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Skin-Derived TSLP Triggers Progression from Epidermal-Barrier Defects to Asthma

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Abstract

Asthma is a common allergic lung disease frequently affecting individuals with a prior history of eczema/atopic dermatitis (AD); however, the mechanism underlying the progression from AD to asthma (the so-called “atopic march”) is unclear. Here we show that, like humans with AD, mice with skin-barrier defects develop AD-like skin inflammation and are susceptible to allergic asthma. Furthermore, we show that thymic stromal lymphopoietin (TSLP), overexpressed by skin keratinocytes, is the systemic driver of this bronchial hyper-responsiveness. As an AD-like model, we used mice with keratinocyte-specific deletion of RBP-j that sustained high systemic levels of TSLP. Antigen-induced allergic challenge to the lung airways of RBP-j−deficient animals resulted in a severe asthmatic phenotype not seen in similarly treated wild-type littersmates. Elimination of TSLP signaling in these animals blocked the atopic march, demonstrating that high serum TSLP levels were required to sensitize the lung to allergic inflammation. Furthermore, we analyzed outbred K14-TSLPtg mice that maintained high systemic levels of TSLP without developing any skin pathology. Importantly, epidermal-derived TSLP was sufficient to trigger the atopic march, sensitizing the lung airways to inhaled allergens in the absence of epicutaneous sensitization. Based on these findings, we propose that in addition to early treatment of the primary skin-barrier defects, selective inhibition of systemic TSLP may be the key to blocking the development of asthma in AD patients.

Introduction

Allergic asthma is a chronic lung disease characterized by T-helper2 (Th2)–mediated inflammation and airway obstruction in response to allergen exposure [1]. Asthma is an increasingly common disorder affecting more than 300 million individuals around the world [2]. Atopic dermatitis (AD) is another prevalent allergic disease, and 17% of children in the United States suffer from this disorder [3]. In comparison to its 4–8% prevalence in the general population, asthma develops in up to 70% of patients with history of severe AD, a phenomenon referred to as “atopic march” [3]. It may be possible to prevent asthma in these at-risk individuals by early diagnosis of AD and blockage of the atopic march. To achieve this, it is critical that we understand the mechanism triggering the development of asthma in AD patients. A survey of the literature has identified several possible mechanisms underlying the clinical link between AD and asthma. These include: (a) a systemic immune system disorder leading to excessive Th2 response at epithelial surfaces exposed to allergens [4], (b) a barrier defect shared by both skin and lung epithelia that leads to overstimulation of the immune cells by invading allergens [5], or (c) systemic consequences of a skin-specific barrier defect causing immune cells to mount an allergic inflammation at any allergen-exposed epithelial surface [6]. Epidemiological data supporting the third hypothesis include the observation that AD tends to be the first manifestation of the atopic march [3]. Another supportive observation relates to filaggrin, a skin cornified envelope protein that is absent from the lung epithelia [7,8]. Although controversial [9], it seems that AD patients with filaggrin loss-of-function mutations exhibit increased incidence of asthma [10–12]. This is consistent with the third hypothesis, that loss of an epidermal-specific barrier protein can trigger systemic atopy in humans. Mechanistically, it is suspected that epicutaneous sensitization with allergens underlies the development of airway hyperreactivity in mice [13] and in humans with filaggrin mutations [10,11]. However, it is unclear whether intrinsic skin-barrier defects can trigger asthma in the absence of any epicutaneous sensitization. If epicutaneous exposure is not required, it will imply that systemic factor(s) produced by AD skin may be involved in sensitizing the bronchial epithelia. Such factors will be important as therapeutic targets in preventing asthma.

To address this question, we studied mice lacking Notch signaling in the skin. Skin keratinocytes are organized in highly interconnected basal, spinous, granular, and cornified layers, forming an elaborate barrier protecting the organism from the outside environment. One of the major molecular regulators of this structure is Notch signaling [14–16]. Notch is a transmembrane receptor interacting with ligands expressed on the surface of neighboring keratinocytes [16,17]. Upon activation, sequential
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Author Summary

Eczema (atopic dermatitis) is a common allergic skin inflammation that has a particularly high prevalence among children. Importantly, a large proportion of people suffering from eczema go on to develop asthma later in life. Although the susceptibility of eczema patients to asthma is well documented, the mechanism that mediates “atopic march”—the progression from eczema to asthma—is unclear. We used genetic engineering to generate mice with chronic skin-barrier defects and a subsequent eczema-like disorder. With these mice, we were able to investigate how skin-specific defects predisposed the lungs to allergic asthma. We identified thymic stromal lymphopoietin (TSLP), a cytokine that is secreted by barrier-defective skin into the systemic circulation, as the agent sensitizing the lung to allergens. We demonstrated that high systemic levels of skin-derived TSLP were both required and sufficient to render lung airways hypersensitive to allergens. Thus, these data suggest that early treatment of skin-barrier defects to prevent TSLP overexpression and systemic inhibition of TSLP, may be crucial in preventing the progression from eczema to asthma.

