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Research Paper

Incubation of Metanephroi with Vitamin D₃ Increases Numbers of Glomeruli

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KEY WORDS

kidney, organogenesis, transplantation

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ABSTRACT

To characterize actions of vitamin D₃ on metanephroi transplanted from rat embryos to adult recipients, we incubated metanephroi with or without 0.01, 0.1 or 1 ug/ml vitamin D₃, 25-hydroxyvitamin D₃ [25(OH)D₃] or 1, 25-hydroxyvitamin D₃ [1,25(OH)₂D₃] prior to implantation. The number of glomeruli in developed metanephroi three weeks post-transplantation that had been incubated with 1.0 ug/ml vitamin D₃ was increased relative to the number in metanephroi that were not incubated with vitamin D₃ (control), an effect that was not recapitulated by administration of vitamin D₃ directly to hosts at the time of transplantation. Incubation of metanephroi with 1.0 ug/ml vitamin D₃ also enhanced inulin clearances of metanephroi measured at 12 weeks post-transplantation. The hydroxylated derivative of vitamin D₃, 25(OH)D₃, increased glomerulus number when applied at 0.01ug/ml but not at higher concentrations, while the twice-hydroxylated derivative 1,25(OH)₂D₃, failed to increase glomerulus number at any concentration tested. We conclude that incubation with vitamin D₃ prior to implantation enhances inulin clearance possibly by increasing the number of glomeruli that develop post-transplantation.

Our findings suggest the vitamin D₃ effect is mediated locally.

INTRODUCTION

We have shown that metanephroi from rat embryos transplanted into the omentum of adult rat hosts undergo growth, differentiation, and vascularization, clear inulin from the host's circulation after anastomosis between the ureter of the transplant and host¹ and can support life in otherwise anephric recipients.² Successful xenotransplantations of metanephroi have also been performed.³⁻⁵ Xenotransplantation (pig into mouse) was optimal under conditions such that renal anlagen were incubated prior to implantation in a solution that contained a number of growth factors and other growth promoting agents including vitamin D₃.⁴

Vitamin D₃, an endogenous steroid synthesized in the skin, is biologically inactive. Formation of its major biologically active product, 1, 25-hydroxyvitamin D₃ [1,25(OH)₂D₃], results from hydroxylations at position C-25 primarily in the liver and C1 α primarily in the kidney.^{6,7} Therefore, an action of vitamin D₃ per se on transplanted metanephroi would be unexpected.

To shed light on whether incubation of metanephroi with vitamin D₃ alone prior to implantation affects subsequent differentiation and function, we incubated metanephroi from rat embryos with or without vitamin D₃, 25(OH)D₃, or 1,25(OH)₂D₃ in the absence of other growth factors. Here we show that incubation with 1.0 ug/ml vitamin D₃ prior to implantation increases the number of glomeruli measured at three weeks post-transplantation, and enhances inulin clearances measured in metanephroi 12 weeks post-transplantation. The action of vitamin D₃ in vitro to increase numbers of glomeruli is not recapitulated by systemic administration of vitamin D₃ to hosts.

METHODS

Transplantation of metanephroi, measurement of inulin clearance and histology. Metanephroi were surgically dissected from embryonic day (E) 15 Sprague-Dawley rat embryos or, where indicated Lewis rat embryos, under a dissecting microscope using previously described techniques.¹ Immediately after dissection metanephroi were placed in 1 ml of a 50:50 mixture of Ham's F12:Dulbecco's modified Eagles medium at 4°C containing 25 nM prostaglandin E1, and iron-saturated transferrin (5 ug/ml) (HF12:DMEM) for 45 minutes.¹⁻⁴ When indicated, the following additions were made to the HF12:DMEM (incubation solution): none (control); 0.01, 0.1, or 1.0

ug/ml vitamin D₃ (Sigma Chemicals, St. Louis MO), 25(OH)D₃, or 1,25(OH)₂D₃ (provided by Dr. Milan Uskokovic, Hoffman-La Roche Nutley NJ); 10⁻⁷ M recombinant human insulin-like growth factor I (IGF I) (Genentech Inc. S. San Francisco CA); or 10⁻⁶ M retinoic acid (RA) (Sigma Chemicals). Metanephroi were removed from the incubation solution, and implanted in the omentum of anaesthetized six week old female (host) Sprague Dawley or

Lewis rats. During the same surgery, host rats had one kidney removed.

