2011

Engraftment of cells from porcine islets of Langerhans following transplantation of pig pancreatic primordia in non-immunosuppressed diabetic rhesus macaques

Sharon A. Rogers
Washington University School of Medicine in St. Louis

Piyush Tripathi
Washington University School of Medicine in St. Louis

Thalachallour Mohanakumar
Washington University School of Medicine in St. Louis

Helen Liapis
Washington University School of Medicine in St. Louis

Feng Chen
Washington University School of Medicine in St. Louis

See next page for additional authors

Follow this and additional works at: https://digitalcommons.wustl.edu/open_access_pubs

Recommended Citation
Rogers, Sharon A.; Tripathi, Piyush; Mohanakumar, Thalachallour; Liapis, Helen; Chen, Feng; Talcott, Michael R.; Faulkner, Chad; and Hammerman, Marc R., "Engraftment of cells from porcine islets of Langerhans following transplantation of pig pancreatic primordia in non-immunosuppressed diabetic rhesus macaques." Organogenesis.7,3. 154-162. (2011).
https://digitalcommons.wustl.edu/open_access_pubs/2537

This Open Access Publication is brought to you for free and open access by Digital Commons@Becker. It has been accepted for inclusion in Open Access Publications by an authorized administrator of Digital Commons@Becker. For more information, please contact engeszer@wustl.edu.
Authors
Sharon A. Rogers, Piyush Tripathi, Thalachallour Mohanakumar, Helen Liapis, Feng Chen, Michael R. Talcott, Chad Faulkner, and Marc R. Hammerman
Engraftment of cells from porcine islets of Langerhans following transplantation of pig pancreatic primordia in non-immunosuppressed diabetic rhesus macaques

Sharon A. Rogers,1 Piyush Tripathi,1 Thalachallour Mohanakumar,2,3 Helen Liapis,1,3 Feng Chen,1,4 Michael R. Talcott,5 Chad Faulkner6 and Marc R. Hammerman1,4,*

Departments of 1Medicine, 2Surgery, 3Pathology and Immunology, 4Cell Biology and Physiology and 5Comparative Medicine; Renal Division; Washington University School of Medicine; St. Louis, MO USA

Key words: β cell, diabetes mellitus, non-human primates, transplantation, xenotransplantation

Abbreviations: E, embryonic day; IEQ, islet equivalents; STZ, streptozotocin

Introduction

In that pigs are plentiful, and because porcine insulin works well in humans, the pig has been suggested to be a pancreas organ donor for humans with diabetes. The severity of humoral rejection due to preexisting natural antibodies effectively precludes their use as whole pancreas donors in non-human primates or humans.1-4 However, isolated islets of Langerhans (islets) can survive following transplantation into immunosuppressed non-human primates2-4 or humans with diabetes1 without initiating humoral rejection. Unfortunately, recent experience with pig to primate islet2-4 or neonatal islet3 transplantation shows that sustained insulin independence can be achieved but only through the use of immunosuppressive agents that are not approved for human use or would result in an unacceptable level of morbidity in humans.2-4

Glucose tolerance can be normalized in streptozotocin (STZ)-diabetic (type 1) LEW rats or ZDF (type 2) diabetic rats following transplantation in mesentery of pig pancreatic primordia obtained very early during organogenesis [embryonic day 28 (E28)], just after the organ differentiates and prior to the time dorsal and ventral anlagen fuse] without host immunosuppression.5-8 Cells expressing insulin and porcine proinsulin mRNA with β-cell morphology engraft in host mesentery, mesenteric lymph nodes, liver and pancreas post-transplantation.5-8 Cells originating from E28 pig pancreatic primordia engraft similarly in non immunosuppressed STZ-diabetic rhesus macaques,9 and glucose tolerance is improved post-transplantation.10

To ascertain whether engraftment of cells originating from E28 pig pancreatic primordia in rats is permissive for engraftment of the same or a similar cell component present in porcine islets from adult swine, we implanted the latter beneath the renal capsule of rats that previously had been transplanted with E28 pig pancreatic primordia in mesentery. Whole islets do not engraft. However, a population of cells originating from donor islets with β-cell morphology that express insulin and porcine
proinsulin mRNA does engraft in kidneys of rats transplanted previously with E28 pig pancreatic primordia. Here we show similar engraftment of cells originating from porcine islets using diabetic rhesus macaques as hosts.

