Diverse voltage-sensitive dyes modulate GABAA receptor function

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Voltage-sensitive dyes are important tools for assessing network and single-cell excitability, but an untested premise in most cases is that the dyes do not interfere with the parameters (membrane potential, excitability) that they are designed to measure. We found that popular members of several different families of voltage-sensitive dyes modulate GABAA receptor with maximum efficacy and potency similar to clinically used GABAA receptor modulators. Di-4-ANEPPS and DiBAC4(3) potentiated GABA function with micromolar and high nanomolar potency, respectively, and yielded strong maximum effects similar to barbiturates and neurosteroids. Never blue oxonols had biphasic effects on GABAA receptor function at nanomolar and micromolar concentrations, with maximum potentiation comparable to that of saturating benzodiazepine effects. ANNINE-6 and ANNINE-6plus had no detectable effect on GABAA receptor function. Even dyes with no activity on GABA receptors at baseline induced photodynamic enhancement of GABA receptors. The basal effects of dyes were sufficient to prolong IPSCs and to dampen network activity in multielectrode array recordings. Therefore, the dual effects of voltage-sensitive dyes on GABAergic inhibition require caution in dye use for studies of excitability and network activity.

Materials and Methods

Hippocampal cultures. Primary cultures were prepared from postnatal day 0–3 rat pups as previously described (Shu et al., 2004). Rat pups were anesthetized with isoflurane and the hippocampus was cut into 500-μm thick slices. The slices were digested with 1 mg/ml papain in oxygenated Leibovitz L-15 medium (Invitrogen) and mechanically triturated in Leibovitz L-15 medium (Invitrogen) containing 5% horse serum, 5% fetal calf serum, 17 mM d-glucose, 400 μM glutamine, 50 U/ml penicillin, and 50 μg/ml streptomycin. Cells were plated in modified Eagle’s medium at a density of ~650 cells/mm² on collagen-coated tissue culture dishes ( Falcon). Cultures were maintained at 37°C in a humidified incubator with 5% CO2/95% air. Glial proliferation was halted 3–4 d after plating with 6.7 μM cytosine arabinoside. At 4–5 d after plating, half the culture medium was replaced with Neurobasal medium plus B27 supplement (both from Invitrogen).
Single-cell electrophysiology. Whole-cell recordings were performed on hippocampal neuron cultures 8–13 d following plating with an Axopatch 200B amplifier (Molecular Devices). Cells were transferred from culture medium to an extracellular recording solution containing the following (in mM): 130 cesium methanesulfonate, 4 NaCl, 5 EGTA, 0.5 CaCl2, and 10 HEPES at pH 7.25. Pipette tips were filled with an internal solution containing the following (in mM): 130 cesium methanesulfonate, 4 NaCl, 5 EGTA, 0.5 CaCl2, and 10 HEPES at pH 7.25. For synaptic experiments, cesium methanesulfonate was replaced with potassium chloride, and with cesium chloride for photopotentiation experiments. When filled with solution, pipette tip resistance was 4–6 MΩ. Cells were clamped at −60 or −70 mV unless otherwise indicated. Access resistance was 8–20 MΩ and was not compensated for exogenous applications or for miniature IPSC recordings, where current amplitudes were small. For evoked autaptic IPSCs, access resistance was compensated 80–90%. A voltage pulse to 0 mV (1.2 ms) triggered a presynaptic action potential that elicited the IPSC (Menernick et al., 1995). Drug applications were made with a multibarrel, gravity-flow local perfusion system. The estimated solution exchange times were 120 ± 14 ms (10–90% rise), measured by the change in junction currents at the tip of an open patch pipette. Whole-cell recordings were performed at room temperature.

