Staphylococcus aureus injection drug use-associated bloodstream infections are propagated by community outbreaks of diverse lineages

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Abstract

Background The ongoing injection drug use (IDU) crisis in the United States has been complicated by an emerging epidemic of Staphylococcus aureus IDU-associated bloodstream infections (IDU-BSI).

Methods We performed a case-control study comparing S. aureus IDU-BSI and non-IDU BSI cases identified in a large US Midwestern academic medical center between Jan 1, 2016 and Dec 21, 2019. We obtained the whole-genome sequences of 154 S. aureus IDU-BSI and 91 S. aureus non-IDU BSI cases, which were matched with clinical data. We performed phylogenetic and comparative genomic analyses to investigate clonal expansion of lineages and molecular features characteristic of IDU-BSI isolates.

Results Here we show that patients with IDU-BSI experience longer durations of bacteremia and have lower medical therapy completion rates. In phylogenetic analyses, 45/154 and 1/91 contemporaneous IDU-BSI and non-IDU BSI staphylococcal isolates, respectively, group into multiple, unique clonal clusters, revealing that pathogen community transmission distinctively spurs IDU-BSI. Lastly, multiple S. aureus lineages deficient in canonical virulence genes are overrepresented among IDU-BSI, which may contribute to the distinguishable clinical presentation of IDU-BSI cases.

Conclusions We identify clonal expansion of multiple S. aureus lineages among IDU-BSI isolates, but not non-IDU BSI isolates, in a community with limited access to needle exchange facilities. In the setting of expanding numbers of staphylococcal IDU-BSI cases consideration should be given to treating IDU-associated invasive staphylococcal infections as a communicable disease.

Plain language summary

Persons who inject drugs are at increased risk of developing a bloodstream infection caused by the bacterium Staphylococcus aureus. To investigate whether this risk is due to transmission of the bacterium within this community, we compared the complete set of genes (genome) of S. aureus isolated from people with bloodstream infections who do and do not inject drugs. S. aureus causing bloodstream infections in persons who inject drugs were much more likely to belong to one of multiple networks of very closely related subtypes, demonstrating that in some communities the bacteria causing this type of infection can likely be directly transmitted from person to person through high-risk injection drug use practices, such as sharing needles. Therefore, invasive infections in persons who inject drugs can spread like communicable diseases and this can inform future policy on how to prevent them.
The injection drug use (IDU) crisis in the United States (US) is complicated by emerging syndemics of infectious diseases among people who inject drugs (PWID). Since 2015 the Centers for Disease Control and Prevention and health departments across the US have identified human immunodeficiency virus (HIV) and viral hepatitis outbreaks attributed to bloodborne transmission within IDU networks. Co-epidemics of invasive bacterial infections among PWID have also been identified. *Staphylococcus aureus* is the most common pathogen causing these invasive infections, which are often associated with staphylococcal bloodstream infection (BSI). PWID are approximately 16.3 times more likely than peers to develop invasive staphylococcal infections, with one in every ten invasive staphylococcal infections in the US now related to IDU. Efforts against invasive staphylococcal infections in general are complicated by the existence of multiple disease subtypes, including central line-associated BSI (CLABSI), endocarditis, osteomyelitis, septic arthritis, epidural abscess. Each disease manifestation necessitates preventative and therapeutic strategies tailored to characteristic epidemiology, pathobiology, and host risk factors. However, the epidemiology and transmission of IDU-associated *S. aureus* invasive infections is poorly defined, as few studies have investigated it as a separate disease entity.

Staphylococcal IDU-associated bloodstream infections (IDU-BSI) may be spurred by factors characteristic of PWID, but absent among individuals with conventional forms of BSI (cBSI). For example, high rates of *S. aureus* contamination of cookers and filters used in preparation of controlled-release opioids for IDU have been observed. Alternatively, it is speculated that person-to-person transmission through contaminated needles contributes to IDU-associated infections. Early reports on an IDU-associated outbreak in Detroit, US identified being unhoused and shared injection equipment as risk factors, and phage typing and antibiotic susceptibility data identified common *S. aureus* lineages shared by many of these cases. Subsequent reports further argued a link between IDU and transmission of methicillin-resistant *S. aureus* (MRSA) lineages among PWID. However, these studies lacked comparator groups and employed low-resolution molecular methods and small cohort sizes, which limits the interpretation of their findings. As community IDU-associated transmission represents an attractive target for BSI prevention, the existence and impact of person-to-person transmission of pathogenic strains among PWID must be clearly established.

