCT-116/APO866 and HCT-116/TP201565 but not in PC-3/TP201565 (Table 1). We also found two mutations that were present in both the HCT-116 parental cell line and in the resistant derivatives. One, 903G->A, was silent (not shown) and previously identified as a SNP (rs11553095, dbSNP build 130) whereas the other, 1025A->G, resulted in a change in protein sequence of K342R. Interestingly, the mutations H191R and D93del occurred in both resistant cell lines of HCT-116 and NYH, respectively. We correlated the positions of the mutations in the protein sequence with their positions in a NAMPT crystal structure published previously [14]. We

Table 1 Acquired and occurring mutations identified in NAMPT inhibitor resistant cell lines and parental line HCT-116

	Mutation	Change in protein sequence	Position
<i>HCT-116</i>	1025A>G	K342R	b-sheet 11
<i>HCT-116/TP201565</i>	1025A>G	K342R	b-sheet 11
	1163A>G	Q388R	Dimer interface
	572A>G	H191R	APO866 binding site
<i>HCT-116/APO866</i>	1025A>G	K342R	b-sheet 11
	572A>G	H191R	APO866 binding site
<i>NYH/CHS</i>	278-280del	D93del	Dimer interface
<i>NYH/APO866</i>	278-280del	D93del	Dimer interface

The structural position of the mutations has been determined from a published crystal structure of NAMPT [14]. All mutations are located in one allele only. In HCT-116/TP201565 the mutations K342R and Q388R are located on the same allele.

found that the acquired mutations were either located in the binding site of APO866 and nicotinamide or at the dimer interface of NAMPT whereas K342R was located distantly from either. All the mutations occurred in only one allele. In HCT-116/APO866 and HCT-116/TP201565, H191R and K342R were located on separate alleles whereas in the latter cell line, Q388R and K342R were located together on the same allele.

We studied the LD₅₀ values of the sensitive and resistant cell lines in a clonogenic survival assay. The parental cell lines were all sensitive with LD₅₀ values of 1.5-11 nM for APO866 (Table 2). TP201565 was a more potent compound in this clonogenic assay compared to APO866 and CHS-828 (10-100 fold). Generally, the cell lines displayed a high degree of cross-resistance (Table 3). NYH/CHS and NYH/APO866 displayed similar resistance to the compounds (data not shown). Further, we found that the resistance observed for each cell line was retained after at least 15 weeks of culture without drug (data not shown). Interestingly, we observed LD50 values of 5-10 μM for CHS-828 in HCT-116/APO866, HCT-116/TP201565 and PC-3/TP201565 (Table 3) that

Table 2 Sensitivity of cell lines as measured in a clonogenic survival assay

	APO866	CHS-828	TP201565
<i>NYH</i>	1.5 ± 0.5	1.7 ± 1.2	0.02 ± 0.01
<i>HCT-116</i>	10.9 ± 6.1	12.2 ± 7.4	0.4 ± 0.3
<i>PC-3</i>	3.8 ± 3.1	8.8 ± 8.4	0.3 ± 0.3

LD₅₀ values (measured in nM) of the parental cell lines NYH, HCT-116 and PC-3 when treated with APO866, CHS-828 or TP201565. LD₅₀ values are represented with standard deviations.

Table 3 Fold increase of LD₅₀ in resistant cell lines for treatment with NAMPT inhibitors

	APO866	CHS-828	TP201565
<i>NYH/CHS</i>	18.3	168	33.5
<i>HCT-116/APO866</i>	946	414	1,723
<i>HCT-116/TP201565</i>	2,364	774	10,103
<i>PC-3/TP201565</i>	8,324	624	19,675

The level of resistance of the resistant cell lines, NYH/CHS, HCT-116/APO866, HCT-116/TP201565 and PC-3/TP201565 is shown for treatment APO866, CHS-828 and TP201565. Values represent the ratio of LD₅₀ of the resistant cell line: LD₅₀ of parental cell line as measured by clonogenic survival assay.

were lower than for APO866. Otherwise, this compound shows similar or higher potency.

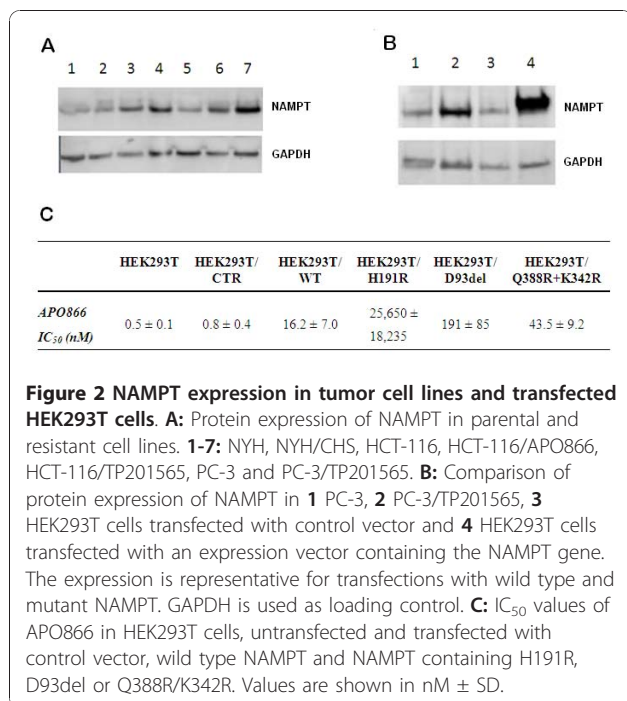
We also studied the protein expression of NAMPT in the parental and resistant cell lines. We found a slight up-regulation around 20% of NAMPT in HCT-116/APO866 compared to HCT-116 (figure 2A). Further, we observed higher levels of NAMPT in PC-3 cells compared to other parental cell lines and a marked up-regulation in PC-3/TP201565 compared to PC-3 (around 80%).