Network recordings. Multielectrode arrays (MEAs) were coated with poly-D-lysine and laminin per the manufacturer’s instructions and dispersed cultures were grown as described above. At day in vitro (DIV) 7 and DIV10, one-third of the medium was removed and replaced with fresh Neurobasal supplemented with B27 and glutamine. Recordings were made with the MEA-60 recording system (MultiChannel Systems) with the headstage in an incubator set at 37°C and equilibrated with 5% CO2 in room air with no additional humidity. The lower temperature was necessary because the electronics in the headstage generate ≈7°C of excess heat. The MEA itself rests on a heating plate inside the headstage that was heated so the cultures were maintained at 37°C. To allow extended recordings in the dry incubator, cultures were covered with a semipermeable membrane that allows diffusion of oxygen and carbon dioxide but not water (Potter and DeMarse, 2001). Di-4-ANEPPS was added directly from stock solutions to the culture medium from a sister culture under sterile conditions. The medium in the MEA was replaced with new medium and allowed to re-equilibrate for ~5 min before recording. Data were amplified ×1100 and sampled at 5 kHz. Spikes were detected by threshold crossing of high-pass filtered data. The threshold was set individually for each contact at 5 SDs above the average root mean squared noise level. Baseline data were recorded immediately before dye treatment, after which the medium was again replaced and a final dataset collected. All datasets were 2 h long. Activity was quantified using the array wide spike detection rate (ASDR) (Wagenaar et al., 2006), defined as the number of spikes detected in all contacts of the MEA during each second of recording. The average ASDR for each 2 h dataset was used as a summary measure of activity. Activity during dye exposure was compared with the average of the baseline and wash activity levels using a paired t test with p < 0.05 considered significant.

Oocyte expression studies. Stage V–VI oocytes from sexually mature female Xenopus laevis (Xenopus One) were harvested under 0.1% 3-aminobenzoic acid ethyl ester anesthesia, according to protocols approved by the Washington University Animal Studies Committee. The follicular layer was removed by shaking for 20 min at 37°C in collagenase (2 mg/ml) dissolved in calcium-free solution containing the following (in mM): 96 NaCl, 2 KCl, 1 MgCl2, and 5 HEPES at pH 7.4. Capped mRNA, encoding rat GABA γ receptor α1, B2, and γ2L subunits, was transcribed in vitro using the mMESSAGE mMachine kit (Ambion) from linearized pBluescript vectors containing receptor-coding regions. Subunit transcripts were injected in equal parts (20–40 ng of total RNA) 8–24 h following defolliculation. Oocytes were incubated up to 5 d at 18°C in ND96 medium containing the following (in mM): 96 NaCl, 1 KCl, 1 MgCl2, 2 CaCl2, and 10 HEPES at pH 7.4, supplemented with pyruvate (5 mM), penicillin (100 U/ml), streptomycin (100 μg/ml), and gentamycin (50 μg/ml).

Oocyte electrophysiology. Oocytes were voltage clamped at −70 mV with a two-electrode voltage-clamp amplifier (Warner Instruments) 2–5 d following RNA injection. The extracellular recording solution was supplemented ND96 medium. Intracellular recording pipettes contained 3 M KCl and had open-tip resistances of 1 MΩ. Drugs were applied from a common tip via a gravity-driven multibarrel delivery system. Drugs were coapplied with no preaplication period. Cells were voltage clamped at −70 mV for all experiments and the peak current was measured for quantification of current amplitudes.

Data analysis and statistical procedures. Data acquisition and analysis of single-cell electrophysiology from oocytes and hippocampal neurons were performed with pCLAMP 9.0 software (Molecular Devices), except for analysis of spontaneous miniature IPSCs, which was performed with MiniAnalysis (SynaptoSoft). Data plotting and curve fitting were performed with SigmaPlot (Systat Software). Curve fitting was performed on potentiation values calculated as (M − G)/G, where M is the response in the presence of GABA plus modulator and G is the response to GABA alone. Empirical fits to concentration–response relationships were achieved using a least-squares minimization to the Hill equation: Xmax [Y0/(EC50 + X0)], where Xmax is the maximum potentiation, Y0 is the Hill coefficient, EC50 is the concentration of modulator producing 50% of maximum potentiation, and X is the test modulator concentration. LogP calculations (logarithm of the octanol:water partition coefficient) were performed using an online calculator that simultaneously calculates LogP estimates from nine independent algorithms (ALOGPS 2.1; http://www.vcclab.org/lab/alogsps/). Multielectrode array data were analyzed and plotted using Igor Pro (WaveMetrics). Data are presented as mean ± SE. Statistical differences were determined using Student’s two-tailed t test.