Other findings support distinguishing IDU-BSI from cBSI. IDU-associated staphylococcal infections have been associated with prolonged bacteremia duration and infectious sequelae such as endocarditis, possibly due to challenges faced by PWID in completing the standard-of-care, multiweek treatment regimens prescribed by guidelines. Despite these poor prognostic indicators, emerging evidence suggests that, compared to cBSI, IDU-BSI exhibits comparable to lower mortality rates. A mix of host and pathogen factors likely influence these discrepant observations. However, the paucity of studies comparing IDU-BSI to non-IDU staphylococcal BSI has resulted in a critical knowledge gap in our understanding of unique factors governing IDU-associated invasive disease.

We hypothesize that features unique to PWID mediate biological differences between *S. aureus* IDU-BSI and cBSI. Firstly, shared behaviors and socioeconomic conditions associated with IDU may predispose to characteristic clonal expansion of *S. aureus* pathogenic strains among PWID, raising the possibility that IDU-BSI is a communicable disease. Secondly, IDU practices predispose to direct inoculation of bacteria into the bloodstream. This bypass of major immunological barriers may obviate the role of microbial factors that mediate early stages of infection in cBSI and permit a wider diversity of otherwise-less virulent *S. aureus* strains to cause invasive disease in PWID. To investigate this, we performed extensive comparative genomics analysis of clinical isolates in a case-control study of *S. aureus* BSI occurring in a large U.S. Midwest medical center over a 4-year period. We identified clonal expansion of multiple *S. aureus* lineages among IDU-BSI isolates, but not non-IDU BSI isolates, in a community with limited access to needle exchange facilities.

**Methods**

**Setting and case definitions.** This study was approved by the Institutional Review Board of Washington University in St. Louis (IRB# 201804183, 201907187, 201911072, 202007171). A waiver of informed consent was granted by the Washington University IRB for isolate collection as all *S. aureus* specimens were obtained during routine clinical care and saved for quality improvement purposes. A waiver of informed consent for data abstraction from the chart was issued as data were pre-existing in the chart and many patients were already deceased or would otherwise have been unable to be contacted. This study was performed at Barnes-Jewish Hospital (BJH), a 1250-bed, academic, tertiary care center serving the greater metropolitan area of St. Louis, Missouri and surrounding areas. Through electronic health record (EHR) review, we retrospectively identified cases between 1/2016–1/2019 associated with *S. aureus* recovered from blood cultures or heart valve surgical specimens during routine medical care, as described previously. We also included cases prospectively identified between 1/2019–12/2019 as part of a local quality improvement initiative and a CDC Developing Healthcare Safety Research Contract. Only the first documentation of infection per patient was included in our analysis, and the date of the first *S. aureus* isolate per patient (“index date”) was identified for each case. Cases were manually reviewed by an infectious diseases physician (L.R.M.) and classified as “IDU-BSI” (defined as IDU directly preceding index date) or “non-IDU” (cases lacking history of any mode of substance use), as described previously. Patient demographics, drug use history, clinical course, and clinical microbiological data were obtained from EHR. Duration of bacteremia was defined as the number of calendar days between index date and date of last positive surveillance blood culture. Prolonged bacteremia was defined as duration of culture-proven bacteremia ≥ 5 days.

