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P-glycoprotein deficiency at the blood-brain barrier increases amyloid-β deposition in an Alzheimer disease mouse model

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Accumulation of amyloid-β (Aβ) within extracellular spaces of the brain is a hallmark of Alzheimer disease (AD). In sporadic, late-onset AD, there is little evidence for increased Aβ production, suggesting that decreased elimination from the brain may contribute to elevated levels of Aβ and plaque formation. Efflux transport of Aβ across the blood-brain barrier (BBB) contributes to Aβ removal from the brain. P-glycoprotein (Pgp) is highly expressed on the luminal surface of brain capillary endothelial cells and contributes to the BBB. In Pgp-null mice, we show that [125I]Aβ40 and [125I]Aβ42 microinjected into the CNS clear at half the rate that they do in WT mice. When amyloid precursor protein–transgenic (APP-transgenic) mice were administered a Pgp inhibitor, Aβ levels within the brain interstitial fluid significantly increased within hours of treatment. Furthermore, APP-transgenic, Pgp-null mice had increased levels of brain Aβ and enhanced Aβ deposition compared with APP-transgenic, Pgp WT mice. These data establish a direct link between Pgp and Aβ metabolism in vivo and suggest that Pgp activity at the BBB could affect risk for developing AD as well as provide a novel diagnostic and therapeutic target.

Introduction

Conversion of amyloid-β (Aβ) into toxic soluble and insoluble forms is accelerated at higher concentrations, implying that pathways influencing Aβ production or elimination could potentially modulate disease progression. Aβ is secreted from neurons into brain interstitial fluid (ISF), where it is eliminated by proteolytic degradation (1, 2), passive bulk flow (3), and active transport across the blood-brain barrier (BBB) (4, 5). The latter, representing efflux across the BBB fluid (ISF), where it is eliminated by proteolytic degradation (1, 2), provides a novel diagnostic and therapeutic target.

Nonstandard abbreviations used: Aβ, amyloid-β; AD, Alzheimer disease; APP, amyloid precursor protein; BBB, blood-brain barrier; ISF, interstitial fluid; LRP1, low-density lipoprotein receptor–related protein 1; MDR1, multidrug resistance-1; Pgp, P-glycoprotein; RAGE, receptor for advanced glycation endproducts.

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in LRP1 expression at the BBB may have partially contributed to Pgp-null mice, which is consistent with previous studies showing mice as compared with Pgp WT mice (Figure 1E). This decrease in brain ISF $A_\beta$ decreased by 50% in brain capillaries of 2- to 3-month-old Pgp-null compared with Pgp WT (FVB) versus Pgp-null, respectively. Similarly, bulk flow pathways, accounting for less than 20% of clearance, which overexpress human amyloid precursor protein (APP), with levels of LRP1 were comparable (data not shown), interestingly, levels of LRP1 were unchanged in the knockout animals as demonstrated by nearly identical levels of inulin remaining in both groups. As shown in Figure 1B, after passive elimination was taken into account, 31% of injected $A_\beta$ were recovered in both groups. (B) Calculation of $[^{252}I]A_\beta_{42}$ clearance via the BBB and ISF bulk flow mechanisms (from data shown in A). Significantly less $A_\beta_{40}$ was cleared via the BBB in Pgp-null mice compared with Pgp WT mice ($P = 0.0033$). (C) Brain recovery of $[^{252}I]A_\beta_{42}$ (left) and $[^{14}C]\text{inulin}$ (right). More $[^{252}I]A_\beta_{42}$ was cleared from the brain in Pgp WT than Pgp-null mice ($P = 0.0047$). Approximately 85% of $[^{14}C]\text{inulin}$ was retained in both groups. (D) Similar to calculations for $A_\beta_{40}$ clearance, significantly less $A_\beta_{42}$ was cleared from the brain of Pgp-null mice ($P = 0.0004$). (E) LRP1 expression was decreased by 51% in cerebral vessels of 2- to 3-month-old Pgp-null mice compared with WT FVB controls ($P = 0.002$). Left: Relative absorbance representing the signal obtained from the 85-kDa LRP band on the Western blots normalized to the loading control is shown on the y axis. Right: representative lanes from an 85-kDa LRP1 peptide (LRP1-85) Western blot and protein loading control. Values are mean ± SEM; $n = 4–8$ per group.