Results

RBP-jCKO Mice Develop an AD-Like Skin Phenotype

The ablation of Notch signaling in skin keratinocytes by removing RBP-j severely impacts the differentiation of basal layer keratinocytes and maintenance of upper spinous and granular cell layers [15,16]. Such a defect in epidermal stratification leads to an aberrant skin-barrier function signified by transepidermal water loss and penetration of dye through the defective barrier in RBP-j–deficient mice at birth [15,16]. After birth, the persistence of barrier defects in RBP-j-deficient skin is evident by the presence of reactive epidermal hyperplasia, TSLP overexpression, and up-regulation of antimicrobial peptides (Figure S2 and Table S1) [15,16,24,25]. Defective skin-barrier function has been suggested to be a hallmark of AD [6,7]. In agreement with this notion, the impaired skin-barrier function in RBP-jCKO mice initiated an inflammatory cascade culminating in the development of an AD-like skin phenotype. The hyperplastic epidermis, acanthosis, hyperkeratosis, parakeratosis, and mast cell infiltration were evident in RBP-jCKO skin as early as 1 wk after birth, followed by dramatic dermal mast cell accumulation, serum IgE elevation, and systemic Th2 cell expansion in adult RBP-j–deficient animals (Figure 1A–1C). Therefore, adult RBP-jCKO mice resemble humans with AD to a degree that allows us to examine the systemic consequences of an allergic inflammation in the skin.

AD-Like Skin Disease Predisposes RBP-j CKO Animals to Allergic Asthma

The lungs of 10-wk-old mice with RBP-j-deficient skins were normal and did not show any sign of allergic inflammation under standard housing conditions; however, the longest-living mutant animals (52 wk) did develop spontaneous lung inflammation (Figure 1D). To determine whether this phenotype was a true indicator of increased susceptibility to an asthmatic phenotype, we used an ovalbumin (OVA)-induced model of allergic inflammation with 5- to 7-wk-old RBP-jCKO mice. This protocol faithfully mimics the development of asthma in humans [18]. OVA-treated RBP-jCKO animals developed a more severe lung inflammation compared with OVA-treated wild-type littermates (Figure 2). Although the wild-type animals tolerated the intranasal OVA challenge well, two out of ten OVA-treated RBP-jCKO mice died during this procedure following a period of severe labored breathing. In the surviving mutants, the number of bronchoalveolar lavage (BAL) leukocytes and the percentage of BAL eosinophils were significantly higher compared with those of the wild-type littermates (Figure 2A and 2B). The absolute number of eosinophils was approximately 7-fold higher in the mutant mice. In addition, IgE was detectable only in the BAL fluid of RBP-j–deficient animals (Figure 2C). Histology of the lungs from OVA-challenged RBP-jCKO and wild-type controls clearly confirmed the existence of severe airway inflammation in RBP-jCKO mice, including significant leukocyte infiltration around the airways and
blood vessels, extensive goblet cell hyperplasia in the large airways, and distinct appearance of goblet cells in medium-sized airways (Figure 2D). Considering that RBP-\(j^{\text{CKO}}\) is not deleted in the lung (Figures S3 and S7), these data clearly show that the skin-barrier defect can serve as a primary risk factor for development of asthma in a normal lung.

**TSLP Signaling Is Required for the Atopic March in RBP-\(j^{\text{CKO}}\) Animals**

It is possible that atopic skin lesions are the essential components downstream of skin-barrier defects that initiate systemic atopy in RBP-\(j^{\text{CKO}}\) mice [3]. If this hypothesis were true, Th2 cells generated at the site of inflamed skin would migrate to other sites, including lung mucosa, and release high levels of Th2-derived cytokines, which would sensitize the lung airways to allergic inflammation. This model, however, is challenged by findings that show that Th2 cells generated in mouse models of AD specifically home to skin [26]. In addition, it is unclear whether the presence of AD lesions is required for initiation of the atopic march [11]. Skin-barrier defects led to systemic TSLP elevation [15], which remained elevated in the serum of RBP-\(j^{\text{CKO}}\) animals throughout life (Figure S2). On the basis of these observations and that localized TSLP overexpression in lung epithelium is capable of inducing asthma [18,19,21], we have articulated an alternative hypothesis: TSLP may be a systemic signal that sensitizes the animals to allergen exposure in the lung. In that case, high systemic levels of epidermal-derived TSLP should render RBP-\(j^{\text{CKO}}\) animals susceptible to the asthmatic phenotype upon exposure to allergen.

