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Barts Health NHS Trust
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Study protocol: MyoFit46—the cardiac sub-study of the MRC National Survey of Health and Development

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Abstract

Background: The life course accumulation of overt and subclinical myocardial dysfunction contributes to older age mortality, frailty, disability and loss of independence. The Medical Research Council National Survey of Health and Development (NSHD) is the world’s longest running continued surveillance birth cohort providing a unique opportunity to understand life course determinants of myocardial dysfunction as part of MyoFit46—the cardiac sub-study of the NSHD.

Methods: We aim to recruit 550 NSHD participants of approximately 75 years+ to undertake high-density surface electrocardiographic imaging (ECGI) and stress perfusion cardiovascular magnetic resonance (CMR). Through comprehensive myocardial tissue characterization and 4-dimensional flow we hope to better understand the burden of clinical and subclinical cardiovascular disease. Supercomputers will be used to combine the multi-scale ECGI and CMR datasets per participant. Rarely available, prospectively collected whole-of-life data on exposures, traditional risk factors and multimorbidity will be studied to identify risk trajectories, critical change periods, mediators and cumulative impacts on the myocardium.

Discussion: By combining well curated, prospectively acquired longitudinal data of the NSHD with novel CMR–ECGI data and sharing these results and associated pipelines with the CMR community, MyoFit46 seeks to transform our understanding of how early, mid and later-life risk factor trajectories interact to determine the state of cardiovascular health in older age.

Trial registration: Prospectively registered on ClinicalTrials.gov with trial ID: 19/LO/1774 Multimorbidity Life-Course Approach to Myocardial Health- A Cardiac Sub-Study of the MRC National Survey of Health and Development (NSHD).

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Background
Unprecedented numbers of people are now surviving into older age [1, 2] which means that the global trends for cardiovascular disease (CVD) prevalence will continue to increase, further adding to the burden of disease [3]. Improving our understanding of the causes of CVD is crucial to developing lifestyle or pharmacological interventions that can either prevent or delay the onset of disease.

The progressive structural, functional and hemodynamic changes affecting the myocardium as a result of ageing, lead to mortality, morbidity, disability and loss of independence in later life [4, 5]. Risk factors for cardiometabolic disease impact the heart from early childhood [6, 7] and with advancing age, the health of the myocardium is progressively compromised by overt stepwise and subclinical cumulative injury [5]. It is currently unclear how the age-dependent influences of multimorbidity, or the effect of early-life exposures, influence the various pathophysiological processes that lead to cardiac disease [8]. Prospectively collected data across the whole life course are required to address these research questions.

The Medical Research Council (MRC) National Survey of Health and Development (NSHD) is the longest running continuous study of human development in the world, which started as an English, Scottish and Welsh representative sample of 5,362 participants, born in one week in 1946 [9–12]. Repeated data collection waves since birth have provided detailed information on early-life exposures, traditional cardiometabolic risk factors and emergent multimorbidity [9, 11]. With members now aged ≥ 75, it is an opportune time to perform advanced cardiac phenotyping to understand which life course trajectories are conducive to the preservation of cardiovascular longevity. We propose to combine high spatial and temporal resolution electrocardiographic imaging (ECGI) and cardiovascular magnetic resonance (CMR) techniques to study the structure, function, electrophysiology (EP) and hemodynamics of the whole heart and correlate this information with the rich life course data that the NSHD cohort provides.

We describe here the research protocol of ‘MyoFit46’, a prospective longitudinal sub-study of 550 NSHD study members which is set to run for a total of 5 years. We summarize the study’s organization, funding, design, participant inclusion criteria, data sharing policy and analysis strategy.

Methods
Study organization
This prospective longitudinal NSHD sub-study will take place at the University College London (UCL) Bloomsbury Centre for Clinical Phenotyping in central London. MyoFit46 is funded by the British Heart Foundation (BHF) special project grant (to G.C. SP/20/2/34841) and supported by the MRC (MCUU00019/1).

Separate ethical approval for the main NSHD study has been provided by Research Ethics Committees (REC) in England and Scotland as outlined in previous papers [9, 10, 13]. Ethical approval for this study was granted by the London Queen Square REC (REC: 19/LO/1774. London REC—Queen Square, IRAS: 254776). All participants will provide written informed consent to participate in the study and for their study data to be stored electronically in accordance with the Data Protection Act (2018) as laid out by the legal requirements of the General Data Protection Regulation (GDPR).