(P = 0.0123 and $P = 0.0047$, respectively), strongly suggesting that $A_\beta$ elimination was hindered in these animals. Removal of $[^{3}H]\text{leucine}$, a Pgp-independent, actively transported molecule, was unaffected at 10 minutes after injection (61.4% ± 3.8% and 62.7% ± 5% retained in Pgp WT [FVB] versus Pgp-null, respectively). Similarly, bulk flow pathways, accounting for less than 20% of clearance, appeared unchanged in the knockout animals as demonstrated by nearly identical levels of inulin remaining in both groups. As shown in Figure 1B, after passive elimination was taken into account, 31% of injected $A_\beta_{40}$ was transported across the BBB in WT mice, whereas only approximately 14% was cleared in the Pgp-null animals ($P = 0.0033$). Similarly, 16% of $A_\beta_{42}$ was transported in Pgp WT mice compared with 6% in Pgp-null mice (Figure 1D). Compared with $A_\beta_{40}$, significantly less $A_\beta_{42}$ was removed from the brain in WT and Pgp-null mice, which is consistent with previous studies showing that $A_\beta_{40}$ is generally actively transported across the BBB more than $A_\beta_{42}$ (7). LRP1 has been previously shown to transport $A_\beta$ across the BBB (5, 7). While overall levels of LRP1 in brain homogenates were comparable (data not shown), interestingly, levels of LRP1 were decreased by 50% in brain capillaries of 2- to 3-month-old Pgp-null mice as compared with Pgp WT mice (Figure 1E). This decrease in LRP1 expression at the BBB may have partially contributed to decreased $A_\beta$ clearance in Pgp-null mice.

Next, we assessed how acute inhibition of Pgp activity affected brain ISF $A_\beta$ levels. We utilized APPsw (also called Tg2576) mice, which overexpress human amyloid precursor protein (APP), with a mutation that in humans causes an autosomal dominant form of early-onset familial AD (14). A selective Pgp inhibitor, XR9576 (15), was used to block Pgp activity at the BBB in APPsw mice. Because brain capillary endothelial cells transport molecules out of the brain ISF into the periphery, we directly assessed this fluid $A_\beta$ levels following intravenous administration of XR9576 at 80 mg/kg, a dose that inhibits Pgp function at the BBB, as determined by $[^{99m}Tc]\text{Sestamibi}$ analysis (data not shown). Using in vivo microdialysis in awake, freely moving mice (16, 17), we first determined a basal concentration of ISF $A_\beta$ over 5 hours, then treated mice with either vehicle or XR9576 and continued to assess ISF $A_\beta$ levels for an additional 10 hours (Figure 2A). Eight hours after treatment, ISF $A_\beta$ levels increased almost 30% compared with vehicle-treated mice ($P < 0.05$), suggesting that acute inhibition of Pgp at the BBB caused decreased elimination of $A_\beta$ from brain ISF. Ten hours after treatment with either vehicle or XR9576, cerebral microvessels were also isolated and assessed for levels of LRP1 by Western blotting. LRP1 levels in endothelial cells did not change (Figure 2B), indicating that, in this case, the change in ISF $A_\beta$ levels is not attributable to LRP1 downregulation but is likely due to specific inhibition of Pgp.

Next, we determined whether the absence of Pgp at the BBB would promote $A_\beta$ deposition. We bred APPsw–/– mice to mdr1a/b–/– animals to produce littermate mice that were positive for the APPsw transgene and either Pgp-null (APPsw–/–, mdr1a/b–/–) or Pgp WT (APPsw–/–, mdr1a/b–/–). To confirm that transgenic expression of mutated human APP did not impact basal function of Pgp in vivo, biodistribution studies utilizing $[^{99m}Tc] \text{Sestamibi}$, a radio-
pharmaceutical that has been validated as a sensitive probe of Pgp transport function (18), were performed in 2- to 4-month-old WT (mdr1a/b+/+) and Pgp-null (mdr1a/b–/–) mice, both in the absence and presence of the APPsw+/– transgene. Normally, [99mTc]Sestamibi is readily transported out of the brain by Pgp. However, in Pgp-null animals, [99mTc]Sestamibi levels increase in the brain as it permeates cerebral capillaries and is not removed effectively. Five minutes after intravenous bolus injection of [99mTc]Sestamibi (2 μCi), brain content of the tracer was 0.22% ± 0.02% injected dose/g (ID/g) in WT controls versus 0.85% ± 0.14% ID/g in Pgp-null mice (Figure 3). There was no difference in blood content of the tracer between the 2 strains (P > 0.2). This approximately 3.5-fold increase in brain penetration of [99mTc]Sestamibi in Pgp-null mice was consistent with previous studies (19). In animals of similar age that expressed the APPsw+/– transgene, brain penetration of the tracer was also increased by approximately 3.5-fold in Pgp-null animals compared with those expressing Pgp (Figure 3). Thus, basal activity of Pgp was independent of the APPsw+/– transgene (P > 0.3).