As a composite comorbidity indicator, we calculated each patient’s Elixhauser Comorbidity Index by screening for 31 comorbidity diagnoses among ICD-10 codes documented prior to the index date. To compare survival distributions for primary endpoint of 1-year mortality (measured from date of hospital admission to account for in-hospital deaths), we examined univariate associations with potential demographic, infection type and medical comorbidity confounders. Variables with $p < 0.1$ in univariate analyses were integrated into a multivariable Cox proportional hazards model. "Drug abuse" was excluded from Elixhauser Comorbidities as this risk factor was already captured under 'injection drug use origin'. Covariates were assessed using backwards stepwise regression, for violation of the proportional hazards assumption and assessed using log-negative-log survival plots. Hazard ratios (HRs) and 95% confidence intervals were calculated. Descriptive statistics were calculated using SPSS v26 (Chicago, IL). Figures were created using GraphPad Prism 9 (San Diego, CA).

**Identification, susceptibility testing and banking of *S. aureus* isolates.** *S. aureus* clinical isolates were cultured and analyzed in the BJH clinical microbiology laboratory as part of the routine medical care. All isolates recovered from BSI are routinely stored.
Whole-genome sequencing (WGS) and genomic analysis. Illumina sequencing was performed on genomic DNA extracted from a subset of clinical isolates (Fig. 1). Genome assembly and analysis, including core genome alignment, multilocus sequence typing (MLST), and identification of antimicrobial resistance and virulence factor genes, were performed based on well-established processing pipelines. Isolates were propagated on sheep’s blood agar, and S. aureus isolate genomic DNA was extracted with the Qiagen Bacteremia Kit (Qiagen, Germantown, MD, USA). DNA was used as input for Illumina sequencing libraries with the Nextera kit (Illumina, San Diego, CA, USA). Pooled libraries were sequenced on a NextSeq HighOutput platform (Illumina) to obtain 2 × 150 bp reads. Following demultiplexing by barcode, reads had adapters removed with Trimmomatic v0.38. Reads were then assembled into draft genomes using de-novo assembler Unicycler v0.4.27. Scaffolds.fasta files were used for downstream analysis. Whole-genome sequence read files are uploaded to NCBI under BioProjects PRJNA694991 and PRJNA695316.

Species identity of each genome was confirmed using the ANIm method from pyANI v0.2.7. ANIm ≥96% compared to the S. aureus reference genome (strain NCTC 8325, GCF_000013425.1) was used as the species cutoff. In silico screen for antibiotic resistance genes and multilocus sequence typing (MLST) was performed with ResFinder v4.0 and MLST-check, respectively. Prokka v1.13.7 was run on scaffold files to identify open reading frames >500 bp in length. Scaffolds were screened for the presence of genes of interest with BLAST using the following parameters: [−evalue 1e−10 -perc_identity 90 -gapopen 5 -gapextend 5] and a cutoff of >90% amino acid identity. Screened genes were selected a priori according to their predicted role in disease, based on literature review and are listed in Supplementary Data 1.

Phylogenetic and clonal analysis. For phylogenetic analysis, the gff files produced by Prokka were used to construct a core genome alignment with Roary v3.13.35. The alignment was used to generate a maximum-likelihood tree with raxML v8.2.11.36, and visualized with iTOL. For clonality analysis, Snp-sites v2.4.0.38 was used to remove indels and create multiFasta alignment containing the single nucleotide polymorphism (SNP) sites for each core genome. Pairwise SNP counts between isolates were calculated and plotted. SNP distances of isolates repeatedly obtained from the same patient were used as reference, to empirically determine SNP distance cutoffs indicating clonal transmission. SNP distances between surveillance isolates obtained from the same patient served as reference to empirically determine a cutoff to define a clonal relationship between isolates obtained from different individuals. Interactions that met the cutoff were visualized using Cytoscape v4.0.39. Source data are available in Supplementary Data 2.
hospitalizations for S. aureus

Clinical characteristics of IDU-BSI

Results

Clinical characteristics of IDU-BSI. We identified 173 and 1261 hospitalizations for S. aureus IDU-BSI and non-IDU cBSI, respectively, from January 2016 through December 2019. The proportion of cases associated with IDU increased from 9.1% in 2016 to 13.4% in 2019, with 12.1% of BSI cases being IDU-BSI. Among BSI cases, AMA discharges and prolonged bacteremia (≥25 days) was more common among IDU-BSI with lower rates of completion of standard of care intravenous (IV) antibiotics but did not translate into higher 1 year mortality rates. Kaplan–Meier survival curves comparing S. aureus IDU-BSI and non-IDU BSI with number at risk below the graph.