Drugs, chemicals, and other materials. Compounds were obtained from Sigma with the following exceptions: Di-4-ANEPPS, Di-8-ANEPPS, and DiSBAC4(3) were from Invitrogen; DiBAC4(5), DiSBAC2(3), DiSBAC4(2), Oxonol V, and Oxonol VI were from AnaSpec; blue oxonol (RH1691, RH1692, and RH1838) were from Optical Imaging; and ANNINE-6 and ANNINE-6plus were from Sensitive Dyes. Dyes were prepared as stock solutions in dimethylsulphoxide (DMSO). Final DMSO concentration was <0.1%. At double this concentration (0.2% DMSO), we observed a small inhibition of GABAα receptor currents (5 ± 2% < p < 0.05, N = 8) but never potentiation, so DMSO effects cannot explain VSD potentiation.

Results

In initial experiments to test whether voltage-sensitive membrane probes alter GABAα receptor function, we expressed a rat α1β2γ2L GABAα subunit complex in Xenopus oocytes and assessed modulation of GABA current amplitudes by simultaneous coapplication of the oxonol VSD DiBAC4(3). For reference, structures of the dyes used in this study are shown in supplemental Figures 1 and 2 (available at www.jneurosci.org as supplemental material). We found strong maximum potentiation (~15-fold) and an EC50 value of 0.36 μM under these conditions (Fig. 1, A, C). Importantly, this modulation was observed in the concentration range of oxonol compounds used for imaging studies. As is observed for high concentrations of many allosteric modulators of GABAα receptors, 3 μM DiBAC4(3) activated small currents in the absence of GABA (Fig. 1A, inset) (63 ± 5% of the response to 2 μM GABA, N = 4 oocytes), indicating that DiBAC4(3) weakly gates the receptor in the absence of agonist in addition to positive modulation of receptor function in the presence of agonist.

DiBAC4(3) is a member of an oxonol family of older, slower VSDs and has a barbituric acid structure that could partly ac-
GABA receptor modulation (trol oocytes that were not injected with GABA receptor subunits, also robustly modulated GABAA receptor currents when simul-
tations were again in the range used in imaging studies (EC50 M GABA alone and GABA with simultaneously coapplied 1, 3, and 10 μM Di-4-ANEPPS. C. Summary concentration–response relationships (N = 4 oocytes per data point). The solid lines are fits of the average data points to the Hill equation and yield an estimated EC50 for DiBAC4(3) of 0.36 μM. The estimated EC50 for Di-4-ANEPPS was 4.1 μM. D, Another ANEP family member, Di-8-ANEPPS, failed to potentiate GABA currents.

Figure 1. Members of two structurally distinct families of voltage-sensitive dyes strongly potentiate GABA<sub>α</sub> receptor currents in Xenopus oocytes expressing α1β2γ2L subunit combinations. A, Representative traces of responses to GABA alone (2 μM) and to simultaneous coapplication of GABA with increasing concentrations (0.03–3 μM) of the oxonol dye DiBAC4(3). Horizontal bar above traces gives duration of exposure. The inset shows the structurally different from the oxonols (supplemental Figs. 1 and 2, available at www.jneurosci.org as supplemental material). Di-4-ANEPPS also robustly modulated GABA<sub>α</sub> receptor currents when simultane-
ously coapplied with GABA (Fig. 1B, C). The concentration requirement was higher than that for DiBAC4(3), but concentrations were again in the range used in imaging studies (EC<sub>50</sub> = 4.1 μM). Similar to DiBAC4(3), maximum potentiation was very strong (Fig. 1B, C), akin to that achieved by neurosteroids and barbiturates under similar conditions. No responses to agonist or to dyes were observed at the highest concentrations tested (3 and 10 μM for DiBAC4(3) and for Di-4-ANEPPS respectively) in control oocytes that were not injected with GABA receptor subunits, confirming that the currents observed indeed resulted from GABA receptor modulation (N = 4 oocytes, data not shown).

