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Prion Protein Repeat Expansion Results in Increased Aggregation and Reveals Phenotypic Variability

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Mammalian prion diseases are fatal neurodegenerative disorders dependent on the prion protein PrP. Expansion of the oligopeptide repeats (ORE) found in PrP is associated with inherited prion diseases. Patients with ORE frequently harbor PrP aggregates, but other factors may contribute to pathology, as they often present with unexplained phenotypic variability. We created chimeric yeast-mammalian prion proteins to examine the influence of the PrP ORE on prion properties in yeast. Remarkably, all chimeric proteins maintained prion characteristics. The largest repeat expansion chimera displayed a higher propensity to maintain a self-propagating aggregated state. Strikingly, the repeat expansion conferred increased conformational flexibility, as observed by enhanced phenotypic variation. Furthermore, the repeat expansion chimera displayed an increased rate of prion conversion, but only in the presence of another aggregate, the [RNQ⁺] prion. We suggest that the PrP ORE increases the conformational flexibility of the prion protein, thereby enhancing the formation of multiple distinct aggregate structures and allowing more frequent prion conversion. Both of these characteristics may contribute to the phenotypic variability associated with PrP repeat expansion diseases.

The accumulation of misfolded proteins is a pathological characteristic common to neurodegenerative disorders such as Alzheimer’s, Parkinson’s, Huntington’s, and prion diseases. Prion diseases are unique in this group by virtue of the self-propagating and transmissible nature of the misfolded prion protein (40). The prion replicates itself by converting soluble protein into the insoluble prion conformation. Only one prion protein has been identified in mammals, PrP, and its conversion into the prion conformation causes neurodegenerative diseases (40). Intriguingly, multiple proteins that behave as prions have been discovered in fungi, but these self-propagating elements do not cause disease (49). Investigation of fungal prions has provided much support for the prion hypothesis and has introduced several new tools to study prion propagation (11).

In Saccharomyces cerevisiae, the essential translation termination factor Sup35p can propagate as a prion called [PSI⁺] (48). Cells containing the [PSI⁺] prion exhibit increased nonsense suppression (48), presumably because the prion aggregates preclude the Sup35 protein from participating as efficiently in translation termination. The increase in nonsense suppression due to the [PSI⁺] state can be observed in yeast cells harboring premature stop codons in nutritional markers, thereby allowing the prion state to be monitored phenotypically (11).

Since the function of the mammalian prion protein is unknown, it is difficult to determine how mutations in PrP affect function and phenotype. Inherited prion diseases arise as a result of mutations in the gene encoding PrP, PRNP, some of which may favor the formation of the infectious prion conformation, PrPSc (52). Intriguingly, patients harboring disease-associated PRNP mutations often present with highly variable phenotypes. Even members of one family carrying identical PRNP mutations can present with unique symptoms, including variations in age of disease onset, disease severity, and disease duration (16, 24, 27).

Mutations that result in expansion of the PrP oligopeptide repeat (ORE) domain (ORD) are associated with dominant, inherited prion diseases (52). Insertional mutations have been identified in patients that expand the number of repeats up to 14 (27). Transgenic mice containing a 14-PrP repeat expansion, Tg PrP(PG14), develop a neurological disease similar to that of humans with inherited OREs (12, 13). The ORD is thought to be dispensable for PrP to form infectious PrPSc (41), but the PrP(PG14) protein accumulates in PrPSc-like deposits that are abundant in the brains of Tg PrP(PG14) mice (12, 13). These aggregates are not infectious (14), but protein aggregates from patients harboring other PrP OREs have been demonstrated to be transmissible to primates (7). Thus, it remains unclear how PrP repeat expansions cause disease.