Over-expression of NAMPT variants induce APO866 resistance in sensitive HEK293T cells

To examine whether the acquired mutations are primarily responsible for the resistance observed in the resistant cell lines we over-expressed wild type NAMPT (NAMPTwt) and NAMPT containing either the H191R, D93del or Q388R/K342R mutations in HEK293T, a sensitive cell line, and measured the resistance induced by the NAMPT over-expression. We created a robust over-expression of NAMPT (Figure 2B) and we found that NAMPTwt induced an increase of resistance towards APO866 by 20 fold ($p < 0.005$). A further relative increase in resistance was induced by all mutant NAMPTs with the order of induced resistance being H191R>D93del>Q388R/K342R (Figure 2C). Over-expression of NAMPT containing the Q388R/K342R, H191R or D93del mutation yielded significantly greater resistance compared with wild type NAMPT over-expression ($p = 0.007$, $p = 0.01$ and $p = 0.002$, respectively). The H191R and D93del mutations resulted in an induced resistance in HEK293T cells towards APO866 similar to or above that of the corresponding resistant cell line. No significant difference was observed between untransfected HEK293T cells and HEK293T/CTR (empty vector).

APO866 binding and enzyme activity is affected by NAMPT mutations in vitro

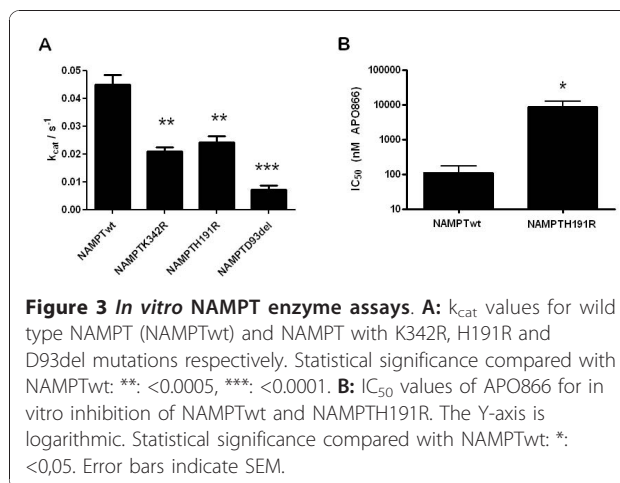
To study further the H191R and D93del mutations we purified recombinant NAMPTwt and NAMPT with either mutation. Using a colorimetric, coupled assay we



determined the k_{cat} for NAMPTwt (measured per active site), NAMPTH191R and NAMPTD93del. We found that the activities of NAMPTH191R and NAMPTK342R were reduced by ~50% by the mutations when compared to wild type whereas NAMPTD93del displayed 20% of the wild type activity (Figure 3A). We examined the inhibitory effect of APO866 in NAMPTwt and NAMPTH191R. The IC_{50} value for inhibition of NAMPTwt was 110 nM while we found an IC_{50} value for NAMPTH191R of 8585 nM (Figure 3B). Further, we found that APO866, CHS-828 and TP201565 did not inhibit NMNAT (data not shown).

NAMPT mutations do not inhibit *in vivo* xenograft growth and maintain resistance towards APO866

We studied whether the resistance induced *in vitro* would have an effect on tumour viability *in vivo*. Xenograft models were performed for parental cell lines NYH, HCT-116 and PC-3 as well as their resistant derivatives, and tumour formation rates were not affected in the resistant cell lines compared with parental cell lines. NYH tumours had a very fast doubling time in xenografts (3.7 days) followed by HCT-116 and PC-3 (8.9 and 10 days respectively). We observed no significant change in tumour doubling times for the resistant cell lines when compared to their wild type counterparts (Figure 4A). However, HCT-116/APO866 had a shorter tumour doubling time, 6.9 ± 1.8 days, compared with HCT-116 (8.9 ± 1.6 days) (Figure 4A). We examined tumour tissues histologically and found no obvious morphological differences between untreated sensitive



xenografts and their resistant counterparts (Figure 4B). Notably, although xenograft tumours from NAMPT inhibitor-resistant derivatives of HCT-116 were found to contain many necrotic areas, similar results were observed for parental HCT-116 cells. Finally, we treated HCT-116 and HCT-116/APO866 xenografts with 20 mg/kg APO866 daily for 14 days. We observed that APO866 increased the lifespan significantly in HCT-116 xenografts, increasing the median time for a tumour to reach the size of 800 mm^3 with 32% from 22 to 29 days, whereas treating the HCT-116/APO866 xenografts with APO866 resulted in no effect on the survival (Figure 4C), confirming *in vivo* resistance. There were no differences in body weight between the groups (data not shown).

PC-3/TP201565 and HEK293T/WT display high NAMPT activity compared to PC-3 and HEK293T

Since PC-3/TP201565 cells contained no mutations in NAMPT we examined whether the increased protein expression could be responsible for the NAMPT inhibitor resistance observed in the cell line. We studied the total *in vitro* activity of NAMPT from lysates of PC-3 and PC-3/TP201565 untreated and in response to inhibition with APO866. We found that the basal total activity of NAMPT was four fold higher in the resistant line compared to PC-3 in accordance with the higher expression of NAMPT (Figure 5A+B). However, the IC_{50} value for *in vitro* lysate treatment with APO866 was not significantly increased. Similarly, we observed a strong increase in total NAMPT activity when over-expressing NAMPT in HEK293T cells compared with wild type HEK293T cells (Figure 5C+D). Likewise, the IC_{50} was not increased by over-expression although the absolute NAMPT activity remained elevated at concentrations above the IC_{50} compared with untreated wild type HEK293T cells. Respective protein expression levels of NAMPT for the cell lines are shown in Figure 2B for