Statistics and reproducibility. Unless otherwise stated, comparisons of categorical data were performed by Fisher’s exact test, while comparisons of continuous variable were tested using the Mann-Whitney U test. All tests were two-tailed, and statistical significance was defined as p ≤ 0.05 or a 95% confidence interval (CI) excluding 1.00 for an odds ratio (OR). Statistical analyses were performed on JMP Pro software (v15) or R software.

Clinical characteristics of cases included in WGS cohort. We obtained genome sequences from 289S. aureus isolates from patients with S. aureus endovascular infections. These included 245 index and 44 surveillance isolates (Supplementary Table 1). The demographic and clinical characteristics of patients with cases associated with index IDU and non-IDU isolates chosen for WGS (n = 154 and 91), respectively, are compared in Table 1. Sequenced S. aureus IDU-BSI strains occurred among younger patients who were more likely to experience homelessness (p < 0.001), and to be co-infected with hepatitis B (HBV) (p = 0.017), hepatitis C (HCV) (p < 0.001), or HIV (p = 0.002). IDU-BSI cases were more likely to be associated with infective endocarditis (p < 0.001), and less likely to be associated with the presence of a central line at diagnosis or to present without metastatic sites of infection (“isolated bacteremia”) (p < 0.001). There was no significant difference in the occurrence of osteomyelitis, necrotizing fasciitis, or septic arthritis. The investigated characteristics of IDU-BSI and non-IDU BSI cases included in WGS analysis, were comparable to those of corresponding subgroups in the total BJH BSI cohort (Supplementary Table 1).

Phylogenetic analysis of BSI isolates. BSI isolate genomes represented 26 different multilocus sequence types (ST), which included 10 ST groups (i.e., groups of single-locus or double-locus MLST variants sharing a common ancestor) containing at least five index isolates (Fig. 3 and Supplementary Table 3). The distribution of these ST groups between IDU-BSI and non-IDU BSI index isolates was not significantly different (p = 0.081, by Fisher exact test). The MRSA-associated ST groups ST8/1181/1750 and ST5/840, composed 67.1% and 25.2% of index isolates,
respectively. The most common non-MRSA ST groups, ST398/4163 and ST15/582, composed 14.0% and 10.5% of index isolates, respectively. The most common non-MRSA ST groups, ST398/4163 and ST672 IDU-BSI isolates represented 29.2% (45/154) of all IDU-BSI isolates, but 78.6% (11/14) and 100% (3/3) of ST398/4163 and ST672 IDU-BSI isolates, respectively.

### Identification and characterization of staphylococcal IDU-BSI transmission clusters.

We next determined the clonality of BSI isolates according to pairwise SNP distance. To standardize clonality analysis across our phylogenetically diverse cohort, we analyzed the 1782 genes shared by all genomes in our cohort (i.e., the 'core genome'). By compiling the pairwise core genome SNP distances between index isolates and between surveillance isolates obtained from single individuals, we determined that isolates likely resulted from clonal transmission if their core genomes differed by <15 SNPs (Fig. 4a). Using this conservative cutoff, we identified 20 transmission clusters containing 46 index isolates, with all but one isolate being from IDU-BSI cases (Fig. 4b). These represented 29.2% (45/154) of all IDU-BSI isolates, but 78.6% (11/14) and 100% (3/3) of ST398/4163 and ST672 IDU-BSI isolates, respectively.

We subsequently investigated demographic and clinical features that could support an epidemiologic link between cases within transmission clusters. Though patients in the entire cohort resided in 117 zip codes throughout Missouri and Illinois, 41/46 patients in transmission clusters shared or lived adjacent to the zip codes of other individuals within their cluster (Fig. 4c).