For neurosteroid actions at GABA<sub>α</sub> receptors, steroid lipophili-
city plays a strong role in potency, at least for a range of structural steroid analogues (Chisari et al., 2009). To determine whether this rule applies to VSD modulation, we compared actions of Di-4-ANEPPS (calculated logP = 4.2 ± 0.3) with Di-8-ANEPPS (logP = 7.6 ± 0.3, see Materials and Methods). The higher logP of Di-8-ANEPPS results from a longer hydrophobic tail (supplemental Fig. 2, available at www.jneurosci.org as supplemental material). In contrast to neurosteroids, we found that the more hydrophobic Di-8-ANEPPS had no detectable effect on GABA<sub>α</sub> receptor function up to 10 μM (Fig. 1D). Currents in the presence of 10 μM Di-8-ANEPPS were 125 ± 13% of GABA currents in the absence of dye (N = 12 oocytes, p > 0.05). Therefore, the ANEP dye family also exhibits structural variability for GABA<sub>α</sub> receptor modulation, and results suggest that with care in choice of dye, basal GABA receptor effects may be avoided.

DiBAC4(3) also modulates certain BK potassium channel subunits, and this modulation has a specific structure-activity relationship (Morimoto et al., 2007). We exploited the structural diversity of the oxonol family, of which DiBAC4(3) is a member, to obtain structure-activity relationship data. We included in this screen three newer members of the oxonol family, so-called blue oxonols, which are rapid sensors of cellular activity (Shoham et al., 1999; Spors and Grinvald, 2002; Petersen et al., 2003). We found that each of the blue oxonols had a concentration-dependent biphasic effect on GABAA receptor function when evaluated at 0.3 μM and at 3 μM (Fig. 2A). The potentiation observed at 0.3 μM was similar to a saturating concentration of lorazepam (Fig. 2B) (lorazepam potentiation was 272 ± 13% of control, N = 3 oocytes). A summary of single-concentration (0.3...
3% and 80 cytes. Again, at modest concentrations, both DiBAC4(3) (0.2–0.5 μM) screened is shown in Figure 2C and demonstrates variability among family members in their ability to modulate GABA<sub>A</sub> receptor function. We used a concentration that was near the EC<sub>50</sub> for DiBAC4(3) so that we could detect stronger and weaker activity compared with DiBAC4(3). Interestingly, DiBAC4(3) was much stronger than any of the other family members at this subsaturating concentration (Fig. 2C). We raised the concentration of oxonol compounds in our screen tenfold, to 3 μM. Normalized current values did not increase dramatically for any of the oxonol compounds (range 0.97–3.61). Only the blue oxonols showed evidence of biphasic modulation (Fig. 2A). Overall, the failure of higher concentrations to increase potentiation suggests that the modest potentiation values reflect low efficacy of modulation rather than low potency. In general, the structure-activity relationship was distinct from that observed for BK channel subunits (Morimoto et al., 2007) (see Discussion).

We also obtained limited quantities of the newer fast indicators ANNINE-6 and ANNINE-6plus dyes (Fromherz et al., 2008). When screened at 0.3 and at 3 μM against responses to 2 μM GABA, we failed to observe any significant potentiation of currents. GABA responses were in fact diminished slightly, but not in a clearly concentration-dependent manner (ANNINE-6: 89 ± 3% and 83 ± 2% depression at 0.3 and 3 μM respectively, N = 3 oocytes; ANNINE-6plus: 81 ± 3% and 80 ± 1% depression at 0.3 and 3 μM respectively).