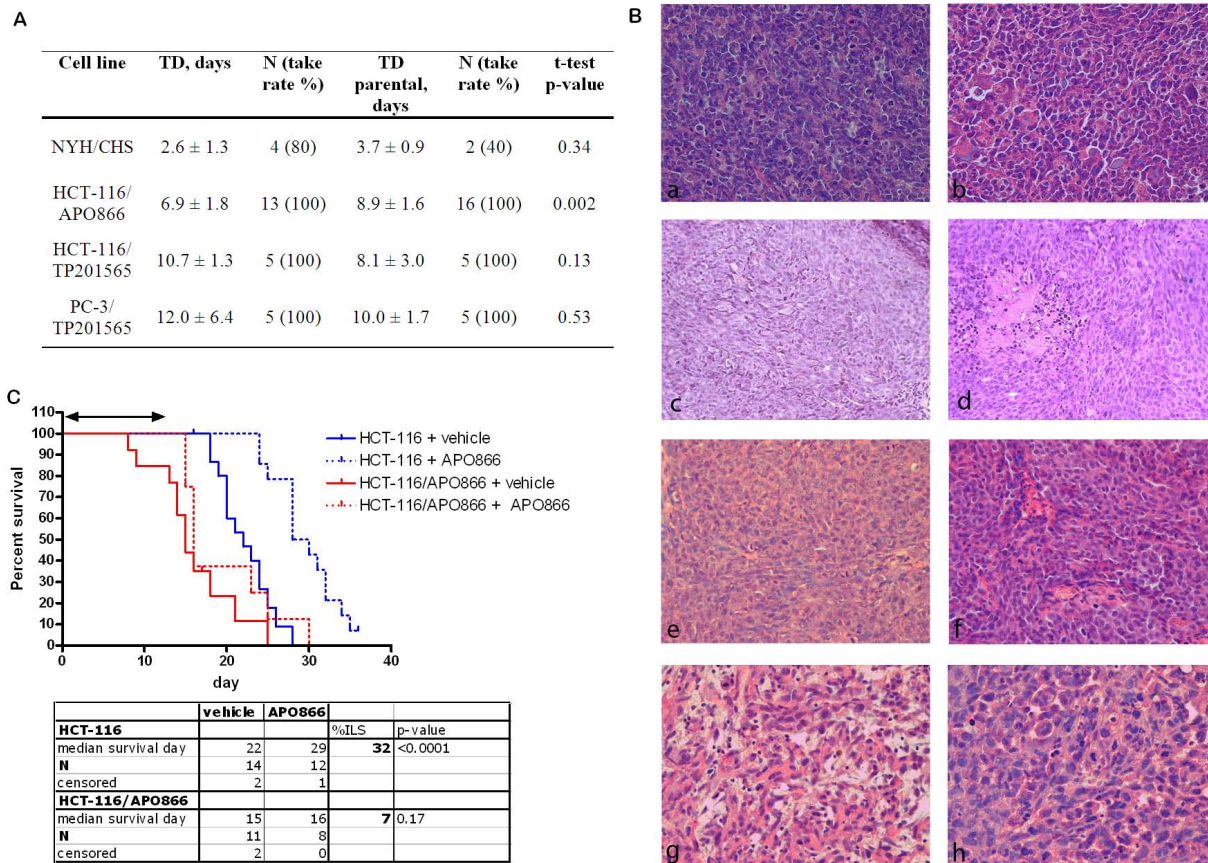


Figure 4 Subcutaneous xenograft experiments of parental and resistant cell lines. **A:** Tumour doubling times of NAMPT resistant cell lines compared to tumour doubling times (TD) of parental cell lines (shown with SD). p-values are determined by t-tests. **B:** H&E stained sections of xenograft tumours at 200x magnification. **a** NYH and **b** NYH/CHS xenografts contained cells of various sizes and showed many mitoses. Tumours of **c+e** HCT-116, **d** HCT-116/APO866 and **f** HCT-116/TP201565 contain many fine connective tissue strands between tumour cells. Also, necrotic foci and densely growing epithelial cells are observed. Morphology of **g** PC-3 and **h** PC-3/TP201565 varied between areas of rounded cells and more spindle-shaped tumour cells. Many pyknotic (apoptotic) cell nuclei were also found. **C:** Cumulative survival of mice with HCT-116 and HCT-116/APO866 xenografts showing the time used by each mouse to grow its tumour to a size of 800 mm³. The mice were treated with 15-20 mg/kg APO866 or vehicle twice daily for 14 days (0-13, double arrow), where day 0 indicates the day tumours had a size of approx. 100 mm³. The result of log-rank analysis comparing APO866 treated mice with control mice in the respective xenografts (HCT-116: blue, HCT-116/APO866: red) is shown on the graph.

comparison. Finally, we detected no up-regulation of MDR 1 and 2 pumps at the mRNA level and we found that PC-3/TP201565 cells show no cross-resistance towards the histone deacetylase inhibitor belinostat, the proteasome inhibitor MG-132, the DNA methyltransferase inhibitor 5-azacytidine, the topoisomerase I inhibitor camptothecin, the topoisomerase II inhibitors and ABC substrates doxorubicin and mitoxantrone, retinoic acid, the IKK inhibitor BMS-345541 or the protein kinase inhibitor staurosporine (data not shown). The drugs were chosen to cover a broad spectrum of mechanisms of action (including drugs sensitive to MDR mechanisms) to give an indication of the molecular basis of resistance in PC-3/TP201565 cells.

Docking studies confirm that APO866, CHS-828 and TP201565 share a common binding site

To study the relationship between the active site of NAMPT and APO866, CHS-828 and TP201565 as well as the H191R mutation, we used computer modelling based on a published x-ray structure of NAMPT [14]. The x-ray crystal structure contains several water molecules located either within or close to the binding site, hence we decided to carry out two separate docking runs; one in which water molecules within 5Å of the APO866 binding site were included in the calculations, and another without water molecules. Traditionally, docking has been carried out using water-free enzyme active-site,