Results

Three rhesus macaques rendered diabetic (Diab) with STZ were transplanted with pig pancreatic primordia in mesentery. Four weeks later, macaques were transplanted with 15,000–20,000 porcine islet equivalents (IEQ) per kg in the renal subcapsular space of one kidney (Diab-E28-Islets group). Eight weeks later, the macaques in the Diab-E28-Islets group were euthanized and tissues obtained for analysis. Three other STZ-diabetic rhesus macaques underwent porcine islet implantation in the subcapsular space of one kidney without prior transplantation of E28 pig pancreatic primordia (Diab-Islets group). Eight weeks later, macaques in the Diab-Islets group were euthanized and tissues obtained for analysis.

Figure 1 shows mesentery from a macaque in the Diab-E28-Islets group (Fig. 1A) and in the Diab-Islets group (Fig. 1B) immediately after euthanasia. We have shown previously that mesentery of macaques transplanted with E28 pig pancreatic primordia becomes opacified relative to mesentery of non-transplanted macaques and contains enlarged mesenteric lymph nodes. Similarly, mesentery of macaques in the Diab-E28-Islets group (Fig. 1A) is opacified relative to mesentery of macaques in the Diab-Islets group (Fig. 1B). An arrow delineates a mesenteric vessel leading to a lymph node (LN) in Figure 1A.

Figure 2 shows sections of a mesenteric lymph node from a macaque transplanted with E28 pig pancreatic primordia in mesentery, followed by porcine islets in the renal subcapsular space stained using anti-insulin antibodies (A, C and E) or control antiserum (B, D and F) or hybridized to an antisense (G) or sense (H) probe for porcine proinsulin mRNA. Arrows, positively staining cells (A, C, E and G). Scale bars, 80 μm (A and B); 25 μm (C and D); 15 μm (E–H).

Three rhesus macaques rendered diabetic (Diab) with STZ were transplanted with pig pancreatic primordia in mesentery. Four weeks later, macaques were transplanted with 15,000–20,000 porcine islet equivalents (IEQ) per kg in the renal subcapsular space of one kidney (Diab-E28-Islets group). Eight weeks later, the macaques in the Diab-E28-Islets group were euthanized and tissues obtained for analysis. Three other STZ-diabetic rhesus macaques underwent porcine islet implantation in the subcapsular space of one kidney without prior transplantation of E28 pig pancreatic primordia (Diab-Islets group). Eight weeks later, macaques in the Diab-Islets group were euthanized and tissues obtained for analysis.

Figure 1. Photographs of mesentery from rhesus macaques. (A) Rhesus macaque transplanted with E28 pig pancreatic primordia in mesentery and implanted with porcine islets in one kidney. Arrow parallels lymphatic vessel (LN, lymph node). (B) Rhesus macaque implanted with porcine islets in one kidney without prior transplantation of E28 pig pancreatic primordia. Scale bar, 2 cm.

Figure 2. Sections of a mesenteric lymph node from a macaque transplanted with E28 pig pancreatic primordia in mesentery, followed by porcine islets in the renal subcapsular space stained using anti-insulin antibodies (A, C and E) or control antiserum (B, D and F) or hybridized to an antisense (G) or sense (H) probe for porcine proinsulin mRNA. Arrows, positively staining cells (A, C, E and G). Scale bars, 80 μm (A and B); 25 μm (C and D); 15 μm (E–H).
macaque (Fig. 3B) and a kidney implanted with porcine islets without prior transplantation of E28 pig pancreatic primordia (Diab-Islets group) (Fig. 3C). As shown following implantation of porcine islets into kidneys of rats under identical conditions, a distinct, whitish, well-demarcated graft can be distinguished from the surrounding renal parenchyma (Fig. 3A, arrows). No graft can be distinguished in the non-implanted (contralateral) kidney (Fig. 3B) or in the kidney from the primate implanted with porcine islets without prior pancreatic primordia transplantation (Fig. 3C).