Participants
Entry criteria to the study are based on maximizing the life course data available for analysis. We will recruit 550 NSHD participants, some of which have been recruited to other recent sub-studies [12] and who have not previously withdrawn, died, or remained untraced from the main study by age 70. Participants will have a rich life course data set available having participated in the majority of data collection sweeps (Table 1). The NSHD is a representative sample of 5362 males and females born in the UK in March 1946 originally intended to answer questions on fertility rates and obstetric services [14]. At the 2014–2016 NSHD data sweep we obtained data on surviving study members that lived in Britain and with whom we still had contact [14], which has provided us with 2502 participants potentially available to invite for examination. The first 550 study members fulfilling these criteria and providing written consent will be included. Excluded individuals will be those with contraindications to contrast perfusion CMR, including but not limited to those with permanent implantable cardiac electronic devices, claustrophobia, renal failure, severe asthma or known trifascicular or higher degree conduction block on their resting surface 12-lead ECG.

Keywords: Subclinical myocardial dysfunction, Cardiovascular health, Life course risk factors, Risk trajectories, Cardiovascular magnetic resonance, Electrocardiographic imaging, Myocardial tissue characterization, Perfusion, 4-dimensional flow
<table>
<thead>
<tr>
<th>Data collections</th>
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<th>Cognitive function</th>
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<tr>
<td></td>
<td>Cognitive function</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
</tr>
</tbody>
</table>

Participants recruited to MyoFit46 may not necessarily have participated in all the data sweeps outlined in this table, but they will have participated in the majority of sweeps. ECG, Electrocardiogram; HR, heart rate; MRI, magnetic resonance imaging; NSHD, National Survey of Health and Development; y, years.
Study outline
All participants meeting the inclusion criteria will be invited to take part through an invitation letter and following verbal agreement, a suitable date/time for the CMR scan will be booked and transport arranged to and from the study center. In light of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) global pandemic specific measures will be implemented to minimize the risk of participant/investigator infection or transmission throughout the study period in accordance with changing government guidelines. A flowchart outlining the study protocol is shown in Fig. 1.

Assessment and management of risk
A duty-of-care protocol based on the NSHD protocol used previously, in accordance with the MRC/Wellcome Trust guidelines [15] will be implemented for the purposes of feeding back health-related findings from the study. All baseline measurements will be routinely reported to the general practitioner with a copy also being sent to the participant.

All CMR images will be pre-reported by a cardiologist clinical research fellow and finalized by a consultant cardiologist. Any extra-cardiac findings considered to be clinically significant will be secondarily reported by a consultant radiologist at University College London National Health Service (NHS) Foundation Trust. The study follows guidelines based on the UK Biobank imaging study [16] and CMR information is only reported back to the GP in the case that an abnormality is considered medically actionable. A non-exhaustive list of potential CMR findings considered reportable are summarized in Table 2.

All serious unexpected study-related adverse events will be reported immediately according to standard operating procedures for research studies carried out at UCL.

Data handling and management
All data will be handled in accordance with the UK DPA 2018 as laid out by the UK GDPR. All clinical data will be pseudonymized by tokenization and will not bear the participant’s name or other directly identifiable data. The participant’s study identification number only, will be used for identification. All participant information including pre-screening questionnaires, relevant medical history, anthropometric measurements, baseline ECG and CMR data points will be imported into the secure research electronic data capture project (REDCap V.7.3.2) controlled by the principal investigator and accessible to named research assistants.

Study protocol description
Blood and urine specimen collection and storage
Blood samples will be collected and stored for future biomarker analysis. Two 4.0 ml ethylenediaminetetraacetic acid and two clotted 3.5 ml serum-separating tube whole blood samples will be collected. All tubes will be spun at 1300 g for 20 min at room temperature to generate supernatant (plasma), buffy coat and red blood cells which will be individually aliquoted into 1 ml, polypropylene, cryovials and stored at −80 °C, for later analysis. One 10.5 ml Paxgene DNA tube and one 9.4 ml Paxgene RNA tube will be collected and stored upright for 24 h at −20 °C, and then transferred to −80 °C, for future analysis. Point-of-care testing will be carried out for creatinine using the StatSensor Xpress Creatinin monitor (Nova Biomedical, USA) and hemoglobin using HemoCue Hb 801 monitor (HemoCue AB, Sweden) that also permits derivation of hematocrit. A random urine sample will be collected by the participant in a polypropylene universal container and aliquoted into slender conical tubes for storage at −80 °C.

Baseline measurements
Height and weight will be measured according to standardized protocols previously described [12]. Bioimpedance body composition measurements (Tanita Cooperation, Japan) will also be recorded. Lying, sitting and 3-min standing blood pressure (BP) will be measured (OMRON MIT ELITE PLUS; OMRON Healthcare UK Ltd. Milton Keynes) on the left arm over 3 acquisitions. A resting 12-lead surface ECG will be recorded using a PC based ECG monitor (Cardioperfect Workstation; Welch Allyn, New York, USA). Resting heart rate (HR) variability will be assessed immediately after the standard ECG using the same lead positions for a total of 5 min.