In some hosts, three weeks following transplantation, end-to-end ureteroureterostomy was performed using microvascular technique (interrupted 10-0 suture) between the ureter of a metanephros implanted in the omentum and the ureter of the kidney that had been removed. Nine weeks later all remaining native renal tissue (the contralateral kidney) was removed from host rats, following which inulin clearances were measured on conscious rats after placement of an indwelling bladder catheter and intravenous line exactly as in previous studies.¹ Baseline measurements for inulin were performed on urine and blood samples obtained prior to beginning the inulin infusions. These "background" values were subtracted from measurements performed after beginning the inulin infusion. Infusion was begun only following removal of all remaining native renal tissue and drainage of all urine remaining in the bladder (10–20 ul). Rats received no immunosuppression.¹

Determination of glomerulus number. Numbers of glomeruli per whole metanephros were determined using a technique described by Nathanson et al.⁸ with modifications. Briefly, whole metanephroi were incubated in 6N hydrochloric acid for 45–60 minutes at 37°C, rinsed with tap water, and stored overnight at 4°C in a volumetric flask. The kidneys were mechanically dissociated and the tubules and glomeruli suspended in water. Three 0.5 ml aliquots were taken, the glomeruli were counted under a dissecting microscope.

Multiple comparisons were made using Student- Newman-Keuls Test.

RESULTS

Initial experiments were performed to determine whether incubation of metanephroi with vitamin D₃ prior to implantation affects numbers of nephrons formed post-transplantation. To this end, we measured numbers of glomeruli in metanephroi incubated prior to implantation in DMEM:HF12 (control media) or 1.0 ug/ml vitamin D₃, RA, known to increase the number of nephrons in developing kidneys several-fold⁹ as a positive control rIGF I, an agent that has been shown to be a more modest (~20%) stimulator of nephron numbers¹⁰ as a negative control.

The number of glomeruli in metanephroi that had been incubated in control media was 4140 ± 596 or about 13% of the number present in an adult rat kidney.¹¹ Incubation with RA enhanced significantly the number of glomeruli to 9010 ± 1207, or to about 30% of normal. Incubation with vitamin D₃ had a comparable effect (9620 ± 1139). In contrast, incubation with IGF I, did not significantly increase glomerulus numbers (Table 1). Numbers of glomeruli expressed per mg metanephros weight are also significantly higher than control in metanephroi incubated with vitamin D₃ or RA (Table 1).

To exclude the possibility that incubation of metanephroi with vitamin D₃ prior to implantation increases the number of glomeruli post-transplantation by virtue of administration of vitamin D₃ to hosts per se (via implantation of vitamin D₃-incubated metanephroi), 1 ug of vitamin D₃ was administered intravenously in 1 ml of sterile saline to Sprague Dawley rat hosts concomitant with transplantation of DMEM:HF12 (control media)-incubated metanephroi. Alternatively, 1 ml of sterile saline was injected.

Table 1 Numbers of glomeruli in metanephroi

Incubation	Control	Vitamin D ₃	Vitamin A	IGF I
Number of experiments	(6)	(6)	(6)	(6)
Metanephros weight (mg)	62 ± 3.7	71 ± 5.9	52 ± 1.8	45 ± 2.0
Glomeruli	4140 ± 569	9620 ± 1139*	9010 ± 1207*	5320 ± 964
Glomeruli per mg metanephros weight	66 ± 8.0	140 ± 23**	174 ± 22**	114 ± 18

Transplantations were carried out from Lewis rat embryos to adult Lewis rats. Data are presented as mean ± SEM different than control. **p<0.05; *p<0.01.