Figure 4 shows sections from islet-implanted kidneys of macaques in the Diab-E28-Islets group. Sections are stained using anti-insulin antibodies (Fig. 4A and C) or control serum (Fig. 4B) or hybridized to an antisense (Fig. 4D) or sense (Fig. 4E) probe for porcine proinsulin mRNA. Cells that stain for insulin are present in the subcapsular space (Fig. 4A, arrow). A high power view of a single insulin-staining cell is shown in Figure 4C (arrow). It is polygonal with a round nucleus and abundant cytoplasm, a β-cell morphology. A cell in the subcapsular space to which the antisense porcine proinsulin mRNA probe binds is shown in Figure 4D (arrow). No staining for insulin is observed in sections incubated with control antiserum (Fig. 4B). No hybridization is observed if a sense probe is substituted for the antisense probe (Fig. 4E).

Figure 5 shows sections from the contralateral (non-implanted) kidney from a macaque in the Diab-E28-Islets group stained using anti-insulin antibodies (Fig. 5A) or control serum (Fig. 5B) and from an islet-implanted kidney from a macaque in the Diab-Islets group (Fig. 5C) stained using anti-insulin antibodies. No cells that stain for insulin are present in the subcapsular space of the non-implanted (contralateral) kidney (compare Fig. 4A with 5A) or the kidney implanted with islets in the macaque not previously transplanted with E28 pig pancreatic primordia in mesentery (compare Fig. 4A with 5C).

To provide additional evidence that cells in mesenteric lymph nodes and the islet-implanted kidneys of macaques in the Diab-E28-Islets group originate from the pig, we performed fluorescent in situ hybridization using probes specific for the pig X chromosome (Fig. 6). Shown in Figures 6A and C are pig X chromosomes in nuclei of cells from a normal porcine pancreas (arrow positive controls) stained using two different probes (green A; pink C). Figures 6B and D show pig X chromosomes (arrows) in the nuclei of cells in the mesenteric lymph node (Fig. 6B) and renal subcapsular space (Fig. 6D, arrowheads). As before, in rat renal tubules, autofluorescence of macaque renal tubular cytoplasm is observed. However, pig X chromosomes are not present in macaque renal tubules.

Multiple organs were excised from a macaque transplanted with E28 pig pancreatic primordia in mesentery and, subsequently, with porcine islets in the renal subcapsular space of one kidney (Diab-E28-Islets group). Tissues were homogenized individually and total RNA purified. RT-PCR was performed using primers for pig or monkey proinsulin mRNA (intron-spanning to eliminate the possibility of false amplification from genomic DNA). Results are shown in Figure 7A. Products were sequenced to confirm identities. The pig primers amplify a band of 193 bps in RNA originating from pig pancreas, corresponding to pig proinsulin insulin mRNA. The rhesus macaque (monkey) primers amplify a band of 199 bps corresponding to monkey proinsulin insulin mRNA in monkey pancreas. Pig proinsulin mRNA is also detected in the islet-implanted monkey kidney. Multiple organs were excised from a macaque transplanted with porcine islets in the renal subcapsular space of one kidney with no prior
transplantation of E28 pig pancreatic primordia in mesentery (Diab-Islets group, and RT-PCR was performed as above. As shown in Figure 7B, no pig proinsulin mRNA was detected in any monkey organ including the transplanted kidney.

Figure 8 is an electron micrograph of the renal subcapsular space from a rhesus macaque kidney into which porcine islets were implanted a month following transplantation of E28 pig pancreatic primordia in mesentery. Shown is a cell with encapsulated granules (arrows) characteristic of endocrine secretory granules.