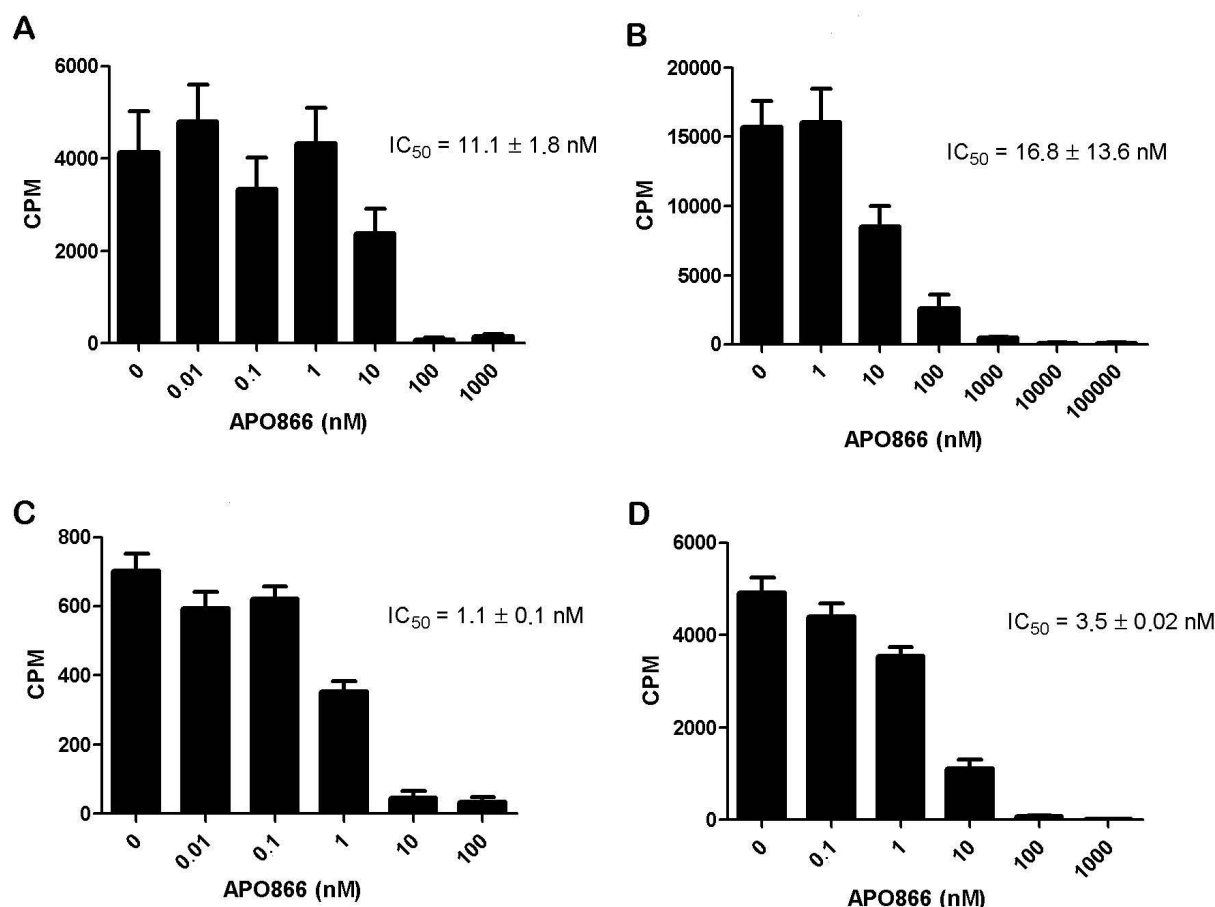


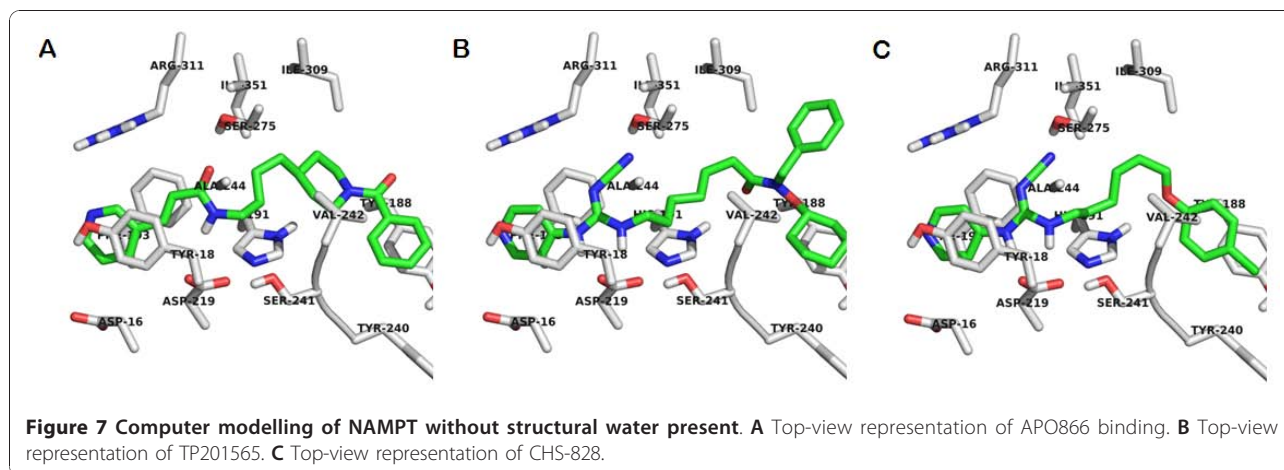
Figure 5 Total *in vitro* activity of NAMPT in cell lysates. NAMPT activity as measured by conversion of C14-labelled nicotinamide to NMN by 50 μ g of total protein from lysate untreated or with increasing concentrations of APO866. Absolute counts per minute (CPM) are presented. The IC₅₀ values have been averaged from at least 3 individual experiments. Cell lines measured are **A** PC-3, **B** PC-3/TP201565, **C** HEK293T and **D** HEK293T/WT. Basal activity of PC-3/TP201565 is 4.0 fold higher than PC-3 and basal activity of HEK293T/WT is 7.4 fold higher than HEK293T. Error bars indicate SEM; IC₅₀ values are represented with standard deviations.

but recently with increased accuracy of crystal structures and increased predictivity of computational chemistry the importance of water molecules has been realized [33]. If two ligands have very similar binding modes one might therefore investigate whether the crystal waters observed experimentally are also suitably positioned for the ligand where no crystal structure is available.

When the water molecules were present in the binding site, the highest-ranking docking conformation of APO866 was in good agreement with the reported X-ray structure (RMS 0.2003Å for non-hydrogen atoms, XP glidescore -8.92, Figure 6Aa). Surprisingly, also TP201565 was able to dock into the solvated active site with the pyridine moiety in the same π -stacking binding pocket between Tyr18' and Phe193. The double-headed TP201565 molecule could be stabilized by displaying either the phenyl or the cyclo-hexane in the opening of the narrow pore where APO866 has