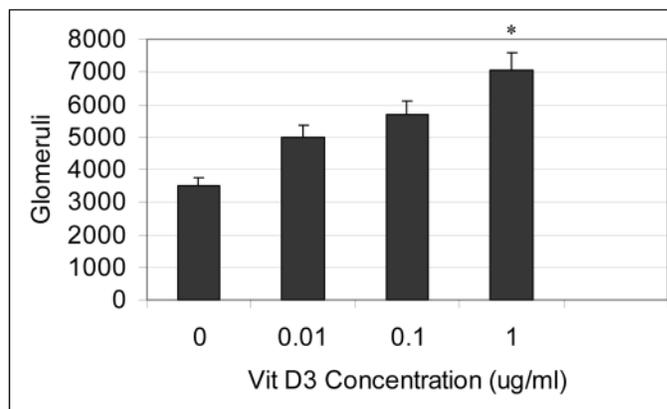


Figure 1. Numbers of glomeruli in metanephroi transplanted from Sprague Dawley rat embryos to adult Sprague Dawley rats. Metanephroi were incubated in DMEM:HF12 without vitamin D₃ (0) (n = 28) or containing 0.01 ug/ml (n = 8); 0.1 ug/ml (n = 9) and 1.0 ug/ml (n = 28) Vitamin D₃. Data are mean ± SEM. * Greater than control p < 0.005.

Table 2 Numbers of glomeruli in metanephroi

Incubation	n	Glomeruli
Control	12	3673 ± 353
25(OH)D ₃		
0.01 ug/ml	9	*7384 ± 610
0.1 ug/ml	8	6092 ± 713
1.0 ug/ml	8	3723 ± 769
1,25(OH) ₂ D ₃		
0.01 ug/ml	12	4464 ± 504
0.1 ug/ml	5	6490 ± 945
1.0 ug/ml	4	5067 ± 363

Transplantations were carried out from Sprague Dawley rat embryos to adult Sprague Dawley rats. Data are presented as mean ± SEM. *Different than control p<0.05.

There was no difference between nephron numbers in metanephroi implanted in vitamin D₃-treated rats (5725 ± 803) (n = 10) or saline-treated rats (4284 ± 396) (n = 12).

We next incubated metanephroi prior to implantation in DMEM:HF12 (control media), or HF12:DMEM containing varying concentrations of vitamin D₃ (0.01, 0.10, or 1.0 ug/ml) and counted glomeruli three weeks post-transplantation. Numbers of glomeruli were increased significantly compared to control in metanephroi that had been incubated in DMEM:HF12 containing 1.0 ug/ml vitamin D₃, but not the two lower concentrations (Fig. 1).

Table 3 Weights, urine volumes and inulin clearances of metanephroi

Incubation	Control	Vitamin D ₃
Number	(4)	(3)
Met Weight (mg)	65 ± 18	79 ± 8.0
Host Weight (g)	245 ± 8.0	270 ± 8.5
Urine vol. (ul/hour)	51 ± 7.1	128 ± 9.3*
Inulin Clearance (ul/min/100mg)	0.43 ± 0.06	1.1 ± 0.06**

Transplantations were carried out from Sprague Dawley rat embryos to adult Sprague Dawley rats. Data are presented as mean ± SEM. *Vitamin D₃>Control p<0.05; **Vitamin D₃>Control p< 0.01.

To compare actions of vitamin D₃ on glomerulus numbers with actions of hydroxylated metabolites of vitamin D₃, we counted numbers of glomeruli three weeks post-transplantation in metanephroi that had been incubated in DMEM:HF12 containing 0.01, 0.10 or 1.0 ug/ml of 25(OH)D₃ or 1,25(OH)₂D₃ prior to implantation. The number of glomeruli was increased significantly relative to control only after incubation in 0.01 ug/ml 25(OH)D₃ (Table 2).

To determine whether incubation with vitamin D₃ affects function in transplanted metanephroi, urine volumes and inulin clearances were measured at 12 weeks post-transplantation, in metanephroi that had been transplanted following incubation with vehicle or vitamin D₃. Incubation with vitamin D₃ increased both parameters (Table 3).