Figures 9A and B illustrate GTTs performed on macaques in the Diab-E28-Islets and Diab-Islets groups, respectively, prior to administration of STZ (pre-STZ) and following administration of STZ (post-STZ) immediately before transplantation. In the Diab-E28-Islets group macaques, GTTs were also performed 4 weeks following transplantation of E28 pig pancreatic primordia prior to implantation of islets (post-E28) and again 7 weeks following subsequent implantation of porcine islets (post-E28-I). In the Diab-Islets group macaques, GTTs were performed 7 weeks following implantation of porcine islets (post-islets). Multiple comparisons were performed (Bonferroni) between mean glucose levels (times 0, 15, 30 and 60) among the Diab-E28-Islets group macaques (pre-STZ, post-STZ, post-E28; and Post-E28-I) (Fig. 9A). Levels were elevated significantly post-STZ relative to pre-STZ (p < 0.001). Thus, administration of STZ rendered macaques diabetic. Levels were reduced significantly post-E28 (p < 0.001) or post-E28-I (p < 0.01) relative to post-STZ but remained elevated relative to pre-STZ (p < 0.001 for each comparison). Thus transplantation of E28 pig pancreatic primordia with or without subsequent implantation of porcine islets improved glucose tolerance but did not render it normal. Levels post-E28 were not different from levels post-E28-I. Thus, implantation of islets subsequent to transplantation of E28 pig pancreatic primordia did not improve glucose tolerance. Multiple comparisons were performed between mean glucose
levels (times 0, 15, 30 and 60) among the Diab-Islets group macaques (pre-STZ, post-STZ and post-islets) (Fig. 9B). Levels were elevated significantly (p < 0.001) post-STZ relative to pre-STZ and were further elevated post-islets relative to post-STZ (p < 0.01). Thus, implantation of porcine islets alone did not improve glucose tolerance.

To show that pig cells within transplanted diabetic macaque kidneys are capable of releasing insulin in response to glucose, we incubated slices from (1) the kidney implanted with porcine islets or (2) the contralateral kidney from one rhesus macaque into which E28 pig pancreatic primordia had been transplanted previously, for 1 h at 37°C in DMEM containing 3 mM glucose. We took aliquots immediately prior to (time 0) and following addition of glucose to incubations so as to render the glucose concentration 20 mM. As illustrated in a representative experiment using weight-matched tissue (Fig. 10A), no insulin could be detected at time 0 in supernatants from the implanted kidney. However, insulin was detectable by 1 min after increasing the DMEM glucose level, consistent with a first-phase insulin release characteristic of β cells. No insulin was detected at any time in any supernatants from the non-implanted kidney (Fig. 10B) or in any supernatants from a kidney of a macaque in which porcine islets were implanted without prior transplantation of E28 pig pancreatic primordia in mesentery (not shown).

**Discussion**

Transplantation of embryonic pancreas (primordia) to replace the function of diseased organs offers theoretical advantages relative to transplantation of either pluripotent ES cells or of fully differentiated (adult) pancreas or islets; (1) Primordia differentiate along defined organ-committed lines with no requirement to steer differentiation and no risk of teratoma formation. The glucose-sensing and insulin-releasing functions of β cells that differentiate from primordia are functionally linked; (2) The growth potential of cells within embryonic pancreas is enhanced relative to those in terminally differentiated pancreas or islets; (3) The cellular immune response to transplanted primordia obtained early during embryogenesis is attenuated relative to that directed against adult islets; (4) The ability of avascular primordia to attract a host vasculature renders them less susceptible to humoral rejection than is adult pancreas with donor blood vessels transplanted across a discordant xenogeneic barrier, and (5) Primordia differentiate selectively. In the case of E28 embryonic pig pancreas, exocrine tissue does not differentiate following transplantation, obviating complications such as the enzymatic autodigestion of host tissues.5,9,11-14