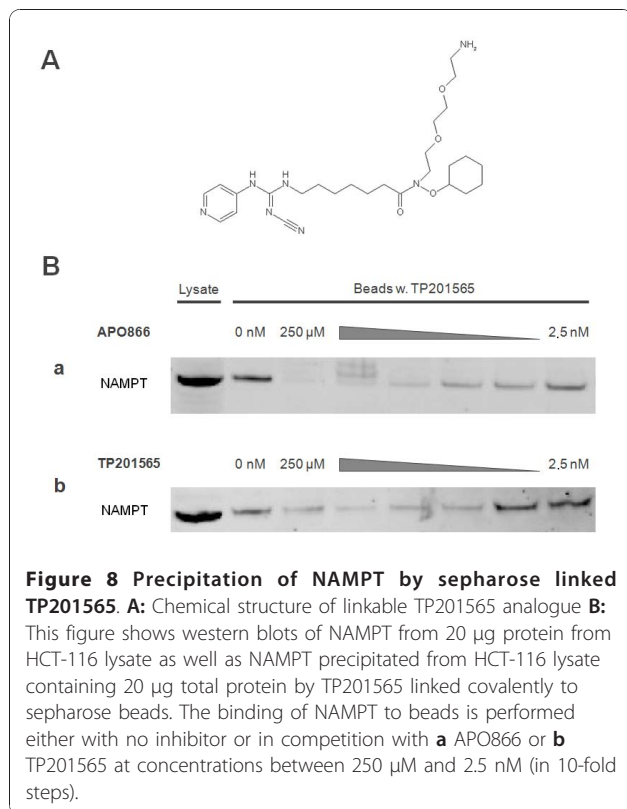
a phenyl group (figure 6Ab+c, XP glidescores -8.29 and -7.53, respectively). For CHS828 we did not obtain any docked conformations with good XP glidescore (lower than -6), indicating that the water-filled active site suitable for APO866 is not compatible with this compound.

Then, we turned our attention to the docking experiments carried out using the empty binding site without water molecules. The best scoring conformation of APO866 now had the phenyl group pointing towards the other pocket (XP glidescore -8.18, shown as top view, Figure 7A), but a conformation similar to the crystal structure was recovered as the fifth best conformation (XP glidescore -6.98, data not shown). TP201565 was able to achieve a good conformation (XP glidescore -7.78, top-view, Figure 7B). With the water molecules removed from the binding site CHS-828 was also able to obtain good interactions (XP glidescore -7.23, top-view, Figure 7C). All three compounds



APO866, it had not been definitively identified as an inhibitor of NAMPT. Furthermore, previous attempts to identify resistance mechanisms for CHS-828 have yielded no definite results [24,34] and resistance to APO866 has not been studied. In this study we introduce a novel potent analogue of CHS-828, TP201565, that shows >10 fold increased activity in sensitive cancer cell lines compared to APO866 and CHS-828. Using computer modelling, we find that CHS-828 and TP201565 most likely function as inhibitors of NAMPT by binding to the same site as APO866 and nicotinamide. This is further

supported by the fact that a sepharose linked TP201565 analogue can precipitate NAMPT from cellular lysate and that this binding can be abrogated by co-incubation with either APO866 or TP201565. We also show how mutations in NAMPT can block the binding of APO866, CHS-828 and TP201565. Together, these data conclusively determine the pyridyl cyanoguanidines, CHS-828 and TP201565, as competitive inhibitors of NAMPT. This is in agreement with very recently reported data, which through the use of metabolic screens, *in vitro* NAMPT assays and computer modelling reach a similar conclusion for CHS-828 [35].



We developed four novel cell lines with acquired drug resistance towards NAMPT inhibitors and including NYH/CHS cells we observed mutations in all except PC-3/TP201565. Interestingly, the H191R mutation was detected in two different resistant cell lines derived from HCT-116. Furthermore, the distinct deletion D93del was located in both NYH/CHS and NYH/APO866. This suggests that these mutations exist in a small subpopulation of the parental cell line and that in these cases resistance has been obtained from selection rather than *de novo* mutations. The fact that we never obtained resistance from treating NYH cells with TP201565 despite prolonged growth with drug may be due to the fact that cells containing the NAMPTD93del were not present in the original flask used for inducing resistance to TP201565. We found that the observed mutations in NAMPT could induce resistance similar to or above that seen in the original resistant cell line which strongly suggests the mutations as the primary mechanistic cause of resistance. Furthermore, we demonstrated that whereas H191R abrogated NAMPT inhibitor binding in computer modelling it only reduced the enzymatic NAMPT activity *in vitro* by 50% - similar to what we found for the K342R mutation. The change in k_{cat} may be due to altered substrate affinity resulting in an

increased K_M value. Also, we showed that the H191R mutation strongly increased the IC_{50} value for APO866 inhibition *in vitro*. The relatively high IC_{50} value of NAMPTwt obtained *in vitro* correlated with the previous results of others [35]. D93del on the other hand reduced k_{cat} *in vitro*. Although this may reduce overall NAMPT activity compared to wild type NYH cells, it is likely that enough NAD is produced to keep up with the cellular demands and thus no growth disadvantage is inferred. NYH/CHS cells have previously been shown to retain the D93del mutation and full resistance after 30 passages without drug present in the media [24] (unpublished results, Olesen, UH). Similarly we have observed retained resistance for the remaining resistant cell lines after culture for more than 15 weeks without drug. We speculate that the induction of resistance by D93del and Q388R may be due to interference with or abrogation of dimerisation of NAMPT as both are located in the dimer interface. The NAMPT monomer may potentially retain lower enzymatic activity while not binding the known NAMPT inhibitors. Alternatively, the resistant cell lines may have adapted through reduced NAD consumption or up-regulation of other sources of NAD such as *de novo* synthesis.

Besides the data on the H191R mutation in figure 6B, the observed difference in kinetics and inhibitory constants between the different mutants cannot reliably be predicted on the basis of the computer docking study performed in this study. To achieve such correlations molecular dynamics calculations of the entire enzyme (including specific solvation) for tens of nanoseconds would be necessary. However, this is far beyond what is feasible with our current computational resources.