Electrocardiographic imaging (ECGI)
ECGI is the process of combining heart and torso geometry with multiple body surface potentials to generate epicardial electrograms and panoramic maps of cardiac excitation [17–19]. ECGI has been extensively validated in ex vivo animal studies using a torso-tank experimental method [20, 21] and in vivo animal experiments. It has also been shown to accurately correlate with invasive EP mapping in ventricular tachycardia (VT) [22], it has been validated when applied to cardiac computed tomography [23] and using a multi-electrode ECGI sock intraoperatively [24]. ECGI has also been successfully combined, by our group, with CMR in patients amyloidosis [25] and arrhythmogenic cardiomyopathy [26]. It is with this method that we can detect subtle EP abnormalities that are often missed by conventional 12-lead ECG [27]. The challenge for the cardiologist is identifying whether
subtle EP abnormalities, which are more prevalent in the ageing myocardium, will put an individual at higher risk of sudden cardiac death or not [28]. In order to bridge this knowledge gap, ECGI will allow us to link regional EP aberrations with myocardial substrate changes for mechanistic insights into the aetiology of arrhythmias.
Our pipeline will enable reconstruction of epicardial surface biopotentials projected onto the cardiac geometry using inverse solution mathematics to measure epicardial activation and repolarization parameters which can then be spatially correlated with myocardial tissue characteristics derived by CMR. [29].

We recently developed a re-usable and dry-electrode-based CMR-compatible vest to permit high-throughput ECGI research and seamless integration with CMR (up to 3 Tesla [T]). This was achieved in collaboration with textile engineers at the École Nationale Supérieure des Arts et Industries Textiles (ENSAIT, Roubaix, France) and with g.Tec medical engineering GmbH (Schiedlberg, Austria). The technology consists of two matching wash-resistant and reusable garments: one embedded with 256 dry electrodes (the electrode vest) and the other (the CMR-safe marker vest) embedded with co-registered fiducial markers (Beekley medical, Bristol, Connecticut).

The signal from each electrode is collected and amplified using the HIamp 256 bundle GT-8016/USBamp GT-0216 and processed for analysis in specially designed software, g.Recorder (g.Tec).

Prior to CMR, the participant will wear the electrode vest which is secured onto the chest using adjustable fasteners. Body surface potentials will be measured from all electrodes at a sampling frequency of 2400 Hz for 5 min at rest, in the supine position. The electrode vest is then removed leaving only the marker vest which the participant wears inside the CMR scanner. Accurate electro-anatomical mapping of ECGI and CMR data will then be achieved through an initial 4 mm contiguous-slice transaxial black-blood thoracic stack for marker co-registration. Our ECGI workflow is summarized in Fig. 2.

**CMR Outline**

Imaging will be performed on a 3 T [200 × 80mT/m x mT/m] MRI system (Magnetom Prisma, Serial Number 166032, Siemens, 60 cm bore) operating VE11C-SP01, with an 18-channel phased-array chest coil and spine array (up to 24-elements) equipped with Gadgetron [30] (Linux box, 24 cores). Participants will be scanned using a protocol which combines standard and advanced CMR imaging techniques and is designed to be completed within a 45 min scanning session. Standardized CMR protocols have been described in detail previously [31, 32]. Brachial and non-invasive central BP assessment using a Cardioscope II BP+ device (USCOM, Sydney, Australia; cuff on the right arm) will be performed at baseline in the scanner bore, prior to the scan starting, followed by resting, peak stress (during adenosine administration) and recovery measurements, also inside the scanner bore. Continuous digital pulse oximeter monitoring will be obtained throughout using the Nonin 7500FO fiber optic MRI table-top pulse oximeter (Nonin,