When allowed to survive to 12 months, APPsw+/– mice exhibited substantial Aβ deposition throughout the neocortex and hippocampus (Figure 4, A and B). The area covered by Aβ immunoreactivity in the hippocampus was significantly greater in APPsw, Pgp-null animals compared with APPsw, Pgp WT littermates (P = 0.0041; Figure 4C). The amount of fibrillar Aβ, as determined by thioflavine S staining, was also significantly elevated in Pgp-null animals (P = 0.0276; Figure 4D). However, when the amount of thioflavine S was normalized to total plaque load, there was not a significant difference in the percentage of fibrillar plaques between groups (Figure 4E). This suggests that while Pgp deletion leads to greater Aβ deposition, Pgp does not preferentially alter the conversion of plaques into fibrillar structures. There was no apparent change in the amount of vascular Aβ accumulation in 12-month-old Pgp-null animals; however, it remains possible that at later ages, when there is a greater amount of cerebral amyloid angiopathy, there may be a noticeable affect. The mass of insoluble Aβ42, as assessed by guanidine extraction and Aβ ELISA, was 2-fold greater in the hippocampus of Pgp-null animals (P = 0.0499; Figure 4F), while there was only a trend toward an increase in insoluble Aβ40 (P = 0.2472; Figure 4G). Carbonate-extractable Aβ40 and Aβ42 was not different between these groups (data not shown). Similar to the hippocampus, there was a trend toward elevated Aβ plaque load in the neocortex of Pgp-null mice; however, this increase was not statistically significant (Figure 4H). These findings indicate that abrogation of the Pgp transporter at the BBB gave rise to greater Aβ accumulation within the brain and suggest that modulation of Pgp activity can directly influence progression of Aβ pathology. Because genetic deletion of Pgp also causes a decrease in the level of LRP1 at the BBB (Figure 1), it remains possible that the changes in Aβ deposition are due to a combined decrease in both Aβ transporters. In fact, these Aβ transporters may be playing a synergistic role, with LRP1 functional on the basolateral surface and Pgp on the luminal surface of brain endothelial cells.
Pgp shares sequence homology and substantial pharmacological cross-reactivity with several members of the ATP-binding cassette superfamily of transporters that are also expressed at the BBB, including multidrug-resistance–associated protein-1 (MRP1, also known as \textit{ABCC1}) (9). In 12-month-old APP\textsubscript{sw}, Pgp-null (\textit{APPsw}+/−, \textit{mdr1a/b}−/−) mice, expression of MRP1 in hippocampal homogenates was elevated compared with Pgp WT (\textit{APPsw}+/−, \textit{mdr1a/b}+/+) mice (\(P = 0.0401\); data not shown). Although Pgp and MRP1 have overlapping pharmacological profiles, it is unknown whether MRP1 can also transport Aβ in vivo. In vitro studies suggest that MRP1 cannot transport Aβ (12). Receptor for advanced glycation endproducts (RAGE), a transporter on the BBB that is responsible for transporting Aβ from the blood into the brain (20, 21), was expressed at similar levels in hippocampal homogenates of 12-month-old APP\textsubscript{sw}, Pgp-null and APP\textsubscript{sw}, Pgp WT mice (data not shown). The potential contribution of each of these transporters to Aβ clearance from the brain and how they might interact or compensate for each other requires further investigation.