To test this hypothesis we deleted the \(\text{IL7R}^{\alpha}\) subunit of TSLP receptor in RBP-\(j^{\text{CKO}}\) animals (\(\text{Msx2-Cre}^{+};\text{RBP-}\text{j}^{\text{flox/flox}};\text{IL7r}^{\alpha/-}\); or RBP-\(j^{\text{CKO}};\text{IL7r}^{\alpha/-}\) [27]). This strategy was chosen because the TSLPR subunit of the TSLP receptor is linked to the RBP-\(j^{\text{CKO}}\) locus. The inhibition of TSLP reception reduced the severity of local AD-like inflammation in the skin of RBP-\(j^{\text{CKO}}\) animals as judged by the reduction of dermal mast cells in RBP-\(j^{\text{CKO}};\text{IL7r}^{\alpha/-}\) mice relative to their RBP-\(j^{\text{CKO}}\) littermates [28]. However, a significant elevation in mast cell number was still detectable in RBP-\(j^{\text{CKO}};\text{IL7r}^{\alpha/-}\) skin compared to wild type (Figure 3A and 3B). In addition, epidermal hyperplasia and TSLP overexpression that mark the presence of postnatal skin-barrier defects persisted in aged RBP-\(j^{\text{CKO}};\text{IL7r}^{\alpha/-}\) mice (Figure 3A and Figure S4). Importantly, deletion of \(\text{IL7R}^{\alpha}\) did not affect the intensity of systemic Th2 response to allergic skin inflammation.
and serum IgE levels remained elevated in RBP-jCKO;IL7rα/- mice (Figure 3C and 3D). This could be due to other cytokines/chemokines made by defective skin barrier in RBP-jCKO;IL7rα/- animals [15], which together with exposure to allergens/pathogens through the compromised skin, sustain a robust systemic Th2 response even in the absence of TSLP signaling [28]. Therefore, RBP-jCKO;IL7rα/- animals provide a suitable system in which to determine if skin-barrier defects and AD-like skin inflammation including a systemic Th2 response with its consequences (e.g., elevated IgE) could confer susceptibility to asthma in the absence of TSLP signaling.

We challenged the lung airways of age- and sex-matched RBP-jCKO;IL7rα/-, RBP-jCKO;IL7rα+/- (RBP-jCKO), IL7rα/-, and wild-type littermates with OVA. To prevent RBP-jCKO lethality, all the animals in this experiment received intranasal OVA challenges only twice. Despite this reduced exposure, RBP-jCKO animals developed a severe asthmatic response, which was absent in RBP-jCKO;IL7rα/- mice (Figure 3E–3H). Histological

Figure 2. Five- to seven-wk-old RBP-jCKO mice develop a severe allergic lung inflammation in an OVA-induced murine model of asthma. (A and B) There are more leukocytes (A) and a higher percentage of eosinophils (B) present in BAL fluid collected from OVA-treated RBP-jCKO mice in comparison to their wild-type littermates (Msx2-Cre/+;RBP-jflox/+;RBP-jflox/flox;RBP-jflox/+), indicating a severe airway inflammation in these mutant animals (M, macrophages; E, eosinophils; L, lymphocytes; N, neutrophils). (C) IgE reaches detectable levels only in BAL fluid from RBP-j-deficient mice, further indicating the high intensity of OVA-induced allergic inflammation in the RBP-jCKO lung. (D) Comparing lung histology of RBP-jCKO with wild-type littermates sensitized and challenged with OVA shows a significantly more intense lung inflammation in RBP-jCKO mice. H&E and PAS staining shows greater immune cell infiltration around the airways and blood vessels (arrowheads), increased airway remodeling, goblet cell hyperplasia (insets), and mucus overproduction in the mutant lung (representative pictures are presented; scale bar: 50 μm). "Controls" refers to RBP-jCKO and wild-type animals treated with PBS instead of OVA (n=4 for each group; *p<0.05, comparing the adjacent groups). These data are confirmed in additional independent experiments.