Conversely, how amino acid sequence changes correlate to alterations in prion propagation has been determined for some yeast prion proteins. The sequence necessary for the conversion of Sup35p into the prion state has been defined by domain mapping and mutagenesis studies (reviewed in reference 54). The prion-forming domain (PFD) has been localized to the N terminus of Sup35p and is necessary and sufficient for prion propagation (30, 46). The PFD has a strikingly high percentage of glutamine (Q) and asparagine (N) amino acids, and mutation of certain Q or N residues eliminates [PSI⁺] (17). Interestingly, one region in the Sup35p PFD is strikingly similar to a region in PrP, the ORD (6). Sup35p contains 5½ OREs, and the ORD of PrP contains five octapeptide repeats. Deletion of one or more Sup35p repeats prevents efficient propagation of...
Yeast cells were grown and manipulated by standard techniques (22). A

\[
\text{GGCGC} \\
\text{GGCGC}
\]

\[
\text{PSI}
\]

in humans. \[RNQ^+\] Additional experiments demonstrated that the 14-PrP ORE could replace the N-terminal Q/N-rich region of the Sup35p PFD and successfully maintain and propagate a prion. Taken together, our results indicate that the ORE of PrP is more prion competent and enhances phenotypic variability compared to the wild-type PrP repeat length. We suggest that these properties may contribute to the variability associated with inherited prion disorders in humans.

**MATERIALS AND METHODS**

**Strain construction.** All of the yeast strains used in this study were derivatives of 74-D694 (MATa or MATa ade1-14 trpl2-289 his3A200 ura3-52 leu2-3,112) (10). Yeast cells were grown and manipulated by standard techniques (22). A 74-D694 [PSI+] diploid with one copy of SUP35 replaced with a kanamycin resistance cassette (sup35::KanMX4) was transformed with a plasmid containing SUP35 (pYK810) (38). Haploid progeny cells containing pYK810 and sup35::KanMX4 were obtained (74-D694 pYK810) from the diploid. The isogenic [psi+] 74-D694 pYK810 strain was created by growth on rich medium (YPD) (22) containing 3 mM guanidine hydrochloride (GdHCl). The plasmid shuffle technique was used to create strains expressing only the Sup35p-PrP chimera (SP5, SP14, and P14MC strains). To obtain SP14/SP35 diploid strains, [SP14+] ∆sup35 cells were mated to wild-type [psi+] cells. The resulting progeny cells showed a 2:2 segregation of the spores harbored the SP14 plasmid. The absence of the plasmid in spores containing SUP35p, but only in the presence of \[RNQ^+\] strains, was determined by color and curability by GdHCl and compared to the total number plated. To determine the prion conversion of [psi+] · [RNQ+] strains, cells were resuspended and plated on YPD plates. The percentage of red colonies was determined, and the n-fold change in stability was calculated relative to the most stable strain (strong [SP14+]). The data presented represent three independent experiments in which ~15,000 colonies were tested per strain variant. To determine the prion conversion of [psi+] · [RNQ+] strains, cells were resuspended, plated on YPD, and analyzed for the number of pink colonies compared to the total number plated. All [PRION+] cells were verified by curability by GdHCl.

**Protein analysis.** Sedimentation properties were analyzed as described previously with the addition of the mini EDTA-free protease inhibitor cocktail (Roche) to the ST buffer (36). Large aggregates were separated by sedimentation centrifugation (SDD-AE) as previously described (36), with the addition of the mini EDTA-free protease inhibitor cocktail (Roche), and the lysates were incubated in sample buffer for 7 min at room temperature before electrophoresis. Sup35p and Sup35p-PrP chimera were detected with a rabbit polyclonal antibody against the middle region of Sup35p that is maintained in all chimeras, anti-Sup35p (39). PFD-GFP microscopy. Cells were transformed with a copper-inducible plasmid containing either an SPS-PFD-GFP or an SP14-PFD-GFP fusion, and fluorescence microscopy was performed after expression of the fusion protein was induced in log-phase cells with 50 μM copper for 1 h (39).