Discussion

Herein, we demonstrate that Pgp is involved in the transport of Aβ across the BBB in vivo and that ablation of Pgp at the BBB enhances Aβ deposition. While Pgp deletion has not been reported in humans, several mechanisms for modulation of Pgp activity at the BBB could influence AD. Several Pgp single nucleotide polymorphisms, such as the exon 26 3435T allele, have been described that decrease Pgp expression and result in functional differences in oral absorption and disposition of drugs (22). It is conceivable that similar polymorphisms in Pgp could decrease Aβ transport out of the brain, thereby increasing the probability of plaque formation. Conversely, polymorphisms that increase Pgp transport activity may decrease the risk of Aβ deposition and development of AD. A detailed genetic analysis of Pgp polymorphisms in relation to AD in humans could test this hypothesis.

In addition to the impact of genetic polymorphisms on Pgp activity, many commonly used drugs and herbal medicinals alter Pgp function. For example, dexamethasone, morphine, rifampin, quercetin, and St. John’s wort are reported to enhance Pgp activity, while verapamil, cyclosporine A, erythromycin, fluoxetine, progesterone, HIV protease inhibitors, and several statins inhibit Pgp activity (for review, see refs. 9, 23). While some of these compounds only affect Pgp activity slightly in short-term assays, with chronic use or in combination, it is possible that these drugs could have a greater effect on Pgp function and, consequently, on brain Aβ levels. Since Pgp function can be modulated pharmacologically, there is potential for drugs that enhance activity to increase Aβ clearance from
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Pgp as a Drug Target for AD

Affecting Aβ transport, and dietary factors can influence Pgp function, thus potentially affecting Aβ transport, it will be interesting to determine whether these features correlate with the incidence of AD.

Methods

Animals. All experimental procedures involving animals were approved by the Animal Studies Committee at Washington University and performed in accordance with their guidelines. We utilized 8- to 10-week-old mdr1a/b–/– double-knockout mice on an FVB background (Taconic) and WT controls (also on an FVB background) for our BBB transport studies. We also bred mdr1a/b–/– mice to APPsw+–/– hemizygous mice (Tg2576 on a C57BL/6-SJL background; a generous gift from K. Ashe, University of Minnesota, Minneapolis, Minnesota, USA). We then bred F1 offspring from this colony to each other to produce litterserm dr1a/b homozygous (+/+) and double-knockout (−/−) mice that were hemizygous for the APPsw+–/– transgene. Animals were screened for the presence of Pgp as well as mdr1a and mdr1b genes by PCR analysis of tail DNA. Aβ content in the brain was analyzed when animals were 12 months old.

BBB transport. Transport of human [125I]Aβ18–23 [125I]Aβ42 across the BBB and calculations were performed as previously described (4, 8). Briefly, a guide cannula was implanted stereotaxically into the right caudate-putamen of anesthetized mice at coordinates 0.9 mm anterior to bregma, 1.9 mm lateral, and 2.9 mm below the dura. Animals were allowed to recover after surgery prior to radiotracer studies. The clearance experiments were performed before substantial chronic processes occurred, typically 4–6 hours following cannula implantation, to allow the BBB to at least partially repair and exclude large molecules (8, 16). Twelve nanomoles of [14C]inulin and either [125I]Aβ18–23 or [125I]Aβ42 was coinjected in a volume of 0.5 μl over 5 minutes using an ultra micropump (World Precision Instruments Inc.). The specific activities of peptide were in the range of 80–120 μCi/μg. The percentage of radioactivity remaining in the brain after microinjection was determined as percentage recovery in brain = 100 × (Ni/N0), where N0 is the radioactivity remaining in the brain at the end of the experiment and Ni is the radioactivity microinjected into brain ISF, i.e., the disintegration per minute for [14C]inulin and the counts per minute for trichloroacetic acid–precipitable [125I]radioactivity (intra-brain Aβ). The percentage of Aβ cleared through the BBB was calculated as [(1 – Ni/(N0/Ni)) – (1 – N(Ni/NI))] × 100, using a standard time of 30 minutes. D-[4,5-3H]leucine (Amersham Biosciences) transport across the BBB was assessed using a method similar to that described above: 750 nmol of [3H]leucine, specific activity 1.6 Ci/mg, was injected in a volume of 0.5 μl followed by measurement of radioactivity within the brain at 10 minutes.