<table>
<thead>
<tr>
<th>Category</th>
<th>Pathology</th>
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<tr>
<td>Ventricular structure</td>
<td>Moderate-severe LV hypertrophy</td>
</tr>
<tr>
<td>Ventricular function</td>
<td>Moderate-severe LV impairment</td>
</tr>
<tr>
<td></td>
<td>Moderate-severe RV impairment</td>
</tr>
<tr>
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<td>Moderate-severe atrial dilation</td>
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<tr>
<td>Valvular pathology</td>
<td>Structural valve abnormalities e.g. bicuspid aortic valve</td>
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<td>Moderate-severe pericardial effusion</td>
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<td>Shunts</td>
<td>Cardiac tamponade</td>
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<td>Cardiac masses</td>
<td>Ventricular septal defect</td>
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<td>Intracardiac mass</td>
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<td></td>
<td>Valvular mass e.g. vegetation/fibroelastoma</td>
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<td>Myocardial inflammation</td>
<td>Acute/chronic myocardial inflammation</td>
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<td>Myocardial infarction</td>
<td>Acute/chronic myocardial infarction</td>
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<td>Perfusion defects</td>
<td>Clinically significant inducible perfusion defects</td>
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<tr>
<td>Extra cardiac findings</td>
<td>Other suspected, significant or life-threatening finding</td>
</tr>
</tbody>
</table>

LGE, Late gadolinium enhancement; LV, left ventricle; RV, right ventricle
Plymouth, USA). The CMR protocol is outlined in Fig. 3 and MR sequence details summarized in Table 3.

**Cardiac structure and function**

Initial CMR scout imaging will include sagittal, coronal and transverse piloting of the chest in single heartbeat, free-breathing, acquisitions. This will be followed by a thin contiguously sliced transaxial set of turbo spin echo (TSE) images (4 mm thickness no gap) across the chest (approximately 90 slices) using a half-Fourier acquisition single-shot turbo spin echo (HASTE) sequence. These T₂ weighted, dark blood, images will localize the fiducial markers on the ECGI vest and facilitate coregistration during post processing. Local 2nd order cardiac
shimming will be prescribed onto HASTE data and automatically copied to the rest of our protocolled sequences. To assess left ventricular (LV) structure and function we will acquire three long axis cines (vertical long axis, horizontal long axis and left ventricular outflow tract) and an LV short axis cine stack (SAX) using breath-held ECG gating. To assess right ventricular (RV) structure and function we will acquire RV outflow tract (sagittal and oblique-sagittal planes) and vertical RV long-axis bSSFp cines. For more detailed aortic valve (AV) assessment we will acquire coronal LV outflow tract (LVOT) and AV cines.

**Tissue characterization**

The native myocardial T$_1$ time, before the administration of gadolinium based contrast agent (GBCA), can lengthen even in the absence of late gadolinium enhancement (LGE) and it is this ability to detect so-called ‘subclinical’ fibrosis that will make it particularly useful for our analysis of myocardial resilience in old age [33, 34]. In order to obtain native T$_1$ maps we will acquire breath-held single shot bSSFp motion-corrected (MOCO) modified Look-Locker inversion recovery (MOLLI) images using prototype 5 s(3 s)3 s at the basal, mid and apical LVSAX levels, with volume selective magnetic field strength ($B_0$) shimming, in diastole [35, 36]. Standard deviation (SD) maps will be automatically generated and used to cross check the reliability of corresponding MOLLI T$_1$ values in co-registered regions of interest, aiming for a SD of <40 ms. A description of this technique has been outlined elsewhere [37].

To create extracellular volume (ECV) maps we will perform two co-registered T$_1$ mapping acquisitions: before and approximately 10 min after infusion of the second dose of GBCA (Dotarem, Gadoterate meglumine, Guerbet, France) using MOLLI prototypes 5 s(3 s)3 s and 4 s(1 s)3 s(1 s)2 s respectively, at the basal, mid and apical levels.

<table>
<thead>
<tr>
<th>Description</th>
<th>Transverse anatomy</th>
<th>LV short axis cine</th>
<th>Native and post GBCA T$_1$ map</th>
<th>Native T$_2$ map</th>
<th>Aortic flow</th>
<th>Perfusion</th>
<th>LGE</th>
<th>4D flow</th>
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<tr>
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<td>Cine imaging bSSFp</td>
<td>bSSFp, MOCO, single shot MOLLI</td>
<td>bSSFp, MOCO, single shot</td>
<td>Gradient echo phase contrast cine</td>
<td>bSSFp, MOCO, single shot</td>
<td>Bright blood MOCO bSSFp with PSIR</td>
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<td>BH</td>
<td>BH</td>
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2-D, 2-dimensional; AP, antero-posterior; BH, breath hold; bSSFp, balanced steady-state free precession; CS, compressed sensing; FB, free breathing; GRAPPA, generalized auto-calibrating partially parallel acquisitions; HASTE, half-fourier single-shot turbo spin-echo; MOCO, motion-corrected; MOLLI, modified look-locker inversion recovery; PE, phase encoding; PSIR, phase sensitive inversion recovery; PT, prospectively triggered; RG, retrospectively gated; RL, right left; SA, short axis; TE, echo time; T-PAT, temporal parallel imaging; TR, temporal resolution; TSE, turbo spin echo. Other abbreviations as in Table 2