Notably, CHS-828 seems to show an upper level in LD_{50} values around 5-10 μM which is significantly lower than what is seen for APO866. We speculate that this may be due to a secondary mechanism of cytotoxicity occurring at above 1 μM . This has also previously been suggested [36,37]. Although this currently unknown mechanism may be less important in cellular and molecular assays, it may well be of interest clinically for CHS-828, specifically since serum concentrations of the drug have been reported to reach 11 μM in clinical trials [26]. However, it is unlikely that it is related to the previously proposed action of CHS-828 as an inhibitor of the I- κ B kinase (IKK) complex [38]. CHS-828 showed no inhibition of IKK α , IKK β or TBK1 enzymes *in vitro* (unpublished results, Olesen, UH). APO866 displayed a surprisingly specific induction of cell death in tumour cells ranging five orders of magnitude from close to 1 nM to more than 30 μM *in vitro* and a similar specificity was seen for TP201565. Finally, the specific nature required of mutations in NAMPT to confer resistance combined with the lack of resistance induced even at

prolonged drug treatment in a number of combinations of cell lines and NAMPT inhibitors may indicate that resistance is not induced frequently unless a suitable mutation in NAMPT is already present in a population of tumour cells. However, we also found that the resistance is not easily reversible. Also, no significant increase in tumour doubling times occurred *in vivo* for the resistant cell lines. Rather, we find that HCT-116/APO866 xenografts displayed reduced tumour doubling times compared to HCT-116. As we did not observe a similar trend for PC-3/TP201565 it seems unlikely to be due to increased NAMPT levels. Rather, the induction of resistance may have resulted in the development of an unidentified growth advantage in HCT-116/APO866. Further, the *in vitro* resistance also conferred insensitivity to APO866 treatment in xenograft models shown for HCT-116/APO866.

The PC-3/TP201565 cell line is highly resistant towards NAMPT inhibitors while displaying up-regulation but no mutations of NAMPT. The up-regulation could be due to increased gene copy number. We found that the basal NAMPT activity in the resistant cell line and also in transfected NAMPT over-expressing HEK293T/WT was much higher than in wild type PC-3 and HEK293T cells. The fact that PC-3/TP201565 cells displayed higher total activity than HEK293T/WT despite the latter having higher expression of NAMPT agrees with previous findings that endogenous NAMPT has higher activity compared to recombinant NAMPT [39]. However, the IC_{50} was not significantly changed and although the absolute NAMPT activity remained high relative to wild type PC-3 and HEK293T at concentrations of APO866 up to 1-10 μM , it would not seem to fully explain the $>10 \mu M$ LD_{50} value observed for APO866 in PC-3/TP201565. Further, the resistance appeared to be specific to NAMPT inhibitors as cross-resistance to other chemotherapeutics was not observed and it was not related over-expression of MDR1 and 2. Also, CHS-828 has displayed only low to moderate sensitivity to mechanisms of MDR based on over-expression of Pgp170 and MRP, increased levels of glutathione and tubulin-associated MDR [23,40]. Thus, we believe that PC-3/TP201565 cells are likely to possess a further, yet unknown, specific mechanism of resistance. Still, the over-expression explains part of the resistance and, as also seen for HEK293T/WT cells, high levels of NAMPT may be sufficient to induce resistance which would be significant in a clinical setting. This is similar to the situation with thymidylate synthase where high expression levels predict resistance to 5-fluorouracil and a poor treatment outcome [41,42].

The crystal structure of NAMPT in complex with APO866 has previously led to suggestions on improving the potency of APO866 based on its interaction with

the NAMPT binding site [14,43]. So far, published APO866 analogues have shown less activity in cellular assays [44,45]. We found that the mutations in NAMPT identified in the resistant cell lines show a tendency to locate either in the binding site or dimer interface. These mutations may directly or indirectly adjust the shape of the binding site so that it is unsuitable for the relatively large NAMPT inhibitors while still accessible for nicotinamide and NMN. Also, others have recently published a single resistant cell line showing a point mutation, G217R, which is close to the active site of NAMPT and is responsible for high-grade resistance towards CHS-828 [14,35]. We hope that the identification of a new class of NAMPT inhibitors and the characterization of resistance inducing mutations in NAMPT may be useful in developing second-generation NAMPT inhibitors with higher potency and potentially less affected by acquired resistance.

Conclusions

We have identified the cyanoguanidines CHS-828 and TP201565 as direct inhibitors of NAMPT. Further, we found that NAMPT mutations located around the enzyme active site or the dimer interface could be expected to be the most frequent cause of acquired resistance towards inhibitors of NAMPT. The location of the mutations could be applied as guide for development of next-generation NAMPT inhibitors with reduced risk of development of resistance.

Abbreviations

(NAMPT): Nicotinamide phosphoribosyltransferase; (PARPs): poly-(ADP-ribose) polymerases; (NMN): nicotinamide mononucleotide; (NMNAT): NMN adenylyltransferase; (NAMPTwt): wild type His-tagged NAMPT;

Acknowledgements

The authors would like to thank Ida Kappel Buhl, Anne Jørgensen, Annette Nielsen and Anja Barnikol-Oettler for invaluable technical assistance and Deirdre McPaul Shiel for linguistic advice during manuscript preparation. AG is supported by a junior research grant from the Medical Faculty, University of Leipzig. JY is supported by the Japan Research Foundation for Clinical Pharmacology, Manpei Suzuki Diabetes Foundation, and Kanae Foundation for the Promotion of Medical Science. PF thanks The Danish Council for Independent Research | Technology and Production Sciences for financial support.

Author details

¹Experimental Pathology Unit, Rigshospitalet, Copenhagen, Denmark.
²TopoTarget A/S, Copenhagen, Denmark. ³Research Laboratory, Hospital for Children and Adolescents, University of Leipzig, Leipzig, Germany.
⁴Department of Developmental Biology, Washington University School of Medicine, St. Louis, MO, USA. ⁵Department of Chemistry, Technical University of Denmark, Lyngby, Denmark. ⁶Department of Medicinal Chemistry, Faculty of Pharmaceutical Sciences, University of Copenhagen, Copenhagen, Denmark.

Authors' contributions

UHO has been involved in the design and planning of the *in vitro* and *in vivo* experiments, carried out identification of mutations and cellular transfections and drafted the manuscript. JGP carried out recombinant NAMPT experiments and linker-assays. AG and WK designed and carried out

the *in vitro* lysate activity studies. JY and SI generated the expression plasmid for wild type human NAMPT protein and contributed to the construction of plasmids for mutant proteins. MKC and FB synthesized the involved compounds. PF performed the *in silico* studies. AT carried out *in vivo* studies. SJN and MS were involved in the planning of the study and revised the manuscript and PBJ likewise revised the manuscript critically. All authors have read and approved the manuscript.