In subsequent studies, we examined whether VSDs also modulate GABA receptors in mammalian cells. Based on results of the oocyte screening studies, we tested DiBAC4(3) and Di-4-ANEPPS as representative positive modulators and Oxonol VI and Di-8-ANEPPS as representative weak potentiators in cultured hippocampal neurons. Results paralleled those from oocytes. Again, at modest concentrations, both DiBAC4(3) (0.2–0.5 μM) and Di-4-ANEPPS (3–10 μM) profoundly potentiated responses to low GABA concentration (Fig. 3A, C), while neither 1 μM Oxonol VI (data not shown) nor 10 μM Di-8-ANEPPS (Fig. 3B) had any reliable potentiating effect in any cells tested (N = 5 and 3, respectively). Responses to DiBAC4(3) and to Di-4-ANEPPS in the presence of GABA were sensitive to application of 100 μM picrotoxin (Fig. 3A, C). Therefore, dye-induced potentiation clearly resulted from interaction of dyes with GABA<sub>A</sub> receptors.

Some VSDs, particularly slow indicators, indicate membrane potential in part by translocation within the membrane in response to changes in the transmembrane voltage (Waggoner and Grinvald, 1977; González and Tsien, 1995). This movement alters position of the environment-sensitive fluorophore and thus fluorescence, and allows use of these fluorophores in FRET studies. We wondered whether voltage-sensitive movement of the fluorophore might register as voltage-sensitive potentiation of GABA currents if intramembrane movement is important to access a VSD receptor site. However, neither DiBAC4(3) (Fig. 3D) nor Di-4-ANEPPS potentiation exhibited strong voltage dependence. Potentiation at −60 mV and at +40 mV was 173 ± 44% and 141 ± 29% respectively for DiBAC4(3) (N = 4 cells). For Di-4-ANEPPS potentiation was 437 ± 81% and 350 ± 30% at the two potentials (N = 8 cells). Furthermore, Di-8-ANEPPS, which was inert at −60 mV, was also inert at +40 mV (N = 3).

Di-8-ANEPPS could exhibit dramatically weaker effects on GABA<sub>A</sub> receptors than Di-4-ANEPPS because it fails to fulfill pharmacophore requirements for binding to a site on the GABA<sub>A</sub> receptor. Alternatively or additionally, Di-8-ANEPPS could fail to reach the binding site because its cellular accumulation and access to a putative GABA receptor site could differ from Di-4-ANEPPS. For instance, the rate of cellular accumulation of the two dyes has been reported to differ in cardiac cells (Rohr and Salzberg, 1994), and this could alter access to transmembrane or cytoplasmic targets on the receptor. As a simple test of whether slow accumulation could participate in the weak Di-8-ANEPPS activity on GABA<sub>A</sub> receptors, we examined responses of oocytes incubated for 10 min in the presence of 10 μM Di-8-ANEPPS. We observed only a minimal increase in Di-8-ANEPPS effect with long incubation. Currents after soaking were 23 ± 4% larger than those observed with acute Di-8-ANEPPS application (N = 4 oocytes).

To determine whether cellular retention patterns in hippocampal neurons differed between Di-4-ANEPPS and Di-8-ANEPPS, we imaged the time course of dye accumulation and maximum fluorescence in hippocampal neurons. We found that the two dyes differed dramatically in their cellular accumulation (Fig. 4). Di-4-ANEPPS exhibited bright fluorescence, initially restricted to the plasma membrane, but which over a course of minutes became strongly internalized within neurons (Fig. 4A). In contrast, 2 min of incubation in 10 μM Di-8-ANEPPS failed to produce cellular fluorescence levels comparable to Di-4-ANEPPS and did not produce fluorescence internalization (Fig. 4A, B). This pattern of comparatively weak fluorescence persisted with incubation times of up to 15 min. Di-4-ANEPPS perimembrane fluorescence at 15 min was >5-fold higher than Di-8-ANEPPS.
The difference in maximum fluorescence of the dyes did not result from a difference in the inherent fluorescence of the two dyes because when dissolved in propanol at 50 μM and imaged in solution in the absence of cells, the dyes exhibited nearly identical fluorescence (Fig. 4B). In summary, we cannot exclude the possibility that cellular accumulation differences that result in differential access to the receptor might participate in the differences in GABA<sub>α</sub> receptor activity between the two tested ANEP dyes.