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analysis could not detect goblet cell hyperplasia or significant inflammation around the airways and vasculature in RBP-jCKO;IL7r-/- lungs (Figure 3E). Total white blood cell count and eosinophils in BAL fluid from OVA-treated RBP-jCKO;IL7r-/- mice was indistinguishable from IL7r-/- littermates (Figure 3F and 3G). In addition, IgE was undetectable in BAL fluid from OVA-treated RBP-jCKO;IL7r-/- mice (Figure 3H). These findings show that without a TSLP signal, no progression from allergic skin inflammation to asthma is observed, indicating that atopic march requires TSLP signals.

Airway Hyper-Responsiveness in RBP-jCKO Mice Is Blocked by Inhibiting TSLP Signaling

Airway hyper-responsiveness is a hallmark of allergic asthma [19]. To determine whether skin-derived TSLP could cause airway hyper-responsiveness in RBP-jCKO mice, we used the OVA-induced model of allergic inflammation and challenged the lung airways of age- and sex-matched RBP-jCKO;IL7r-/-, RBP-jCKO, IL7r-/-, and wild-type littermates twice with OVA. Thereafter, we measured the total lung resistance of OVA-treated mice at basal state (vehicle alone) or in response to increasing doses of nebulized methacholine. As expected, OVA-treated RBP-jCKO mice mounted a more severe airway resistance compared with their wild-type littermates when exposed to low doses of methacholine, and could not tolerate the higher doses of methacholine (Figure 4). RBP-jCKO;IL7r-/-, however, did not develop any significant airway resistance even when exposed to high doses of methacholine (Figure 4). These results confirm that skin-barrier defects predispose animals to asthmatic phenotypes and that TSLP is required for this predisposition.

TSLP Overexpression by the Skin Is Sufficient to Trigger the Atopic March

Although we showed that TSLP signaling is necessary for the development of asthma in RBP-j-deficient animals with chronic allergic skin inflammation, it remains unclear whether other factors associated with AD-like skin lesions contribute to this atopic march. TSLP overexpression in mouse skin led to an AD-like pathology on an inbred background [26]. However, we discovered that on an outbred background, the same K14-TSLP mice [15] maintained high serum TSLP levels under normal conditions (approximately 450 pg/ml) without any skin or lung inflammation (Figure 5 and Figure S5) [29]. Normal skin morphology and the lack of mast cell hyperplasia evident in K14-TSLP mice confirmed that in this outbred background and in the absence of skin-barrier defects, TSLP overproduction was insufficient to attract mast cells to the skin (Figure 5B). In addition, serum IgE levels and Th2 cell numbers in the peripheral lymph nodes of K14-TSLP mice were indistinguishable from the wild type, confirming the absence of allergic skin inflammation in K14-TSLP animals that were not exposed to any external stimulus (e.g., allergen exposure; Figure 5C and 5D). Therefore, outbred K14-TSLP mice constituted a
suitable model in which to examine whether epidermal TSLP overexpression alone would increase susceptibility to asthma upon allergen exposure.

Next, we sensitized and challenged the lung airways of 5- to 7-wk-old K14-TSLP<sup>tg</sup> mice and their age- and sex-matched wild-type littermates three times with allergen. As seen in OVA-treated RBP<sub>p</sub>CKO animals exposed to three doses of intranasal OVA, K14-TSLP<sup>tg</sup> mice developed a severe asthmatic response and significant lethality, which were not seen in the control group (Figure 6). Two out of eight OVA-treated transgenic mice died during the intranasal challenge. Histological examination of lungs from surviving K14-TSLP<sup>tg</sup> animals revealed pronounced airway remodeling with goblet cell hyperplasia and inflammation around the lung airways and vasculature (Figure 6A), which were negative for TSLP expression prior to OVA treatment (Figures S6 and S7). Total leukocyte count was less than 5-fold higher in BAL fluid from K14-TSLP<sup>tg</sup> mice compared with similarly treated wild-type animals (Figure 6B), which was mainly due to a severe BAL fluid eosinophilia in the transgenic animals (Figure 6C). In addition, IgE reached detectable levels only in BAL fluid from OVA-treated K14-TSLP<sup>tg</sup> mice (Figure 6D), which also experienced significantly higher serum IgE levels (Figure 6E) (1.6 μg/ml versus 0.18 μg/ml in OVA-treated wild-type serum, p<0.05). Thus, K14-TSLP<sup>tg</sup> mice developed an intense Th2 response upon allergen exposure. In the airway reactivity test, OVA-treated K14-TSLP<sup>tg</sup> mice showed a severe airway hyper-responsiveness to low doses of nebulized methacholine, which was not seen among similarly treated wild-type littermates (Figure 6F).