The median time between index dates of cases within a cluster was 28 days (mean 95 days, range 1–355 days), compared to a median of 293 days (mean 393, range 1–1394 days) between cases in differing clusters (p = <0.0001). Fourteen of the 20 clusters (70%) demonstrated HCV serocordance between ≥2 individuals (Fig. 4b). Lastly, compared to cohort individuals not assigned to transmission clusters, patients in clusters were more likely to be under the age of 50 (p < 0.001), to experience prolonged bacteremia (p < 0.001), to be co-infected with HIV or HCV (p < 0.001), and to use injection opioids (p < 0.001) (Fig. 4d). Together these data describe multiple geographically and temporally localized S. aureus transmission networks, which share objective epidemiologic and biologic markers of needle sharing (i.e., report of IDU, HIV/HCV serology), exclusively among PWID.

Secondary analysis reveals nonequivalent distribution of ST groups between cBSI and IDU-BSI isolates. As stated above, primary analysis revealed no difference in the distribution of ST groups between IDU-BSI to non-IDU BSI isolates. However, we noted a high incidence of CLABSI cases among the non-IDU cases in some lineages, such as ST398/4163 (Fig. 3). When non-IDU BSI isolates were sub-grouped into CLABSI and cBSI (n = 36 and n = 53, respectively), ST group distribution differed between cBSI and IDU-BSI cases (Supplementary Table 3, p = 0.035, Fisher exact test). When comparing observed to expected occurrence, underrepresented ST groups (i.e., ST groups observed at <75% of expected occurrence within a BSI subtype) were ST188, ST398/4163, ST45/256/536, ST97, and ST15/582 for cBSI, and ST30/39 and ST72 for IDU-BSI (Fig. 5a, b). These findings were consistent with differential selection of staphylococcal lineages according to BSI type.

### ST groups lacking multiple canonical virulence factors are underrepresented among cBSI isolates.

Since genomic repertoire influences the differential ability of microbes to cause different disease forms, we evaluated for the presence of genomic antibiotic resistance and canonical staphylococcal virulence determinants in our cohort (Supplementary Data 3). As expected, gene distribution was principally according to ST group lineage, and the presence of resistance determinants correlated with lab susceptibility testing (Fig. 3 and Fig. 5c, d). The distribution of MRSA isolates did not differ between cBSI (20/53), CLABSI (8/36) and IDU-BSI isolates (64/154) (p = 0.094, Fisher exact test).

Regarding the a priori selected list of fitness and virulence determinants (Supplementary Material 1), 100% of genomes contained the genes involved in iron acquisition and desiccation tolerance, and many genes involved in toxin production, immune evasion, and adhesion/biofilm formation were present in greater than 95% of isolates (Supplementary Table 1). For genes that occurred in less than 95% of total isolates, sequence types that were underrepresented in cBSI (Fig. 5c) lacked genes in all other sequences types (Fig. 5d, e), such as the staphylokinase and hyaluronidase genes sak and hysA, respectively. ST398/4163 isolates demonstrated the greatest paucity of canonical virulence factors (Fig. 5c).

### Discussion

To our knowledge, this is the first molecular epidemiological analysis comparing S. aureus isolates acquired from IDU-associated bloodstream infections (IDU-BSI) and conventional BSI (cBSI). Our case-control approach allowed us to identify characteristic features of IDU-BSI that are critical to consider in developing future investigations and interventions for this emerging disease.