*LV SAX cine stack is 8 mm slice thickness
LVSAX slice. We will then generate pixel-wise ECV maps by determining the ratio of the change in T₁ values, following correction for blood cell density (using validated point-of-care hematocrit readings) [31, 38, 39]. This will be done by transposing regions of interest (ROIs) in the septal midwall (or alternative remote myocardial zone) and blood pool from native onto the post-GBCA T₁ maps. Two types of ECV maps will be generated per participant for cross-validation: (1) A synthetic ECV map, which will be generated directly from the scanner using a synthetic hematocrit derived from native T₁ values, as described previously [39]; (2) A conventional ECV map, which will be generated at the end of each scan using the measured blood hematocrit (derived from the hemoglobin value obtained from point of care testing).

We will perform single-slice breath-held T₂ mapping using a MOCO single shot bSSFP sequence at the level of the mid LV short axis and prior to GBCA administration.

**Aortic flow imaging**

With advancing age, biochemical, histological and blood flow changes in the main arteries result in wall stiffening [40]. Arterial stiffness has traditionally been measured in peripheral vessels due to ease of access, even though aortic stiffness is a stronger prognostic marker because of its resultant increase in LV afterload and wider downstream end-organ damage [41]. To analyze the effects of aortic stiffening in older age we will measure local aortic distensibility (derived from aortic area measurements) and regional pulse wave velocity (PWV) across the ascending aorta and aortic arch. Both regional and local measurements of aortic stiffness capture variation in aortic tissue composition across its course, but have different sensitivities to pathology related to arterial stiffening and are associated with different cardiovascular outcomes [42]. In order to obtain measurements of aortic length, we will initially acquire an ECG-gated, free-breathing sagittal cine of the aorta (‘candy cane’ view). This image will then serve as a localizer to pilot blood-flow velocity maps at the level of the bifurcation of the main pulmonary artery transecting the ascending and descending thoracic aorta. We will use a high temporal resolution phase-contrast free-breathing ECG-gated gradient echo sequence as previously described [31, 32, 43] (imaging parameters provided in Table 3). Aortic flow imaging will be performed at baseline and repeated during peak stress (immediately before the stress perfusion acquisition) alongside simultaneous pressure waveform and pulse oximetry measurements. An outline of the methods used to derive pulse wave velocity is summarized in Fig. 4. Measurements of flow velocity and pressure waveforms will be used to derive wave intensity, wave separation, reservoir pressure and reflection magnitude as previously described [44].

**Stress perfusion imaging**

For myocardial ischemia evaluation we will perform stress perfusion imaging using adenosine on all participants using a dual-cannula approach (typically sited in the antecubital fossae) which has been previously described [45]. Intravenous adenosine will be infused peripherally at 140 µg/kg/min from one of the cannulas for a minimum of 3 min and a maximum of 4 min to achieve an adequate response (assessed through a HR increment of 10 beats per minute and/or the development of symptoms). A higher dose of 175 µg/kg/min may be infused for a further 2 min in the event of limited symptoms or failure to increment HR by a minimum of 10 beats per minute. The first dose of Dotarem (0.05 mmol/kg) will be injected via the other canula at a rate of 4 mL/second along with 20 mL of saline. The adenosine infusion will be stopped during the first pass perfusion image acquisition which will be acquired using a free-breathing MOCO saturation-recovery bSSFP sequence to cover 3 LVSAX slices across every heartbeat. Approximately 10 min following peak stress, rest perfusion imaging (using the same sequences described above) will then be completed without adenosine, using the second dose of Dotarem.

**Late gadolinium enhancement**

To assess the presence of LGE we will acquire an LVSAX stack of conventional (‘bright-blood’) LGE images using a free-breathing MOCO bSSFP sequence with phase-sensitive inversion-recovery (PSIR) [46]. LGE acquisitions will commence approximately 5 min after the second dose of Dotarem.

**4-Dimensional flow**

Four-dimensional (4-D) intracavitary flow CMR provides comprehensive hemodynamic flow assessments, taking into account the multidirectional and multidimensional nature of blood flow through the cardiac chambers [47] with the potential to improve our understanding of the ageing cardiovascular phenotype. We will compare the use of respiratory-navigated vs. free-breathing retroflexibly ECG-gated, spoiled gradient echo sequences (work-in-progress CS785B, Siemens Healthineers, Erlangen, Germany), with field of view covering the LVOT, proximal aorta, and all 4 cardiac chambers. The acquisition will be performed at the end of the CMR protocol with GBCA on board to allow for improved blood myocardial contrast. Sequence parameters based on the published literature [48] are provided in Table 3.