Figure 5. K14-TSLP<sup>tg</sup> mice exhibit no sign of skin or lung allergic inflammation under normal conditions. (A) Skin-derived TSLP overexpression leads to high systemic TSLP levels in the serum of K14-TSLP<sup>tg</sup> mice (n = 10 for each group; *p < 0.01). (B–D) Under normal conditions, skin TSLP overproduction on an outbred C57BL/6-CD1 background does not elicit any allergic inflammation in K14-TSLP<sup>tg</sup> mice. Note the normal skin histology (B), normal serum IgE levels (n = 10 in each group)(C), and normal number of peripheral IL-4-producing Th2 cells (D) in K14-TSLP<sup>tg</sup> mice at postnatal day 120 (P120). (E) H&E and PAS stained lung sections of K14-TSLP<sup>tg</sup> animals confirm that no inflammation or airway remodeling occurred at P120 (scale bar: 50 μm).

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Figure 6. OVA treatment reveals that K14-TSLP\textsuperscript{tg} mice are prone to asthmatic phenotype. (A) Histological analysis shows severe inflammatory cell infiltrates (arrowheads), airway remodeling, and goblet cell hyperplasia (arrow; insets) in the lung airways of 5– to 7-wk-old K14-TSLP\textsuperscript{tg} animals that are sensitized and challenged with OVA, but not in similarly treated wild-type littermates (scale bar: 50 μm). (B–D) BAL fluid analysis shows more leukocytes (B), a higher percentage of eosinophils (C), and specific appearance of IgE in lung airways of OVA-treated K14-TSLP\textsuperscript{tg} mice (D), confirming the higher intensity of the asthmatic phenotype conferred by the TSLP overexpression in the morphologically normal skin (M, macrophages; E, eosinophils; L, lymphocytes; N, neutrophils). (E) The severe allergic inflammation in OVA-treated K14-TSLP\textsuperscript{tg} lungs leads to drastic elevation of serum IgE in the animals. ''Controls'' refers to K14-TSLP\textsuperscript{tg} and wild-type mice treated with PBS alone (*p<0.05, comparing the adjacent groups). (F) Total lung resistance (R\textsubscript{L}) measurements show that OVA-treated K14-TSLP\textsuperscript{tg} lungs are hyper-responsive to increasing doses of nebulized methacholine as compared to OVA-treated wild types (V: vehicle-only basal measurement; *p<0.05). These findings are confirmed in additional independent experiments.

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Even though outbred K14-TSLP\textsuperscript{tg} animals developed a severe asthmatic phenotype, their skin retained its normal appearance after OVA treatment, confirming that the lung inflammation did not depend on a concurrent skin lesion (Figure S8). Although low levels of TSLP were expressed in the trachea, the K14-TSLP transgene was not expressed in the lung of K14-TSLP\textsuperscript{tg} animals and TSLP protein was not detectable in their BAL fluid (Figures S6 and S7). Therefore, these observations are most consistent with a model in which systemic TSLP is sufficient to predispose mice to allergic lung inflammation.

Discussion

This report describes the mechanism that mediates the progression of a local allergic skin disease to a disseminated allergic inflammation downstream of skin-barrier defects in mice. We show that creating an intrinsic skin-barrier defect (by removing Notch signaling specifically from some skin keratinocytes) results in the sensitization of the lung airways to allergens. Similar to the clinical cases, barrier-defective mice require a secondary insult by allergens to expose their predisposition to asthma. Importantly, we show that epidermal sensitization with a common allergen [3] is not required to achieve this pronounced lung hyper-responsive-ness. Therefore, RBP-\textsuperscript{K14-TSLP\textsuperscript{tg}} animals demonstrate that skin-barrier impairment provides a signal that promotes development of a distant atopy in the lung, most likely by a systemic, diffusible cytokine. We identified this signal to be TSLP, a central player in the sensitization of the lung airways to allergens. In contrast to TSLP overexpression by lung epithelia, which does not result in systemic accumulation of this cytokine [18], overexpression of TSLP in the skin results in high systemic availability [15]. Therefore, skin is capable of acting as a signaling organ, driving susceptibility to allergic inflammation in another barrier organ (i.e., lung) by releasing TSLP.