### Table 1 Demographics of index isolates included for whole-genome sequencing.

<table>
<thead>
<tr>
<th>Demographics</th>
<th>Non-IDU patients, N = 91</th>
<th>PWID, N = 154</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (mean, SD)</td>
<td>58 ± 14</td>
<td>38 ± 10</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Female</td>
<td>34 (37.4%)</td>
<td>77 (50%)</td>
<td>0.045</td>
</tr>
<tr>
<td>Homeless</td>
<td>0 (0.0%)</td>
<td>12 (7.8%)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Discharged AMA</td>
<td>0 (0.0%)</td>
<td>43 (27.9%)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Substance use patterns</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Opioid use (fentanyl or heroin)</td>
<td>0 (0.0%)</td>
<td>142 (92.2%)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Methamphetamine use</td>
<td>0 (0.0%)</td>
<td>48 (31.1%)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Comorbidities</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hepatitis B virus infection</td>
<td>0 (0.0%)</td>
<td>6 (3.9%)</td>
<td>0.017</td>
</tr>
<tr>
<td>Hepatitis C virus infection</td>
<td>1 (1.1%)</td>
<td>95 (61.7%)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>HIV infection</td>
<td>0 (0.0%)</td>
<td>10 (6.5%)</td>
<td>0.002</td>
</tr>
<tr>
<td>Elixhauser comorbidities</td>
<td>9.0 (3.4)</td>
<td>8.6 (3.9)</td>
<td>0.484</td>
</tr>
<tr>
<td>(mean, SD)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Clinical syndromes caused by Isolate

| Infective endocarditis | 21 (23.0%) | 99 (64.3%) | <0.001 |
| Osteomyelitis | 17 (18.6%) | 33 (21.4%) | 0.576 |
| Septic arthritis | 10 (10.9%) | 27 (17.5%) | 0.149 |
| Necrotizing skin and soft tissue infection | 6 (6.6%) | 22 (14.2%) | 0.054 |
| Isolated bacteremia | 46 (50.5%) | 15 (9.7%) | <0.001 |
| S. aureus infection characteristics | | | |
| Hospital Day of S. aureus | 1 ± 0.2 | 1 ± 0.3 | 0.641 |
| Isolation for WGS (mean day, SE) | | | |
| Duration of bacteremia | 3 ± 0.3 | 4 ± 0.3 | 0.122 |
| Central line associated bacteremia | 38 (41.8%) | 22 (14.3%) | 0.082 |
| Outcomes | | | |
| 1 year Mortality | 27 (29.7%) | 22 (14.3%) | 0.082 |

AMA against medical advice, HIV human immunodeficiency virus, IDU injection drug use, PWID person who injects drugs, SD standard deviation, SE standard error, WGS whole-genome sequencing.
Firstly, WGS revealed that person-to-person transmission of multiple MRSA and methicillin susceptible *S. aureus* (MSSA) lineages are fueling the epidemic of invasive infections among PWID in Missouri and Southern Illinois. The transmission of *S. aureus* within IDU networks had been previously proposed, but evidence was limited to studies with low-resolution molecular methods or small cohort sizes. More recently, Packer et al. employed WGS to investigate MRSA colonization and disease isolates in PWID in Bristol, United Kingdom. They identified a single ST5 lineage predominated in their cohort, but their study was not designed to establish whether this phenomenon was exclusive to PWID, to identify transmission events, or to investigate dynamics of other lineages. By applying strict, empirically-derived clonality criteria comparable to those employed by other groups, we found strong evidence of clonal expansion of pathogenic *S. aureus* IDU-BSI isolates, and that epidemiologically-linked BSI transmission clusters almost exclusively occurred among PWID. In multiple clusters, ≥3 IDU-BSI isolates shared a common ancestor, reflecting that staphylococcal IDU-BSI cases can result from community outbreaks, similar to HCV and HIV outbreaks linked to needle sharing practices among PWID. Though we detected clonal expansion only among IDU-BSI cases, the relative underrepresentation (and, thus, reduced relative sequencing depth) of non-IDU BSI isolates in our genomic analysis could have decreased the likelihood of discovering clusters among non-IDU BSI cases, as well. However, the substantial difference in the proportion of matched IDU BSI and non-IDU BSI isolates belonging to transmission clusters (i.e., 29.2% [45/154] versus 1.1% [1/91]) firmly supports that clonal expansion is characteristic of the IDU-BSI epidemic.