ISF Aβ in vivo microdialysis. In vivo microdialysis to assess brain ISF Aβ1–42 in the hippocampus of awake, freely moving 3-month-old APPsw+–/– mice was performed exactly as described previously (16, 17). Aβ1–42 represents all Aβ peptides beginning at amino acid 1 of the N-terminus with the C-terminus being variable, but predominantly either ending at position 40 or 42. An initial 4– to 6-hour recovery period elapsed after guide implantation and probe insertion to allow for tissue recovery, followed by collection of microdialysis samples for 5 hours at 1-hour intervals at a flow rate of 1.5 μl/min to establish basal level of ISF Aβ. XR9576 (80 mg/kg body weight; Xenova QLT Inc.; diluted in 5% dextrose) or vehicle was injected into the jugular vein, and an additional 10 one-hour microdialysis samples were collected. Concentrations of ISF Aβ1–42 for each mouse were expressed as the percentage of basal level for each mouse (mean of 5 hours prior to treatment).

Histological analysis. Tissue sections were cut in the coronal plane at 50 μm on a freezing sliding microtone from the genu of the corpus callosum through the caudal extent of the hippocampus. The percent surface area covered by Aβ-immunoreactivity deposits (% Aβ load), as identified with a mouse monoclonal antibody against the N terminus of human Aβ, m3D6 (Eli Lilly and Co.), was quantified following unbiased stereological principles as described (28). Thioflavin S staining was performed stereologically and quantified as previously described (29). Aβ and thioflavin S load were determined in the cingulate cortex and hippocampus in 3 brain sections, each separated by 300 μm.

Aβ quantification. Aβ18–22 and Aβ40 levels in brain tissue were determined by sandwich ELISA as previously described (16). Briefly, m266, m2G3, or m21F12 were used as coating antibodies to capture Aβ1–42, Aβ40, or Aβ42, respectively. Biotinylated m3D6, a human-specific antibody against the N terminus of Aβ, was used as the reporting antibody in each ELISA. To evaluate the carbonate-soluble and -insoluble pools of Aβ in brain tissue, we performed a carbonate extraction (100 mM carbonate, 50 mM NaCl, protease inhibitors, pH 11.5) of hippocampal tissue (1:10, wt/vol) on ice followed by a 5 M guanidine, pH 8.0 extraction for 3 hours at room temp. Tissue samples were dounce homogenized and spun in a microcentrifuge at 21,000 g for 15 minutes at 4°C following each extraction.

Western blotting. Hippocampal tissue from 12-month-old APPsw, Pgp WT, and APPsw, Pgp-null littermates was homogenized in 150 mM NaCl, 50 mM Tris pH 7.4, 0.5% deoxycholic acid, 0.1% SDS, 1% Triton X-100, 2.5 mM EDTA, and protease inhibitors. Western blotting for MRPI, LRPI, and RAGE were performed using 3–8% NuPAGE Tris-Acetate gels (Invitrogen Corp.) with 30 μg of protein loaded per lane. Cerebral microvessels from 2- to 3-month-old Pgp WT (FVB) and Pgp-null mice were isolated as described (30). Microvessels were homogenized in cold lysis buffer: 150 mM NaCl, 50 mM Tris pH 7.4, 0.1% SDS, 1% Triton X-100, and complete protease inhibitor cocktail (Roche Diagnostics Corp.). Individual protein homogenates were separated under nonreducing conditions, transferred to nitrocellulose membrane, and probed with rabbit anti-LRP1-85 (LRP, β-chain specific; EMD Biosciences Inc.), rabbit anti-RAGE (Santa Cruz Biotechnology Inc.), or rat anti-MRP1 (Signet Laboratories), followed by goat anti-rabbit or donkey anti-rat secondary antibodies conjugated to HRP (Jackson ImmunoResearch Laboratories Inc.). Mouse anti-tubulin (Sigma-Aldrich) or anti-β-actin (Santa Cruz Biotechnology Inc.) were used to detect control proteins in each lane. Bands were detected with Lumigen-TMAE (Amersham Biosciences) and captured digitally using Kodak ImageStation 440CF. The relative absorbance of each band was detected by densitometry and normalized to protein loaded in each lane. Analysis was performed using Kodak 1D Image Analysis software version 3.6.