**Quality assurance**

Normal values for key CMR biomarkers collected in MyoFit46 will be generated from healthy volunteer
datasets acquired using the same sequences on the same magnet system. The upper and lower range of normal values will be defined as the mean ± 1.96 standard deviations from the normal data, according to standardized recommendations [38].

Prior to commencing native myocardial $T_1$ mapping in MyoFit46, two quality assurance steps will be implemented.

A quality assurance phantom testing protocol for $T_1$, $T_2$ and ECV mapping will be undertaken to verify the stability of our magnet-sequence combination over time and throughout the project lifecycle. We will use the medical-device grade $T_1$ mapping and ECV standardization ($T_1$MES) phantom [49] (field-strength specific for 3 T with regulatory clearance from the Food and Drug Administration and Conformité Européene-marking) as well as the $T_2$ Phantom [50] (field-strength agnostic). Both devices have been previously developed by our group in collaboration with Resonance Health (Perth, Australia) and with the US and German National Metrology Institutes (National Institute of Standards and Technology and Physikalisch-Technische Bundesanstalt respectively) [49, 51]. At study start and study close both phantoms (stored in the temperature-controlled MRI scanner environment) will be interrogated by flip angle maps and be scanned using slow overnight inversion recovery spin echo (IRSE) and multi-echo SE, to establish reference $T_1$ and $T_2$ values. At study start, study close
and once every 2 months between these timepoints, the T₁ MES phantom will additionally be scanned in accordance with the T₂ MES user manual [51, 52] using the fat/water sequence for field maps and using 3 repeats of the MOLLI 5 s(3 s)3 s and 4 s(1 s)3 s(1 s)2 s prototypes. At study start, study close and once every 2 months between these timepoints, the T₂ phantom will additionally be scanned using the fat/water sequence and three repeats of the T₁ mapping sequence. Phantom body temperature will be recorded prior to the start of each scan session, aiming for an ambient CMR room temperature of 21–22 °C. A schematic of our phantom-based quality assurance framework for MyoFit46 is presented in Fig. 5.

**Imaging pipelines and analyses**

**Primary imaging analyses**

**Imaging pipelines**

All CMR image DICOMs (Digital Imaging and Communications in Medicine), derived results and associated ROIs will be stored on a customized web-based server running XNAT 1.6.5 and hosted at UCL. DICOM-compliant imaging format will be used for all MRI data including raw list mode data and images that have been reconstructed on the scanner using the Gadgetron framework. CMR data will be analyzed using Circle Cardiovascular Imaging, cvi [42] version 5.3.2 (Calgary, Canada).

**CMR analysis**

Standardized CMR image interpretation and post-processing will be performed as previously described [31, 32, 53, 54]. Briefly, analyses will include volumetric measurements of the 4 cardiac chambers (using manual epicardial and endocardial contour tracings), assessment of biventricular and valvular function, analysis of native myocardial T₁ and T₂, stress perfusion imaging and LGE assessment. Papillary muscles and trabecular tissue will be consistently included in the LV mass measurements, as previously described [53, 54]. Left ventricular volumetric and maximal wall thickness analyses will additionally be performed using the previously validated openCARE artificial intelligence (AI) analysis platform. This platform has already been shown to have improved precision when compared to human observers in independent studies [55].

A standard operating procedure has been developed for the manual analysis of LGE, T₁, T₂ and ECV by trained researchers. For T₁, T₂ and ECV analysis, epicardial and endocardial contours will be drawn manually with 10% erosion from the blood myocardial boundary and divided according to the 16-segment American Heart Association (AHA) model. On the native and post-GBCA MOLLI T₁ maps, a ROI will be drawn in the mid septum and co-registered with automatically generated pixel-wise SD maps. Semi-automated LGE quantification will be carried out using a 2 SD and a 5 SD approach following manual selection of an ROI in an area of remote/dark myocardium [36, 56]. We will also use the full width half maximum technique [57] for comparison against the other methods. Segments with LGE will be annotated to allow for subsequent interpretation of the mapping data [54].

**Aortic flow analysis**

Proximal ascending and descending thoracic aortic contours will be traced semi-automatically using validated software (ArtFun, INSERM U678, Paris, France) to derive flow-velocity waveforms [43, 58]. Aortic distensibility will be measured at local, specified regions of the aorta and calculated according to established methods [43, 59]. PWV over the upper thoracic aorta will be measured using length and transit time (from derived aortic velocity profiles) [43].