Remarkably, some lineages (e.g., ST398/4163, ST672) were strongly associated with these IDU-BSI transmission clusters, suggesting that their pathogenicity depends heavily on IDU-associated risk factors. Many staphylococcal lineages underrepresented among cBSI cases lacked various canonical virulence factors associated with staphylococcal pathogenesis, suggesting a
selective pressure favoring “well-equipped” lineages during conventional pathogenesis. Others have observed similar enrichment of virulence factors in invasive versus colonizing \textit{S. aureus} isolates, with implicated virulence factors having a cumulative effect on invasiveness. Conversely, direct inoculation of bacteria into the bloodstream and bypass of early stages of cBSI (e.g., abscess formation, mucosal invasion, early immune evasion, etc.), permits disease by “less virulent” lineages. The comparable-to-lower mortality of invasive Staphylococcal disease among PWID relative to disease in non-PWID populations, despite the former’s association with prolonged bacteremia and often suboptimal antibiotic therapy, may largely be due to the fact that the former occurs in a younger patient population. However, it is conceivable that the portion of IDU-BSI associated with lineages with reduced superantigen production or toxin secretion (both implicated in \textit{S. aureus} sepsis) may also contribute to tempering IDU-associated infection severity. These implications, however, should not detract from the morbidity of IDU-BSI with “less virulent” lineages, as they were repeatedly implicated in BSI complications, including endocarditis, osteomyelitis, and septic arthritis in our cohort. Additional observational surveys of larger cohorts and preclinical research examining lineage-dependent pathogenesis, are required to confirm the impact of “less virulent” lineages in \textit{S. aureus} invasive disease.

A notable example of a “less virulent” lineage underrepresented among cBSI cases was the clonal complex 398 (CC398), which includes ST398 and ST4163 (Supplementary Table 3). CC398 isolates in our cohort characteristically demonstrated macrolide resistance putatively mediated by \textit{ermT} (Fig. 2), but they all lacked superantigens and many other canonical virulence factors. CC398 was first identified as a PVL-negative MRSA lineage in livestock in Europe, but was subsequently detected among MSSA colonizing diverse settings including community households and inmates in a jail holding tank. Rarely implicated in human disease, CC398 was identified in only one of 81 MSSA soft tissue infections in a prior study in our area. By contrast, CC398 was the most abundant exclusively MSSA lineage in our BSI cohort. CC398 was strongly associated with IDU transmission clusters, suggesting the lineage depends on intravenous inoculation in order to cause BSI. Indeed, though several non-IDU BSI cases were linked to CC398 in our cohort, they were almost exclusively CLABSI cases, where nosocomial bloodstream inoculation likely occurred. This demonstrates how preclinical models of IDU-BSI must account for the broader array of “less virulent” lineages that may be historically underrepresented among cBSI cases.

The dynamics mediating the high degree of relatedness among \textit{S. aureus} IDU-BSI isolates may serve as a target for curbing the IDU-BSI epidemic and merits close examination. One explanation is that the intimate interactions characteristic to some PWID populations, such as congregation in non-traditional housing, sharing of drug use paraphernalia and transactional sexual exchanges, which may predispose to clonal expansion through sequential asymptomatic carriage within a PWID network. This, in turn, increases the likelihood that two individuals in a network develop IDU-BSI due to related \textit{S. aureus} strains. However, this model presumes periods of genomic pool expansion during asymptomatic carriage punctuated by pool bottlenecking during colonization of new hosts and subsequent introduction into the bloodstream. It is unclear whether these population shifts would result in BSI cases with isolates displaying the high degree of genomic relatedness as what we observed between isolates subsequently obtained from a single BSI case and isolates within a single IDU-BSI transmission cluster (i.e., <15 core genome SNPs).
An alternative explanation is subsequent infections resulting from sharing of drug preparation or delivery equipment colonized with S. aureus. Indeed, Kasper et al. previously found that 14% of cookers/filters used for injection of controlled-release opioids in a community were contaminated with S. aureus which could have been injected intravenously during routine substance use. In either scenario, socioeconomic status factors such as homelessness and the low accessibility of needle exchanges in Missouri (which is a needle non-exchange state) may have contributed to the relatively high prevalence of transmitted isolates in the current study.