While glucose tolerance can be normalized in non-immunosuppressed diabetic rats by transplantation of E28 pig pancreatic primordia9,10 and improved in rhesus macaques following transplantation of E28 pig pancreatic primordia10 (Fig. 9A), it has proven to be more difficult to render macaques normoglycemic without an exogenous insulin requirement.9 The most likely explanation is the size difference between the species, with macaques weighing 20–30 times as much as rats.8 A STZ-diabetic rat can be rendered normoglycemic by transplantation of 5–8 pig pancreatic primordia.9 Extrapolating, it would take 100 primordia to normalize glucose tolerance in a macaque. In lieu of increasing the number of transplanted primordia, we determined whether porcine islets, a more easily obtainable and possibly more robust source of insulin-producing cells, could be substituted in animals transplanted previously with E28 pig pancreatic primordia. Our first step was to show, using rats, that prior transplantation of primordia permits engraftment of a β-cell component from porcine islets.9 Here, we document using immunohistochemistry for insulin and in situ hybridization for porcine proinsulin mRNA (Fig. 4); fluorescent in situ hybridization (Fig. 6) and RT-PCR for porcine proinsulin mRNA (Fig. 7) that pig cells engraft in macaque kidney following transplantation of E28 pig pancreatic primordia in mesentery and subsequent implantation of porcine islets in kidney. Pig cells do not engraft in kidney following implantation of islets without prior transplantation of E28 pig pancreatic primordia or in the contralateral (non-implanted) kidney (Fig. 5). Thus, cells illustrated in Figure 4 are almost certainly derived from porcine islets and not from embryonic pancreas transplants. Furthermore, the cells engrafted in macaque kidney release insulin in a glucose-dependent manner in vitro (Fig. 10A). The ELISA we used for these studies is sensitive to porcine insulin but also detects primate
insulin. While it is possible that the implanted kidney contains native cells that secrete primate insulin rather than implanted cells that secrete porcine insulin, this is rendered unlikely by the absence of any insulin in the non-implanted kidney (Fig. 10B).

In rodent models, the renal subcapsular space is the most common site for islet implantation. However, in large animals and humans, islet transplantation beneath the kidney capsule is limited due in part to the success of intraportal infusion. Islets in the portal vein release insulin into the portal vein, from which it accesses the liver directly, an advantage relative to release into the systemic circulation as occurs following renal subcapsular implantation. We utilized the renal subcapsular site in our previous study and in the present studies, because 

\[ \beta \]

cells originating from E28 pig pancreatic primordia transplanted in mesentery do not engraft in kidney, permitting the distinction between cells originating from E28 pig pancreatic primordia and porcine islets at the renal subcapsular site. Glucose tolerance in diabetic rats not fully normalized by prior transplantation of E28 pig pancreatic primordia in mesentery, was corrected following subsequent implantation of porcine islets in kidney. In contrast, such a correction was not observed following E28 pig pancreatic primordia transplantation and porcine islets implantation in macaques (Fig. 9A), consistent with previous observations that glucose tolerance is more difficult to correct in macaques than in rats. Our present experiments were designed to demonstrate engraftment of a cell component originating from porcine islets in kidneys of diabetic macaques transplanted previously with E28 pig pancreatic primordia, not necessarily to normalize glucose tolerance. It is possible that the cell component, although of sufficient mass to normalize glucose tolerance in diabetic rats following implantation of islets [5,000 IEQ in 150 g rats (approximately 33,000 IEQ per kg)], is insufficient following implantation of 15–20,000 IEQ/kg in rhesus macaques. Implantation of more islets (isolated from more than one adult pig pancreas) in kidney or, alternatively, infusion of porcine islets into the portal vein of macaques following transplantation of E28 pig pancreatic primordia in mesentery may be a better way to normalize glucose tolerance.