DISCUSSION

The kidney is a target organ for active forms of vitamin D. The pattern of intracellular 1,25(OH)₂D₃ vitamin D receptor (VDR) expression during renal ontogenesis has been defined in the rat. Starting at E15, epitopes for the VDR are found in cells of branching ureteric buds and in the surrounding mesenchyme, and at later stages, in glomerular parietal and visceral epithelial cells.¹² The VDR cannot be detected in metanephroi from E13 rats. However, beginning after 3 days in organ culture, the pattern of expression is identical to that detected *in vivo*.¹²

Presently we have shown that incubation of metanephroi with vitamin D₃ prior to implantation increases the number of glomeruli three weeks post-transplantation and enhances inulin clearance 12 weeks post-transplantation. In developed kidneys numbers of glomeruli reflect numbers of nephrons since each nephron has a single glomerulus. However, this equation cannot necessarily be made in developing metanephroi. Therefore, it is possible, but not proven that the enhanced inulin clearance reflects larger numbers of nephrons.

It is of interest that incubation of metanephroi with 0.01 ug/ml 25(OH)D₃, but not higher concentrations increases the number of glomeruli. Possibly, the biological activity of 25(OH)D₃ *in vitro* is explained by its 1 α hydroxylation in metanephroi^{13,14} to form 1,25(OH)₂D₃. However, this is unlikely because 1,25(OH)₂D₃ does not recapitulate the 25(OH)D₃ effect.

The mechanisms for these biological actions of vitamin D₃ are not defined in our studies, and the identity of the vitamin D₃ metabolite or metabolites that are biologically active is not delineated. However, since systemic administration of vitamin D₃ at the time of transplantation does not recapitulate the effect of incubation prior to

implantation, our findings suggest the effect is mediated locally, and provide a rationale for the use of vitamin D₃ in media for incubation of metanephroi prior to implantation into hosts.

References

1. Rogers SA, Lowell JA, Hammerman NA, Hammerman MR. Transplantation of developing metanephroi into adult rats. *Kidney International* 1998; 54:27-37.
2. Rogers SA, Hammerman MR. Prolongation of life in anephric rats following de novo renal organogenesis. *Organogenesis* 2004; 1:22-25.
3. Rogers SA, Hammerman MR. Transplantation of rat metanephroi into mice. *Am J Physiol* 2001; 280:R1865-R1869.
4. Rogers SA, Talcott M, Hammerman MR. Transplantation of pig metanephroi. *ASAIO J* 2003; 49:48-52.
5. Dekel B, Burakova T, Arditti FD, Reich-Zeliger S, Milstein O, Aviel-Ronen, et al. Human and porcine early kidney precursors as a new source for transplantation. *Nat Med* 2003; 9:53-60.
6. Horst RL, Reinhardt TA. Vitamin D Metabolism. In: Feldman D, Glorieux FH, Pike JW, eds. *Vitamin D*. San Diego, CA: Academic Press, 1997:13-31.
7. Kumar R. Vitamin D and the kidney. In: Feldman D, Glorieux FH, Pike JW, eds. *Vitamin D*. San Diego, CA: Academic Press, 1997:275-92.
8. Nathanson S, Moreau E, Merlet-Binichou C, Gilbert T. In utero and in vitro exposure to β lactams impair kidney development in the rat. *J Am Soc Nephrol* 2000; 11:874-84.
9. Vilar J, Gilbert T, Moreau E, Merlet-Binichou C. Metanephros organogenesis in highly stimulated by vitamin A derivatives in organ culture. *Kidney International* 1996; 49:1478-87.
10. Rogers SA, Powell-Braxton L, Hammerman MR. Insulin-like growth factor I regulates renal development in rodents. *Developmental Genetics* 1999; 24:293-8.
11. Bertram JF. Analyzing renal glomeruli with the new stereology. *Int Rev Cytol* 1995; 161:111-72.
12. Johnson JA, Grande JP, Roche PC, Sweeney Jr WA, Avner ED, Kumar R. 1 α ,25-Dihydroxyvitamin D₃ receptor ontogenesis in fetal renal development. *Am J Physiol* 1995; 269:F419-28.
13. Hosseinpour F, Norlin M, Wikvall K. Kidney microsomal 25- and 1 α -hydroxylase in vitamin D metabolism: Catalytic properties, molecular cloning, cellular localization and expression during development. *Biochimica et Biophysica Acta* 2002; 1580:133-44.
14. Yamagata M, Kimoto A, Michigami T, Nakayama M, Ozono K. Hydroxylases involved in vitamin D metabolism are differentially expressed in murine embryonic kidney: Application of whole mount *in situ* hybridization. *Endocrinology* 2001; 142:3223-30.