We recently showed that GABA receptors are subject to photodynamic effects in the presence of fluorophores that gain proximity to the GABA receptor, with fluorescent neurosteroid analogues serving as particularly potent photosensitizers (Eisenman et al., 2007). To test whether VSDs also elicit photodynamic analogues serving as particularly potent photosensitizers (Eisenman et al., 2007). The photodynamic effect, like the baseline effect, was not detectably sensitive to voltage (Fig. 5A). We also tested Di-8-ANEPPS, a VSD with no baseline activity at GABA<sub>α</sub> receptors. At 10 μM, we confirmed that Di-8-ANEPPS had little or no baseline activity (Fig. 5C) (46 ± 17% potentiation, N = 11). However, upon 480 nm light excitation, GABA responses were potentiated 483 ± 168% (N = 11) over baseline GABA responses. ANNINE-6plus (3 μM) dyes, in the absence of light excitation, on GABA synaptic function. Figure 6, A and B, shows the effect of RH1691 (0.3 μM) and Di-4-ANEPPS (5 μM) on GABA-mediated evoked autaptic responses (IPSCs) from hippocampal neurons in culture. Di-4-ANEPPS elicited reversible increases in the holding current (−69 ± 30 pA, N = 5) that are not evident in Figure 6B, where baseline currents were subtracted. This likely resulted from direct activation of receptors in the absence of GABA. This change in holding current was not evident with 0.3 μM RH1691 (8 ± 7 pA, N = 5). The major change produced by both dyes on synaptic events was a prolongation of IPSCs (Fig. 6A,B), similar to that observed with other positive modulators of GABA<sub>α</sub> receptor activity (Hemmings et al., 2005). On average peak IPSCs were changed by −2 ± 8% and 28 ± 13% by 0.3 μM RH1691 and 5 μM Di-4-ANEPPS respectively. IPSC decays were prolonged by 32 ± 5% and by 173 ± 30% respectively (p < 0.05, N = 5 cells for both compounds).

The prolongation of IPSCs, with weak effects on peak amplitude, suggests a primarily postsynaptic locus of dye effect, with little effect on the presynaptic voltage-gated channels responsible for action potential propagation and Ca<sup>2+</sup> influx. As a further test of a postsynaptic locus, we also examined effects of 3 μM Di-4-ANEPPS on spontaneous miniature IPSCs, recorded in the presence of 0.5 μM tetrodotoxin (Fig. 6C). Dye again produced a substantial change in holding current (data not shown) and increased membrane noise (Fig. 6C1,C2), consistent with direct gating of receptors by dye. Miniature IPSCs were prolonged by dye in a reversible manner (Fig. 6C4) (96 ± 20% increase in decay time, N = 5). In addition we observed a significant increase in peak amplitude of miniature IPSCs (50 ± 16% of control, N = 5).
cells) and an apparent increase in frequency of miniature IPSCs (152 ± 58% of control). Based on the significant miniature IPSC frequency increase, we cannot exclude a presynaptic effect of dye, although this apparent change could in part result from increased membrane noise in the presence of dye, leading to an increase in detection of falsely positive synaptic events (Fig. 6C).