**Curing by overexpression of Hsp104p.** Hsp104p overexpression was induced from a galactose-inducible plasmid expressing Hsp104p (pYS-Gal104) by growth in minimal medium lacking uracil (liquid or plates) supplemented with 2% galactose and 1% raffinose, and final colony color was assessed on YPD (10). **Protein transformation.** (i) Spheroplasting of cells. 74-D694 [psi+] cells were grown to an OD600 of ~0.5 and harvested. The cell pellet was washed successively with 10 ml of water, 10 ml of 1 M sorbitol, and 1 ml of SCE buffer (1 M sorbitol, 10 mM EDTA, 10 mM dithiothreitol, 100 mM sodium citrate, pH 5.8). The cells were spheroplasted in 500 μl of SCE buffer with lyticase (in sodium citrate, pH 5.8) for 30 to 40 min at 30°C. After spheroplasting was complete, cells were carefully washed twice with 1 ml 1 M sorbitol and twice with 1 ml SCE buffer (1 M sorbitol, 10 mM Tris, pH 7.5), and resuspended at 800 × g for 1 min at 4°C. Spheroplasted cells were resuspended in 1 ml of SCE buffer at 4°C.

(ii) Prion particle preparation. [PRION+] and [psi+] cells were grown to an OD600 of ~0.5 and harvested. Cell pellets were washed successively with 10 ml of 1 M sorbitol, and 1 ml of SCE buffer (1 M sorbitol, 10 mM EDTA, 10 mM dithiothreitol, 100 mM sodium citrate, pH 5.8). The cells were spheroplasted in 500 μl of SCE buffer with lyticase (in sodium citrate, pH 5.8) for 30 to 40 min at 30°C. After spheroplasting was complete, cells were carefully washed twice with 1 ml 1 M sorbitol and twice with 1 ml SCE buffer (1 M sorbitol, 10 mM Tris, pH 7.5), and resuspended at 800 × g for 1 min at 4°C. Spheroplasted cells were resuspended in 1 ml of SCE buffer at 4°C.
of water, 10 ml of 1 M sorbitol, and 1 ml of SCE buffer. Cells were resuspended in 600 μl of SCE buffer (containing 1 mM phenylmethyisulfonl fluoride and protease inhibitor cocktail [Sigma Chemical Co.]) and lysed with 300 μl of sterilized glass beads by vortexing 10 times for 10 s each at 4°C. The crude lysate was centrifuged twice at 4°C for 5 min at 800 × g and 1,000 × g. The protein concentration of the supernatant was determined with the Bio-Rad protein assay (Sonic Dismembrator; Fisher Scientific).

(iii) Transformation procedure. The transformation mixture contained 5 μl of pRS316 DNA (~300 ng/μl), ~300 μg of sonicated protein lysate, 10 μl of carrier DNA (10 mg/ml), 150 μl of spheroplast cells, and 5 volumes of PEG 8000 buffer (20% [wt/vol] PEG 8000, 10 mM CaCl₂, 10 mM Tris, pH 7.5) and was incubated for 45 min at 25°C. The transformation mixture was washed at 1,000 × g and resuspended in 150 μl of SOS buffer (1 M sorbitol, 7 mM CaCl₂, 0.25% yeast extract, and 0.5% Bacto peptone supplemented with 0.3 μg of all of the amino acids in which the yeast strain is deficient) and incubated for 30 min at 30°C. The SOS-cell mixture was plated on SD-ura/sorbitol plates (16.4% sorbitol, 3% glucose, 2% agar, complete supplement mixture without uracil, 0.67% yeast nitrogen base without amino acids) overlaid with top agar (1.2 M sorbitol, 2.5% agar, 2% glucose, complete supplement mixture without uracil, 0.67% yeast nitrogen base without amino acids). The plates were incubated at 30°C for 5 days. Transformants were spotted onto YPD, SD-ade, and 3 mM GdHCl media to score the colonies for the prion state.