Though not observed under all conditions, engraftment of pancreatic progenitors transplanted across a xenogeneic barrier to non-immunosuppressed immune-sufficient hosts has been reported twice previously. Eloy et al. described normalization of glucose post-transplantation of E15 but not E18 embryonic chick pancreas into liver of non-immunosuppressed STZ-diabetic rats. Abraham et al. described successful xenograftment in multiple organs of human pancreatic islet-derived progenitor cells infused in
which is dependent on antigen transport via afferent lymphatics into the draining mesenteric lymph nodes. Harada et al. have proposed a similar co-opting of oral tolerance to explain the muted immune response in vivo and by cells from mesenteric lymph nodes in vitro to a colon carcinoma of BALB/c origin or a human CD80-transfected DBA/2 mastocytoma injected into the subserosal space of cecum in BALB/c mice relative to tumors injected subcutaneously.18

In previous studies that did not employ transplantation of E28 pig pancreatic primordia prior to implantation of porcine islets in non-human primates,19–21 complete destruction of grafts was evident within two weeks. We found no evidence that porcine islets survive post-implantation in macaque kidneys without prior transplantation of E28 pig pancreatic primordia transplantation in mesentery (Figs. 5 and 7). However, a β cell-like component (containing insulin and porcine proinsulin mRNA) originating from porcine islets engrafts if macaques are transplanted previously with E28 pig pancreatic primordia in mesentery. To our knowledge, ours is the first report describing sustained survival of such cells following transplantation of porcine islets beneath the renal capsule of non-immunosuppressed immune sufficient non-human primates. The findings establish a rationale for further investigations to ascertain whether porcine islets can normalize glucose tolerance in diabetic primates transplanted previously with E28 pig pancreatic primordia.

Methods

Animal care and induction of diabetes mellitus. Animal care followed Institutional Animal Care and Use Committee (IACUC) standards. Insulin-dependent diabetes mellitus was induced in prepubescent (2.5–3.5 kg) rhesus (Rh) macaques (Macaca mulatta) obtained from Alphagenesis Yemasee SC by infusion of STZ intravenously in a single 60 mg/kg bolus.9 Macaques had free access to water, were fed Harlan 2050 Teklad Global 20% protein diet (3.5% of their body weight per day divided in two meals, each after administration of insulin) and fresh fruit and were monitored twice daily for non-fasting blood glucose. In all diabetic macaques, blood glucose was maintained at or below 200 mg/dl using two daily Lantus insulin injections plus 1–2 units of Lispro as needed.

Isolation of pig pancreatic primordia and porcine islets and transplantation/implantation into diabetic rhesus macaques. Pancreatic primordia from 20–30 E28 Yorkshire pig embryos (Oak Hill Genetics) were isolated and transplanted between tissue planes of mesentery without suture, exactly as described previously in reference 9. Islets of Langerhans from adult female Yorkshire pigs were isolated as before8 and implanted beneath the kidney capsule. A midline abdominal incision (10–15 cm) was made and the kidney exposed, with care taken not to tear the capsule surrounding the kidney. A small incision was made in the capsule of one kidney (usually the left kidney) and a 22G catheter placed under the capsule through which all islets isolated from one adult pig pancreas (15,000 to 20,000 IEQ per kg) were infused. The capsule was closed with a single suture, and the abdomen was closed in standard fashion.

Figure 8. Electron micrograph of rhesus macaque kidney following sequential transplantation of E28 pig pancreatic primordia in mesentery and implantation of porcine islets in the kidney. Arrows, endocrine granules. Scale bar a 2 μm.

Figure 9. Intravenous glucose tolerance in rhesus macaques. Glucose in peripheral venous blood was measured in (A) three macaques in the Diab-Isets group and (B) three macaques in the Diab-Isets group before any transplantations prior to administration STZ (pre-STZ) and 5 days following administration of STZ (post-STZ). Glucose tolerance was measured in Diab-E28 Isets macaques 4 weeks post-transplantation of E28 pig pancreatic primordia (post-E28) and again 7 weeks after subsequent implantation of porcine islets (post-E28-I) and in Diab-Isets group macaques 7 weeks following implantation of porcine islets (post-Isets). Data are shown as mean ± SE (three macaques).
Glucose tolerance testing. Levels of glucose were measured using the Hemocue B-glucose Analyzer (Hemocue) in whole blood. Intravenous glucose tolerance testing (IV GTT) was performed as before. Exogenous insulin was held for 24 hours in macaques fasted for 18 hours prior to administration of glucose (0.5 g/kg) intravenously over 30 seconds as 50% dextrose. Peripheral venous blood samples were taken prior to injection (Time 0) and at several times after administration. Multiple comparisons were performed using the Bonferroni multiple comparisons test (GraphPad Instat 3). Differences were considered significant if p < 0.05 by two-tailed analysis.