Importantly, we were able to show that TSLP was necessary to predispose RBP-\textsuperscript{K14-TSLP\textsuperscript{tg}}-deficient animals to the asthmatic phenotype. Concomitant removal of IL7rα subunit of the TSLP receptor in RBP-\textsuperscript{K14-TSLP\textsuperscript{tg}} mice prevented the atopic march despite persistent AD-like pathology and elevated serum IgE levels. Complementing this experiment is the observation that overexpression of TSLP by skin keratinocytes leads to high serum levels of TSLP that are sufficient to sensitize the lung airways to allergic inflammation in the absence of any skin pathology. If the low levels of TSLP in the trachea of K14-TSLP\textsuperscript{tg} mice contribute to the phenotype, they do so by adding to the already high levels of skin-derived serum TSLP. Because TSLP can activate dendritic cells [30], T cells [31,32], and myeloid cells [33], we speculate that its high systemic levels directly contribute to the atopic march downstream of TSLP-mediated activation remains a subject for future investigation. Of note, elevated TSLP expression is also reported in psoriatic skin [34]; however, there is no evidence linking psoriasis to predisposition to asthma. This phenomenon could be explained by the dominance of Th1 response in patients with psoriasis and the unresponsiveness of their immune cells to TSLP [34].

The findings outlined in this report have a potentially important implication for human health. They provide a plausible explanation of why a large number of AD patients develop asthma and other allergic disorders later in life [35,36]. Because TSLP overproduction is triggered by skin-barrier defects and faithfully mirrors the severity of these defects [15], our data emphasize that an early and aggressive treatment of the underlying skin-barrier defects in AD-prone patients [10,11] may be more beneficial in preventing asthma than treating the outbreaks of AD lesions [6]. Although serum TSLP levels in AD patients are yet to be determined, high TSLP expression levels in human AD lesions [37] suggest that this diffusible cytokine could also reach systemic levels sufficient to trigger atopic march in AD patients. If indeed serum TSLP levels are elevated in AD patients, our findings suggest that an aggressive management of TSLP levels in these patients will lower the incidence of asthma later in their lives.

Materials and Methods

Mice

\textit{Mice\textsuperscript{3}}

Mixed C57BL/6 and CD1 outbred genetic background, which are generated as previously described [15]. All animals were kept in the animal facility under specific pathogen-free conditions. All procedures were performed in accordance with University of Pennsylvania institutional animal care and use committee guidelines.

OVA Treatment

The allergic sensitization and lung airways challenge with OVA was carried out as previously outlined [18]. In brief, wild-type and mutant mice at 5–7 wk of age were sensitized on days 0 and 14 by 100 μg OVA (Sigma) dissolved in 250 μl intraperitoneal injection of antigen solution containing 50 μg OVA (Sigma) dissolved in 1.3 mg aluminum hydroxide gel (Sigma) and PBS. On days 1, 2, and 3, mice were intranasally challenged with 100 μg OVA (Sigma) dissolved in 40 μl of PBS. Mice cohorts designated as "controls" included the mutant and wild-type littermates used in each experiment. All mice were maintained in the animal facility under University of Pennsylvania institutional animal care regulations.
however, within-sex analyses were performed to avoid any sex effect on phenotypes observed.

Airway Reactivity Test

The airway responsiveness to aerosolized methacholine was determined in OVA-treated animals by measuring total lung resistance and dynamic compliance as previously outlined [38]. In brief, mice were treated with OVA as described in the preceding section (two intranasal OVA challenges for all mice in rescue experiments (Figure 4) and three intranasal OVA challenges for all mice in sufficiency experiments (K14-TSLP<sup>Δ</sup> and wild-type; Figure 6). The animals were then anesthetized for airway reactivity test on day 24 post OVA sensitization. They were ventilated through a tracheotomy and monitored for intrapleural pressure using an oroesophageal tube. PBS (vehicle) or methacholine (Sigma) through a tracheotomy and monitored for intrapleural pressure using an oroesophageal tube. PBS (vehicle) or methacholine (Sigma) in PBS were delivered at 3-min intervals using an in-line nebulizer. The respiratory flow signals were collected between deliveries using a pneumotach (SenSym SCXL004, Buxco Electronics). For detailed description of the device, refer to [39].