A third explanation for the clonal nature of IDU-BSI isolates would be bloodborne transmission. We confirmed IDU-BSI was associated with prolonged durations of bacteremia and frequent AMA discharges. So, although S. aureus invasive infections are generally considered to have acute and fulminant presentations, these infections conceivably take a more chronic course in a) individuals receiving intermittent, abbreviated antimicrobial therapy resulting in a “lower-grade” bacteremia or b) BSI patients with incompletely treated, secondary infections (e.g., endocarditis, osteomyelitis, occult abscesses, etc.) that can serve as chronic sources for recurrent bacteremia. Indeed, our cohort included an individual who had clonal S. aureus isolates obtained from blood specimens over a seven month period, without intervening negative blood cultures. Furthermore, seven transmission clusters contained IDU-BSI patients from whom clonal S. aureus isolates were obtained from at least two separate hospitalizations. Thus, IDU-BSI patients who defer medical attention could plausibly serve as reservoirs for bloodborne transmission through sharing...
of needles or drug preparation equipment. This provocative scenario requires further investigation, as it would shift priority towards clearance of bacteremia as a means to curb IDU-BSI propagation and inform the approach for assessing risk factors for \textit{S. aureus} BSI in this population.

Our investigation was limited to a single region where needle exchanges are prohibited, thus, limiting the generalizability of our findings. However, this represents a unique opportunity to examine the indirect impact of social IDU mitigation strategies practiced in other states on the BSI epidemic. Because the validity of contact tracing is complicated by recall limitations and socioeconomic instability often experienced by PWID, we were unable to validate our genomically-derived transmission clusters with retrospective contact tracing. However, our conservative SNP threshold for case clustering was deliberately chosen to minimize false-positive effects. Since our WGS cohort did not include isolates from non-endovascular host sites, we could not investigate whether asymptomatic colonization is a prerequisite for BSI among PWID. Further, the impact of different inpatient interventions such as ICU admission or cardiac surgery which could affect mortality differences between PWID and non-PWID groups was not assessed in this study. Lastly, our follow-up was limited to data present in the electronic medical record and out of hospital deaths which could contribute to mortality differences would not be captured in this study. Despite these limitations, the comparative design of our study combined with robust clinical and genomic data provides valuable insight into the epidemiological and pathophysiological impact of the propagation of multiple \textit{S. aureus} lineages among PWID. 

Conclusion

We identified clonal expansion of multiple \textit{S. aureus} lineages among IDU-BSI isolates, but not non-IDU BSI isolates, in a community with limited access to needle exchange facilities. In the setting of expanding numbers of staphylococcal IDU-BSI cases consideration should be given to treating IDU-associated invasive staphylococcal infections as a communicable disease.

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

The data that support the findings of this study are available within the paper and its supplementary information files. The source data underlying Figs. 3, 4, & 5 are derived from whole-genome sequencing of the strains in this project. Whole-genome sequence read files are uploaded to NCBI under BioProjects PRJNA694991 and PRJNA695316. Source metadata for Fig. 3 & 5 can be accessed as Supplementary Data 2. Source Data for Fig. 4a can be accessed as Supplementary Data 3. The remaining source data on individual patient outcomes and locations used in Figs. 2 and 4b–e cannot be provided because it contains elements of protected health information and the ethical approval does not cover placing individual patient level data into publicly open repositories. Relevant portions of those data can be accessed from the authors upon relevant ethical approval by contacting the corresponding author on reasonable request.

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Author contributions
L.R.M. and J.J.C. conceived and designed the study. L.R.M., M.J.D., and J.A.W. performed all chart review. C.A.B. and M.A.W. collected samples. L.R.M., J.J.C., J.A.W., M.A.W., E.M.R., and S.S. performed specimen handling, genome sequencing, data management and statistical analysis. L.R.M., J.A.W., J.J.C. prepared figures and tables. L.R.M. and J.J.C. wrote the first draft of the manuscript. C.A.B., J.P.H., M.J.D., and G.D. analyzed and discussed data and critically revised the manuscript. All authors agreed to final submission.

Competing interests
The authors declare no competing interests.

Additional information
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