Competing interests

TopoTarget A/S has funded parts of this study. UHO, JGP, MKC, AT, FB, PBJ, SJN and MS have recently been fully or partially employed by TopoTarget A/S and several authors also hold stock in TopoTarget A/S. TopoTarget A/S has licensed the rights to APO866 and owns the rights to TP201565 and has provided the drugs used for this study.

Received: 7 June 2010 Accepted: 12 December 2010

Published: 12 December 2010

References

1. Janne PA, Gray N, Settleman J: Factors underlying sensitivity of cancers to small-molecule kinase inhibitors. *Nat Rev Drug Discov* 2009, **8**(9):709-723.
2. Gorre ME, Mohammed M, Ellwood K, Hsu N, Paquette R, Rao PN, Sawyers CL: Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. *Science* 2001, **293**(5531):876-880.
3. Gorre ME, Sawyers CL: Molecular mechanisms of resistance to STI571 in chronic myeloid leukemia. *Curr Opin Hematol* 2002, **9**(4):303-307.
4. O'Hare T, Corbin AS, Druker BJ: Targeted CML therapy: controlling drug resistance, seeking cure. *Curr Opin Genet Dev* 2006, **16**(11):92-99.
5. Ziegler M: New functions of a long-known molecule. Emerging roles of NAD in cellular signaling. *Eur J Biochem* 2000, **267**(6):1550-1564.
6. Diefenbach J, Burkle A: Introduction to poly(ADP-ribose) metabolism. *Cell Mol Life Sci* 2005, **62**(7-8):721-730.
7. Saunders LR, Verdin E: Sirtuins: critical regulators at the crossroads between cancer and aging. *Oncogene* 2007, **26**(37):5489-5504.
8. Kroemer G, Pouyssegur J: Tumor cell metabolism: cancer's Achilles' heel. *Cancer Cell* 2008, **13**(6):472-482.
9. Hageman GJ, Stierum RH: Niacin, poly(ADP-ribose) polymerase-1 and genomic stability. *Mutat Res* 2001, **475**(1-2):45-56.
10. Lengauer C, Kinzler KW, Vogelstein B: Genetic instabilities in human cancers. *Nature* 1998, **396**(6712):643-649.
11. Nomura F, Yaguchi M, Togawa A, Miyazaki M, Isobe K, Miyake M, Noda M, Nakai T: Enhancement of poly-adenosine diphosphate-ribosylation in human hepatocellular carcinoma. *J Gastroenterol Hepatol* 2000, **15**(5):529-535.
12. Revollo JR, Grimm AA, Imai S: The NAD biosynthesis pathway mediated by nicotinamide phosphoribosyltransferase regulates Sir2 activity in mammalian cells. *J Biol Chem* 2004, **279**(49):50754-50763.
13. Revollo JR, Grimm AA, Imai S: The regulation of nicotinamide adenine dinucleotide biosynthesis by Nampt/PBEF/visfatin in mammals. *Curr Opin Gastroenterol* 2007, **23**(2):164-170.
14. Khan JA, Tao X, Tong L: Molecular basis for the inhibition of human NMPRTase, a novel target for anticancer agents. *Nat Struct Mol Biol* 2006, **13**(7):582-588.
15. Kim MK, Lee JH, Kim H, Park SJ, Kim SH, Kang GB, Lee YS, Kim JB, Kim KK, Suh SW, et al: Crystal structure of visfatin/pre-B cell colony-enhancing factor 1/nicotinamide phosphoribosyltransferase, free and in complex with the anti-cancer agent FK-866. *J Mol Biol* 2006, **362**(1):66-77.
16. Wang T, Zhang X, Bheda P, Revollo JR, Imai S, Wolberger C: Structure of Nampt/PBEF/visfatin, a mammalian NAD⁺ biosynthetic enzyme. *Nat Struct Mol Biol* 2006, **13**(7):661-662.
17. Hasmann M, Schemainda I: FK866, a highly specific noncompetitive inhibitor of nicotinamide phosphoribosyltransferase, represents a novel mechanism for induction of tumor cell apoptosis. *Cancer Res* 2003, **63**(21):7436-7442.
18. Clark JB, Ferris GM, Pinder S: Inhibition of nuclear NAD nucleosidase and poly ADP-ribose polymerase activity from rat liver by nicotinamide and 5'-methyl nicotinamide. *Biochim Biophys Acta* 1971, **238**(1):82-85.
19. Jackson MD, Schmidt MT, Oppenheimer NJ, Denu JM: Mechanism of nicotinamide inhibition and transglycosylation by Sir2 histone/protein deacetylases. *J Biol Chem* 2003, **278**(51):50985-50998.