Histology

Dorsal skin samples were harvested from the mice and fixed in 4% PFA at 4°C overnight. Lungs were inflated through the trachea to 25-cm water pressure with 4% PFA prior to excision from the chest and fixation. These lungs were PFA-fixed for 24 hr at 4°C and embedded in paraffin. The paraffin-embedded tissues were sectioned at 5-6 μm and stained with hematoxylin and eosin (H&E), toluidine blue, or periodic acid-Schiff (PAS). For RBP-J immunohistochemical staining, anti-RBP-J antibody (clone T6709, Institute of Immunology) and biotinylated anti-rat secondary antibody were used. HRP-conjugated streptavidin and DAB substrate kit (Pierce) were used to visualize the signal. Hematoxylin was used to counterstain the sections. For TSLP immunostaining, paraffin-embedded tissue samples and biotinylated anti-TSLP antibody (R&D Systems) were used. Sections were counterstained with DAPI nuclear stain.

BAL Analyses

BAL fluid was collected by infusing the lungs of the anesthetized mouse with 1-ml PBS through tracheal insertion of a Surflo catheter (Terumo Medical). Leukocyte count in BAL fluid was determined in OVA-treated animals by measuring total lung resistance and dynamic compliance as previously outlined [38]. The respiratory flow signals were collected between deliveries using a pneumotach (SenSym SCXL004, Buxco Electronics). For detailed description of the device, refer to [39].

ELISA

Serum TSLP levels were determined using Quantikine mouse TSLP kit according to manufacturer’s instructions (R&D Systems). Serum and BAL fluid IgE levels were measured using Mouse IgE ELISA kit (Immunochemistry Consultants Laboratory).

Flow Cytometry

Intracellular cytokine staining was conducted to estimate Th2 cell census as previously described [26]. Single cell suspensions were prepared from spleen and lymph node samples and cultured in presence of PMA (50 ng/ml), ionomycin (1 μg/ml) and monensin (10 μg/ml) for 4 h. Cells were then stained with phycoerythrin (PE)-cy7 conjugated anti-CD4 antibody (552775, BD Bioscience Pharmingen). After fixation in 2% PFA and permeabilization with 0.5% saponin, cells were stained with PE conjugated anti-IL4 (554389) and APC conjugated anti-IFN-γ (554412) antibodies from BD Bioscience Pharmingen [40].

Conventional PCR for the RBP-J<sup>Δ</sup> allele was performed on genomic DNA isolated from skin and lung of adult RBP-J<sup>Δ</sup>-/ mice using KlenTaq10 (DNA Polymerase Technology) supplemented with 1.3 M final concentration of betaine (amplification cycles = 32). The following primers were used to distinguish between deleted (RBP-J<sup>Δ</sup>) and floxed (RBP-j<sup>flox</sup>) alleles of RBP-J: Deleted allele: 5'-TGTTTGGCACCAGAATTGTTGG-TATTGTG-3' and 5'-ATTGTTGAGGTCTGATTGTTCGTGATTGC-3'. Floxed allele: 5'-TGTTTGGCACCAGAATTGTTGG-TATTGTG-3' and 5'-AGGTACCTGGTACTAAGTCTGTG-GACCG-3'.

Statistical Analysis

The quantitative measurements were assessed using Student t-test as the test of significance and presented as mean ± standard deviation in bar graph format. Our studies were conducted in outbred cohorts of animals; therefore, we used nested ANOVA to exclude any effect of between-family differences and make certain the significant differences observed were solely attributable to the gene removed (RBP-j<sup>Δ</sup>-/ and RBP-j<sup>flox</sup>−/+ or overexpressed (K14-TSLP<sup>Δ</sup>). Statistical significance was determined in OVA-treated animals by measuring total lung resistance and dynamic compliance as previously outlined [38]. The primers used to amplify TSLP were: 5'-CCAGGCTACCTGAAACTGA-3' and 5'-CTCTGA-GATTGCATGAGG-3'.

Supporting Information

**Figure S1** RBP-j<sup>Δ</sup>-/ animals die prematurely due to their severe skin phenotype (*p<0.001 compared to wild-type life span, log-rank test). A few mutant mice that survive up to one year (red circles), however, develop spontaneous lung inflammation. Found at: doi:10.1371/journal.pbio.1000067.s001 (78 KB TIF)

**Figure S2** Serum TSLP levels in RBP-j<sup>Δ</sup>-/ mice are highly elevated. TSLP overproduction is evident in RBP-j<sup>Δ</sup>-/ serum at 1 wk after birth, reaching extreme levels in the adult animals (*p<0.01, comparing the mutants to the wild-type littermates). Found at: doi:10.1371/journal.pbio.1000067.s002 (55 KB TIF)