**4-Dimensional flow analysis**

Intracavitary 4-D flow CMR will allow us to derive parameters such as turbulent kinetic energy, wall shear stress, PWV and variations in LV vortices [47, 60]. For example, previous studies have shown that there is an inverse relationship between LV diastolic vortices [61] and ageing and that turbulent kinetic energy could be a promising subclinical marker of LV dysfunction [62]. 4-D flow CMR also allows for detailed valve quantification with particularly promising results in mitral regurgitation, since it allows for direct quantification of flow at the level of the valve which remains valid even in the presence of other valvular lesions or shunts [63]. These markers of intracavitary hemodynamics will be used to characterize pathophysiological responses to ageing beyond conventional methods of flow. 4-D flow image post-processing will be performed using CAAS (PIE Medical Imaging, Maastricht, The Netherlands). During analysis we will ensure that any contours drawn in the magnitude images are cross-referenced with velocity images to take into account large movements and the effects of partial volume. Velocity data will be encoded into individual vectors, allowing us to assess local
Fig. 5 (See legend on previous page.)
patterns of flow such as vortices, visualize and quantify wall shear stress and study kinetic energy [64].

**ECGI analysis**

Body surface ECGs collected by g.Recorder will be stored and analyzed offline using previously established methods [18, 25, 26]. Signal averaging will be performed using in-house, customized, software (Matlab, MathWorks, Natick, MA) to enhance the quality of the signals from body surface potentials. The location of the fiducial markers on the torso corresponding to the electrode positions will be combined with the heart-torso geometry obtained from the CMR HASTE stack and reconstructed to create epicardial meshes using commercially available software (Amira, ThermoFisher, MA, USA, version 2020.3). Unipolar epicardial electrograms will then be reconstructed by solving the inverse problem in collaboration with the Rudy laboratory [19, 26]. Generated epicardial maps will enable calculation of epicardial activation and repolarization times and corresponding isochrone maps will be generated for visualization. These maps will be used to identify discrete areas of abnormal electrophysiology and correlated with matching substrate abnormalities detected by CMR. An exemplar pilot isochrone map that we have created using this CMR-ECGI pipeline is reproduced in Fig. 6. At scale validation of the co-registration process with CMR has not been performed before and in order to address this we will carry out test–retest reproducibility and intra-/inter-observer variability studies. All raw and post-processed ECGI data will be stored securely in the project-specific UCL XNAT repository.

**Cardiac computational modelling**

In order to extend our multi-dimensional deep phenotyping of the ageing heart we will perform advanced cardiac computational modelling in collaboration with the Barcelona Supercomputing center (Barcelona, Spain). Using the established ‘Alya Red’ [65] supercomputer driven data modelling protocol [65–67], we will combine epicardial electrophysiological data (from ECGI) with the rich myocardial structural, functional, perfusion and flow data obtained from CMR to create personalized electro-mechanical-flow whole-heart simulations per participant [68].

![Fig. 6 ECGI analysis workflow.](image)
Study analysis Plan

Statistical analysis

Both overt and subclinical cardiovascular disease biomarkers (binary or continuous variables, such as LV ejection fraction, native T₁, ECV, LGE, etc.) will be determined through this proposed CMR-ECGI deep phenotyping protocol and such biomarkers will serve as either outcome or predictor variables in the various analyses planned. On the one hand, MyoFit46 will unravel how key exposures during early-, mid- and later life (e.g. environmental pollution, communicable disease exposure, socioeconomic status, etc.) interact with traditional cardiovascular risk factors (e.g. smoking, obesity, alcohol intake, etc.) and multimorbidity to determine cardiovascular disease (outcome) in older age. On the other hand, we will also prospectively study the association between discovered cardiovascular disease biomarkers (predictors) and key outcomes of interest ascertained during long-term follow-up of the birth cohort participants (e.g. functional status or major adverse cardiovascular events such as hospital admissions, nonfatal stroke, myocardial infarction, cardiovascular death, etc.).

Since several of our exposures of interest are either categorical or represent trajectories/time-varying exposures; associations between exposures and outcomes will be analyzed using a range of statistical approaches relevant for life course data [69–71]. Associations between trajectories of exposures and cardiovascular disease biomarkers will be analyzed using latent growth curve analysis [71]. In order to subsequently link with outcomes we will use a two-step process as previously described [72]. We will use structural equation models using full information maximum likelihood to account for missing data in NSHD under missing at random assumptions.

Sample size estimates have been calculated to permit detection of a small effect size (Cohen's $f^2 = 0.03$) using multivariable regression analysis. A sample size of 533 achieves 80% power to detect an $f^2 \geq 0.030$ attributable to up to 9 independent variable(s) using an F-Test with a significance level (alpha) of 0.050. For categorical outcomes a logistic regression of a binary response variable on a continuous, normally distributed variable with a sample size of 533 observations achieves 92% power at a 0.05 significance level to detect a change corresponding to an odds ratio of 1.5.