It was clear that effects of Di-4-ANEPPS were largely reversible upon washout. This reversibility may be somewhat unexpected, since many experiments load cells or tissue with Di-4-ANEPPS, washout-free dye, and perform subsequent imaging of retained dye for many minutes (Yuste et al., 1997; Wachowiak and Cohen, 1999; Arata and Ito, 2004). To determine whether this reversibility was paralleled by cellular fluorescence, we imaged wash-on and washout of Di-4-ANEPPS (Fig. 7). Figure 7A shows that when applied for 40 s, Di-4-ANEPPS was localized mainly to the plasma membrane (Fig. 4A), and was largely reversible over the subsequent 90 s of wash (Fig. 7A,C). This is consistent with washout times observed in our electrophysiology studies. In contrast, with longer incubations of 15 min (Fig. 7B), dye was internalized, reached brighter fluorescence, and failed to readily reverse over a subsequent 90 s wash (Fig. 7B,C). Therefore, reversibility of GABA actions is expected of brief incubations, consistent with our electrophysiology protocols. Over longer incubations, the impact of cell internalization and slow washout times might be expected to influence effects.

To test the impact of longer dye incubation on synaptic function, we assessed the currents directly gated by Di-4-ANEPPS after prolonged incubations (15 min) in 10 μM Di-4-ANEPPS. Evaluation was performed on sibling cultures after vehicle con-
control incubation, in the continued presence of dye, or following free dye removal, to simulate the way Di-4-ANEPPS is often used in slice experiments (incubation followed by imaging in dye-free solutions). Glutamate blockers were present in all recording solutions, and cells were clamped at −70 mV using a CsCl-filled pipette. After prolonged dye exposure, we recorded from neurons and locally perfused 100 μM picrotoxin to evaluate the standing GABA_A receptor-gated current under the various conditions. In control cells, there was a barely detectable picrotoxin-sensitive standing current (3.4 ± 1.0 pA, N = 8 cells). Prolonged incubation with recordings in the continued presence of dye yielded larger picrotoxin-sensitive currents (46.3 ± 9.7 pA, N = 7). After prolonged Di-4-ANEPPS incubation with recordings performed in the absence of dye, we recorded intermediate sized GABA_A receptor currents (10.0 ± 1.4 pA, N = 7; p < 0.01 relative to control and relative to the persisting incubation condition). These results suggest that despite removal of free dye, some effects on GABA receptors linger as a result of retained dye, although these effects are not as strong as the acute dye effects.

To test directly the impact of VSD actions at GABA receptors on network activity, we examined the effects of Di-4-ANEPPS on network spiking activity recorded in dissociated hippocampal cultures plated on multielectrode arrays. Overall spontaneous activity in the cultures was strongly inhibited when the medium was switched to medium containing 10 μM Di-4-ANEPPS (Fig. 8). Note that no light stimulation of the fluorophore was used in these studies. Cells were maintained in a dark incubator for the entire time period, in the continued presence of dye, or following wash of free dye (Fig. 8). Similar array-wide spike rates from 10 min segments obtained from another culture subjected to control medium exchanges with no dye.