20. Sauve AA, Schramm VL: **Sir2 regulation by nicotinamide results from switching between base exchange and deacetylation chemistry.** *Biochemistry* 2003, **42**(31):9249-9256.
21. Nahimana A, Attinger A, Aubry D, Greaney P, Ireson C, Thougard AV, Tjornelund J, Dawson KM, Dupuis M, Duchosal MA: **The NAD biosynthesis inhibitor APO866 has potent antitumor activity against hematologic malignancies.** *Blood* 2009, **113**(14):3276-3286.
22. Holen K, Saltz LB, Hollywood E, Burk K, Hanauske AR: **The pharmacokinetics, toxicities, and biologic effects of FK866, a nicotinamide adenine dinucleotide biosynthesis inhibitor.** *Invest New Drugs* 2008, **26**(1):45-51.
23. Hjarnaa PJ, Jonsson E, Latini S, Dhar S, Larsson R, Bramm E, Skov T, Binderup L: **CHS 828, a novel pyridyl cyanoguanidine with potent antitumor activity in vitro and in vivo.** *Cancer Res* 1999, **59**(22):5751-5757.
24. Olesen UH, Christensen MK, Bjorkling F, Jaattela M, Jensen PB, Sehested M, Nielsen SJ: **Anticancer agent CHS-828 inhibits cellular synthesis of NAD.** *Biochem Biophys Res Commun* 2008, **367**(4):799-804.
25. Hovstadius P, Larsson R, Jonsson E, Skov T, Kissmeyer AM, Krasilnikoff K, Bergh J, Karlsson MO, Lonnebo A, Ahlgren J: **A Phase I study of CHS 828 in patients with solid tumor malignancy.** *Clin Cancer Res* 2002, **8**(9):2843-2850.
26. Ravaud A, Cerny T, Terret C, Wanders J, Bui BN, Hess D, Droz JP, Fumoleau P, Twelves C: **Phase I study and pharmacokinetic of CHS-828, a guanidino-containing compound, administered orally as a single dose every 3 weeks in solid tumours: an ECSG/EORTC study.** *Eur J Cancer* 2005, **41**(5):702-707.
27. Pishvaian M, Marshall J, Hwang J, Malik S, He A, Deeken J, Kelso C, Cotaria I, Berger M: **A phase I trial of GMX1777, an inhibitor of nicotinamide phosphoribosyl transferase (NAMPT), given as a 24-hour infusion.** 2009 *ASCO Annual Meeting: 2009: ASCO* 2009.
28. Schou C, Ottosen ER, Petersen HJ, Bjorkling F, Latini S, Hjarnaa PV, Bramm E, Binderup L: **Novel cyanoguanidines with potent oral antitumor activity.** *Bioorg Med Chem Lett* 1997, **7**(24):3095-3100.
29. Jensen PB, Christensen IJ, Sehested M, Hansen HH, Vindelov L: **Differential cytotoxicity of 19 anticancer agents in wild type and etoposide resistant small cell lung cancer cell lines.** *Br J Cancer* 1993, **67**(2):311-320.
30. Olesen UH, Thougard AV, Jensen PB, Sehested M: **A preclinical study on the rescue of normal tissue by nicotinic acid in high dose treatment with APO866, a specific nicotinamide phosphoribosyltransferase inhibitor.** *Mol Cancer Ther* 2010, **9**(6):1609-1617.
31. Elliott GC, Ajioka J, Okada CY: **A rapid procedure for assaying nicotinamide phosphoribosyltransferase.** *Anal Biochem* 1980, **107**(1):199-205.
32. Garten A, Petzold S, Barnikol-Oettler A, Korner A, Thasler WE, Kratzsch J, Kiess W, Gebhardt R: **Nicotinamide phosphoribosyltransferase (NAMPT/PBEF/visfatin) is constitutively released from human hepatocytes.** *Biochem Biophys Res Commun* 2009, **391**(1):376-381.
33. Mattos C: **Protein-water interactions in a dynamic world.** *Trends Biochem Sci* 2002, **27**(4):203-208.
34. Gullbo J, Lovborg H, Dhar S, Lukinius A, Oberg F, Nilsson K, Bjorkling F, Binderup L, Nygren P, Larsson R: **Development and characterization of two human tumor sublines expressing high-grade resistance to the cyanoguanidine CHS 828.** *Anticancer Drugs* 2004, **15**(1):45-54.
35. Watson M, Roulston A, Belec L, Billot X, Marcellus R, Bedard D, Bernier C, Branchaud S, Chan H, Dairi K, et al: **The Small Molecule GMX1778 is a Potent Inhibitor of NAD+ Biosynthesis: Strategy for Enhanced Therapy in NAPRT1-Deficient Tumors.** *Mol Cell Biol* 2009, **29**(21):5872-5888.
36. Hassan SB, Jonsson E, Larsson R, Karlsson MO: **Model for time dependency of cytotoxic effect of CHS 828 in vitro suggests two different mechanisms of action.** *J Pharmacol Exp Ther* 2001, **299**(3):1140-1147.
37. Martinsson P, de la Torre M, Binderup L, Nygren P, Larsson R: **Cell death with atypical features induced by the novel antitumor drug CHS 828, in human U-937 GTB cells.** *Eur J Pharmacol* 2001, **417**(3):181-187.
38. Olsen LS, Hjarnaa PJ, Latini S, Holm PK, Larsson R, Bramm E, Binderup L, Madsen MW: **Anticancer agent CHS 828 suppresses nuclear factor-kappa B activity in cancer cells through downregulation of IKK activity.** *Int J Cancer* 2004, **111**(2):198-205.
39. Revollo JR, Korner A, Mills KF, Satoh A, Wang T, Garten A, Dasgupta B, Sasaki Y, Wolberger C, Townsend RR, et al: **Nampt/PBEF/Visfatin regulates insulin secretion in beta cells as a systemic NAD biosynthetic enzyme.** *Cell Metab* 2007, **6**(5):363-375.
40. Hovstadius P, Lindhagen E, Hassan S, Nilsson K, Jernberg-Wiklund H, Nygren P, Binderup L, Larsson R: **Cytotoxic effect in vivo and in vitro of CHS 828 on human myeloma cell lines.** *Anticancer Drugs* 2004, **15**(1):63-70.
41. Formentini A, Henne-Bruns D, Kornmann M: **Thymidylate synthase expression and prognosis of patients with gastrointestinal cancers receiving adjuvant chemotherapy: a review.** *Langenbecks Arch Surg* 2004, **389**(5):405-413.
42. Peters GJ, Smorenburg CH, Van Groeningen CJ: **Prospective clinical trials using a pharmacogenetic/pharmacogenomic approach.** *J Chemother* 2004, **16**(Suppl 4):25-30.
43. Khan JA, Forouhar F, Tao X, Tong L: **Nicotinamide adenine dinucleotide metabolism as an attractive target for drug discovery.** *Expert Opin Ther Targets* 2007, **11**(5):695-705.
44. Galli U, Ercolano E, Carraro L, Blasi Roman CR, Sorba G, Canonico PL, Genazzani AA, Tron GC, Billington RA: **Synthesis and biological evaluation of isosteric analogues of FK866, an inhibitor of NAD salvage.** *ChemMedChem* 2008, **3**(5):771-779.
45. Kang GB, Bae MH, Kim MK, Im I, Kim YC, Eom SH: **Crystal structure of Rattus norvegicus Visfatin/PBEF/Nampt in complex with an FK866-based inhibitor.** *Mol Cells* 2009, **27**(6):667-671.

Pre-publication history

The pre-publication history for this paper can be accessed here:
<http://www.biomedcentral.com/1471-2407/10/677/prepub>

doi:10.1186/1471-2407-10-677

Cite this article as: Olesen et al.: Target enzyme mutations are the molecular basis for resistance towards pharmacological inhibition of nicotinamide phosphoribosyltransferase. *BMC Cancer* 2010 **10**:677.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