**Figure S3** Lung epithelium is normal in RBP-j<sup>Δ</sup>-/ animals. (A) PCR analysis of DNA isolated from adult RBP-j<sup>Δ</sup>-/ (Msx2-Cre/+/RBP-j<sup>flox</sup>/flox) and wild-type (RBP-j<sup>flox</sup>/flox) skin and lung shows that RBP-J locus is intact (i.e., RBP-j<sup>Δ</sup>-/ is not deleted) in the lung (Δ-deleted allele; M: molecular marker; S: skin; L: lung). (B) Immunohistochemical analysis for RBP-J protein confirms that RBP-j is present in the lung airway epithelium. Skin sections stained under the same condition are presented as controls (scale bar: 50 μm). Found at: doi:10.1371/journal.pbio.1000067.s003 (7.35 MB TIF)

**Figure S4** Serum TSLP levels of 10-wk-old RBP-j<sup>Δ</sup>-/ mice are highly elevated. This indicates that skin-barrier defects caused by the loss of RBP-J in epidermal
keratinocytes persist in the absence of IL7Rx [a = 5 for each group; * p<0.01]. Note that serum TSLP levels in RBP-JCKO:IL7rx-/- mice are consistently and significantly higher than in RBP-JCKO animals. We are currently investigating the underlying reason for this surge.

Found at: doi:10.1371/journal.pbio.1000067.s004 (50 KB TIF)

**Figure S5**
The ear and skin of K14-TSLP<sup>tg</sup> mice appear normal at P120. This emphasizes that in an outbred genetic background (C57BL/6 and CD1 mix) these transgenic animals do not develop any skin inflammation under normal conditions.

Found at: doi:10.1371/journal.pbio.1000067.s005 (1.3 MB TIF)

**Figure S6**
There is no TSLP overexpression detectable in K14-TSLP<sup>tg</sup> lung airways or parenchyma. Because the K14 gene is expressed in basal cells located in the trachea, we analyzed mRNA levels by qRT-PCR on samples isolated from epidermis, trachea and lung of K14-TSLP<sup>tg</sup> and wild-type mice. This analysis shows that the K14-TSLP transgene is active in basal cells within the trachea, but the overall levels are 200-fold lower than those made by epidermal keratinocytes (###p<0.0000001 and *p<0.01 compared to wild type).

Found at: doi:10.1371/journal.pbio.1000067.s006 (145 KB TIF)

**Figure S7**
Lung epithelial does not overexpress TSLP in adult K14-TSLP<sup>tg</sup>, RBP-JCKO, or RBP-JCKO:IL7rx-/- animals. Immunofluorescence staining for TSLP protein (red) confirms that TSLP is overexpressed only in the epidermal keratinocytes of the mutant mice. All sections are stained under the same conditions. Dotted lines outline the basement membrane and asterisks highlight the epidermal keratin cysts present in RBP-JCKO skin (scale bar: 50 μm).

Found at: doi:10.1371/journal.pbio.1000067.s007 (5.3 MB TIF)

**Figure S8**
The skin of OVA-treated K14-TSLP<sup>tg</sup> mice remains normal. H&E and toluidine blue staining of K14-TSLP<sup>tg</sup> and wild-type skin show no significant signs of cutaneous inflammation in the transgenic mice (scale bar: 30 μm).

**References**


**Table S1**
Major antimicrobial peptides are overexpressed in postnatal epidermis of RBP-JCKO animals. This overexpression indicates the persistence of the skin-barrier defect in RBP-JCKO mice after birth [25]. Data are extracted from a microarray study on P9 epidermis of RBP-JCKO and wild-type littermates as previously described [15]. The fold increase of antimicrobial peptide mRNA in RBP-J deficiency epidermis relative to wild type is presented.

Found at: doi:10.1371/journal.pbio.1000067.s009 (37 KB PDF)

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**Author Contributions**

The author(s) have made the following declarations about their contributions: Conceived and designed the experiments: SD RK.

Performed the experiments: SD MM MJH.

Analyzed the data: SD MM.

Contributed reagents/materials/analysis tools: MJH.

Wrote the paper: SD RK.

**Note Added in Proof**

After this manuscript was accepted for publication, similar observations were reported by Zhang et al: Zhang Z, Hener P, Frossard N, Kato S, Metzger D, et al. (2009). Thymic stromal lymphopoietin overproduced by keratinocytes in mouse skin aggravates experimental asthma. Proc Natl Acad Sci U S A 106: 1536–1541.

**References**