**Discussion**

We show strong, positive modulation of GABA_A receptors by several structurally diverse VSDs at concentrations relevant to their use as voltage indicators. Effects of several VSDs are similar to neurosteroids and to barbiturates, with 15–20-fold maximum potentiation at low GABA concentrations. DiBAC4(3) has potency similar to neurosteroids and much greater than barbiturates. Di-4-ANEPPS exhibits potency comparable to or greater than barbiturates. Although drawbacks of VSDs have been noted...
photodynamic effects on GABA<sub>A</sub> receptors may be evident at the GABA receptor by binding in the absence of light stimulation. However, because of selective proximity gained by some VSDs to inhibition at 3<sup>rd</sup> division or permeation, contributes to binding-site access (Akk et al., 2009). In this case, structural features affecting the ligand’s interaction with the membrane could affect its ability to access the receptor site, in addition to any true pharmacophore effects (Makriyannis et al., 2005; Chisari et al., 2009). Low concentrations of DiBAC<sub>4</sub>(3) and Di-4-ANEPPS exhibited very slow onset kinetics relative to GABA in both oocytes (Fig. 1) and hippocampal neurons (Fig. 2). Similarly slow kinetics are also hallmarks of neurosteroid actions (Shu et al., 2004) and could indicate that a process not limited by aqueous diffusion, such as membrane partitioning or permeation, contributes to binding-site access (Akk et al., 2005; Chisari et al., 2009). Complicating matters further is our observation that blue oxonols exhibit evidence for a biphasic effect on GABA<sub>A</sub> receptor function, with mixed potentiation and inhibition at 3 µM (Fig. 2A). Pending a full understanding of the number of distinct sites involved in VSD modulation of GABA<sub>A</sub> receptors and how the VSDs access these sites, investigators will have to evaluate potential confounding effects of GABA receptor interactions on a case-by-case basis.

We also found that at least for our test compound Di-4-ANEPPS, length of incubation affected the compartmentalization of dye (Fig. 7), the ability to effectively remove cellularly retained dye (Fig. 7), and the effects on neuronal network activity (Fig. 8). The basis for the rebound of activity during prolonged Di-4-ANEPPS incubation is unclear. However, the rebound could be associated with the internalization of dye observed in Figure 7 with prolonged incubation. Alternatively, because we observed that directly gated picrotoxin-sensitive currents persisted with prolonged dye incubation, it is possible that the rebound in network activity was associated with compensatory synaptic or excitability changes.

Fluorescent dyes photodynamically modulate voltage-gated channels (Oxford et al., 1977; Duprat et al., 1995; Antic et al., 1999), giving rise to well known effects on action potential shape (Antic et al., 1999). These effects, like photodynamic effects on GABA<sub>A</sub> receptors, can be minimized by limiting light exposure. However, because of selective proximity gained by some VSDs to the GABA receptor by binding in the absence of light stimulation, photodynamic effects on GABA<sub>A</sub> receptors may be evident at light levels below those that elicit effects on other ion channels. For instance, fluorescent neurosteroid analogues that bind the GABA<sub>A</sub> receptor exhibit more potent photodynamic GABA<sub>A</sub> effects than analogues that do not bind the receptor (Eisenman et al., 2007; Shu et al., 2009). For reasons that are unclear, ANNINE-6plus, a dye that proved inert at baseline, was particularly efficient at generating photodynamic effects on GABA<sub>A</sub> receptor function in our experiments (Fig. 5D).

The light-independent effects of dyes described here are more difficult to circumvent. Similar to recently reported effects on membrane capacitance of protein voltage indicators (Akemann et al., 2009), the effects reported here on GABA<sub>A</sub> receptors are likely to be ubiquitous. Virtually all neurons express GABA<sub>A</sub> receptors, and we found that all hippocampal neurons tested were sensitive to GABA<sub>A</sub> modulation by oxonol and by Di-4-ANEPPS. Furthermore, 10 µM Di-4-ANEPPS significantly depressed network activity. Several of the oxonol compounds had weaker effects on GABA receptors than either DiBAC<sub>4</sub>(3) or Di-4-ANEPPS. However, use of these compounds is risky because the efficacy of these compounds approached that of benzodiazepines, another class of high potency, weak efficacy potentiatators. Because benzodiazepines have well described effects on neuronal activity and excitability, it seems likely that weak oxonols will have similar effects. Clearly, careful dye choice is critical to avoid inadvertent VSD effects on the GABA system and neuronal excitability.

In summary, we have demonstrated dual light-independent and light-dependent actions of voltage-sensitive dyes on GABA<sub>A</sub> receptor function. These effects are of particular concern because of the ubiquity of GABA signaling throughout the nervous system and the importance of GABA to cellular and network excitability. Our results suggest caution in choice of dye and in experimental design to avoid inadvertent interference with neuronal activity.

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