Histology. Tissue fixation, sectioning, staining and photography were performed as in previous studies in reference 9. Polyclonal rabbit anti-insulin antiserum that recognizes both porcine and rhesus macaque insulin was obtained from Accurate Chemicals. Normal rabbit serum was substituted for the anti-insulin serum for control stains. Hematoxylin was used as a counterstain in Figures 2, 4 and 5.

Detection of insulin transcripts using RT-PCR or in situ hybridization. Tissues were excised, RNA was purified and reactions carried out as before. Products were separated by electrophoresis on 3% agarose gels and their identities confirmed by sequencing in the Washington University Core Protein and Nucleic Acid Chemistry Laboratory. The primers are intron-spanning to eliminate the possibility of false amplification from genomic DNA. For amplification of pig proinsulin transcripts (GenBank, AY044828), the primers used are SsInsF2: 5’-AAC CCT CAG GCA GGT GCC GGA GCA GCA-3’; SsInsR2: 5’-GGG GTG CGG GGA GCA GCA-3’, using conditions: 30 minutes at 55°C, 2 minutes at 94°C and followed by 40 cycles of 94°C, 15”; 64°C, 30”; 68°C, 25”. For rhesus macaque (monkey) proinsulin transcripts (Human Genome Sequencing Center Baylor College of Medicine: Contigs 422635 and 421061), the primers used are MmIns1F2: 5’-GAC CCT CAG GTG GGG CAG-3’; MmInsR2: 5’-AGG AGG CGG AGG GTG TGG-3’ using conditions: 30 minutes at 55°C, 2 minutes at 94°C and followed by 40 cycles of 94°C, 15”; 61°C, 35”; 68°C, 25”. The pig primers amplify a band of 193 bps in RNA originating from pig pancreas corresponding to pig proinsulin mRNA. The monkey primers amplify a band of 199 bps corresponding to monkey proinsulin mRNA in monkey pancreas RNA.

In situ hybridization was performed on 5 μm paraffin-embedded sections using digoxin-labeled antisense probes that are specific for porcine proinsulin mRNA (do not hybridize to rhesus macaque proinsulin mRNA). A sense probe was used for control stains.

Fluorescent in situ hybridization. Fluorescence in situ hybridization in paraffin-embedded tissue sections was performed exactly as in our previous studies in reference 8 and visualized using an Olympus BX61 epifluorescence microscope system.

Electron microscopy. Electron microscopy was performed by one of us (H.L.) exactly as in previous studies in reference 8.

Detection of insulin in supernatants. Kidney tissue was removed from diabetic rhesus macaques, and tissue samples were placed individually in 1 ml of Dulbecco’s modified Eagle’s medium (DMEM) containing 3 mM glucose at 37°C as before. After 1 h, glucose was added to the incubation to make the total concentration 20 mM, and 100 μl aliquots were removed beginning 1 min later. Levels of insulin in aliquots were measured using a porcine insulin-sensitive ELISA (DAKO, #K6219, sensitivity 0.5 μU/ml).

Institutional review board approval. Animal studies were approved by the Washington University Animal Studies Committee (Approval 20100212).

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Acknowledgments
Supported by George M. O’Brien Center DK079333 and grant 1-2008-37 from the Juvenile Diabetes Research Foundation (JDRF). We are grateful for the technical assistance of Nick Benshoff.