Data sharing

A data sharing policy will be published and managed through MRC Skylark website (https://skylark.ucl.ac.uk/Skylark) including access to research publications, talks presented at international conferences and links to academic social media accounts. Raw data will be archived for a minimum of 10 years within the UCL open XNAT server (https://ucl-open-xnat.cs.ucl.ac.uk/app/template/Login.vm#/) and these data will be shared with bona fide researchers along with the relevant clinical and demographic metadata that will be archived in REDCap. CMR and ECGI standard operating procedures will be electronically archived alongside the data, and as hard copies. Software pipelines will be stored in electronic lab books, including software version numbers.

Discussion

MyoFit46 aims to combine the rich life course data of the NSHD with cutting-edge CMR and ECGI technologies and multi-dimensional cardiac simulations to provide new insights into the long-term myocardial sequelae of key exposures from birth. Uniquely the NSHD provides us with detailed, prospectively collected data since birth, allowing us to study the individual, combined and cumulative impacts of risk factors, and their mediators on myocardial health by older age. Previous older age cohorts have carried out CMR imaging, but none have performed CMR-ECGI integration with stress perfusion, multi-parametric mapping, and 4-D intracardiac flow as is being proposed here. ECGI will enable us to correlate overt and subclinical markers of arrhythmogenesis with CMR-derived structural and functional abnormalities. By merging these complex structural, functional, hemodynamic and electrophysiological data into cardiac computational models we be deepening our understanding of the ageing myocardium, representing a world first in terms of CMR-driven personalized care. We expect our results to impact public health by identifying novel risk factors for cardiovascular disease and addressing the growing challenge of multi-morbidity with the aim of maintaining myocardial health into older age for the benefit of humanity globally.

Abbreviations

AHA: American Heart Association; AI: Artificial intelligence; AV: Aortic valve; BHF: British Heart Foundation; BP: Blood pressure; bSSFP: Balanced steady state free precession; CMR: Cardiovascular magnetic resonance; COVID-19: Coronavirus disease 2019; CVD: Cardiovascular disease; Dicom: Digital Imaging and Communications in Medicine; DPA: Data protection act; ECG: Electrocardiogram; ECGI: Electrocardiographic imaging; ECV: Extracellular volume; EP: Electrophysiology; FSE: Fast spin echo; 4-D: Four-dimensional; GBCA: Gadolinium-based contrast agent; GDPR: General Data Protection Regulation; HASTE: Half Fourier acquisition single shot turbo spin echo; HR: Heart rate; IRSE: Inversion recovery spin echo; LGE: Late gadolinium enhancement; LV: Left ventricle; LVOT: Left ventricular outflow tract; MOCO: Motion corrected; MOLLI Modified look locker inversion recovery; MRC: Medical Research Council; MRI: Magnetic resonance imaging; NHS: National Health Service; NSHD: National Survey of Health and Development; PSIR: Phase-sensitive inversion-recovery; PWV: Pulse wave velocity; REC: Research Ethics Committee; REDCap: Research electronic data capture project; ROI: Regions of interest; RV: Right ventricle; SARS-CoV-2: Severe acute respiratory distress coronavirus 2; SAX: Short axis stack; SD: Standard deviation; T: Tesla; TSE: Turbo spin echo; UCL: University College London; VT: Ventricular tachycardia.
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Authors’ contributions
MW and GC designed the study, obtained funding and ethical approval for the study, wrote this manuscript and will be responsible for data collection, analysis and interpretation of the results. DF, MAF, GJ, FC, CD and LHH have assisted with pilot data collection. AH, C, AW, AR, JS and RH provided expert review of the manuscript on aspects relating to the NSHD cohort and statistical analysis. AB provided expert input on aortic flow analysis and RHD on the technical CMR aspects. PL, MO and YR provided expert input on the ECG aspects. CM, JAS and MV provided input on the cardiac simulations. XT, ST and CG fabricated the CmR-ECGI vest. All authors read and approved the final manuscript.

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Availability of data and materials
Not applicable.

Declarations

Ethical approval and consent to participate
All participants will undergo formal written consent prior to all procedures as per our ethical approval. Ethics approval has been obtained for the study: London REC—Queens Square (Ref: 19/LO/1774). London REC—Queens Square, IRAS: 254776.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing of interests.

Study status
This study is ongoing having commenced participant recruitment.

Related articles

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References


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