RESULTS

Chimeric Sup35-PrP proteins form prions. To test the effects of the repeat expansion of PrP in a genetically tractable system, we expressed SUP35-PrP chimeras in yeast (Fig. 1). We replaced the ORD of Sup35p with the wild-type number of 5 PrP repeats and an expanded length of 14 PrP repeats (Fig. 1A). The chimeric molecules, termed SP5 (for Sup35-PrP 5 repeats) and SP14 (for Sup35-PrP 14 repeats), replaced the wild type by plasmid shuffle and were expressed as the only copy of SUP35. Since SUP35 is essential in yeast, this approach allowed us to evaluate the functionality of the chimeras in the absence of wild-type Sup35p. The expression of the chimeric proteins was similar to endogenous Sup35p protein levels, as determined by Western blotting (Fig. 1B). A control plasmid with wild-type SUP35 (terned pSup35) was generated and used to ensure that the episomal expression of Sup35p mimicked that of chromosomal Sup35p.

To test if the chimeric proteins could propagate the prion state, each chimera was introduced into a [PSI+] strain. The strain contains the ade1-14 allele, which harbors a premature stop codon that is read through in [PSI+] cells, thereby producing full-length, functional Ade1p. Thus, [PSI+] cells with the ade1-14 allele are adenine prototrophs and appear as light pink colonies on a rich medium (YPD). In contrast, in [psi−] cells, Sup35p is soluble and functional and therefore faithfully terminates translation. As such, [psi−] cells that harbor the

FIG. 1. Chimeric Sup35-PrP proteins demonstrate yeast prion properties. (A) Schematic depiction of Sup35p and PrP highlighting the consensus sequence of each ORD. PrP(PG14) contains nine ad-

(D) Cell lysates of the chimeras were subjected to ultracentrifugation, and the chimeric proteins in the total (T), supernatant (S), and pellet (P) fractions were analyzed by Western blotting. (E) [SP5+], [SP5−], [SP14+], and [sp14−] cells expressing the corresponding PFD-GFP constructs were analyzed by fluorescence microscopy. (F) Prion particles isolated from [SP5+] and [SP14+] cells can convert [prion−] cells. Protein harvested from [sp5−] or [sp14−] cells was transformed into [sp5+] or [sp14+] cells, respectively (top, [prion−] → [prion+]). Protein harvested from [SP5+] or [SP14+] cells was transformed into [sp5+] or [sp14+] cells, respectively (bottom three lines, [PRION−] → [PRION+]). Individual isolates were spotted onto YPD and SD-ade, cured on YPD with 3 mM GdHCl, and subsequently spotted onto YPD (post GdHCl).
allele are adenine auxotrophs and colonies appear red on YPD. If SP5 or SP14 protein is able to propagate the prion state, then cells containing these chimeric proteins will display a nonsense suppression phenotype in the absence of wild-type Sup35p and remain pink on YPD. Both SP5- and SP14-containing cells remained pink on YPD and grew on SD-ade medium after replacement of wild-type Sup35p in a [PSI⁺] strain (Fig. 1C), suggesting that these chimeric proteins maintain the prion phenotype. We refer to the prion states of SP5 and SP14 as [SP5⁺] and [SP14⁺], respectively. To determine if the [SP5⁺] and [SP14⁺] phenotypes could be eliminated (cured) in a manner similar to that of [PSI⁺], the cells were grown on YPD medium containing 3 mM GdmHCl. The [SP5⁺] and [SP14⁺] cells became red after growth on medium containing GdmHCl (Fig. 1C) and could no longer grow on SD-ade medium (data not shown). Moreover, when the chimeras were assessed as the only copy of SUP35 in a [psi−] strain, the cells remained red and could not grow on SD-ade medium (data not shown), demonstrating that both chimeras are functional in translation termination. Since the PrP repeat expansion consisting of 14 PrP repeats is pathogenic in humans, we asked if other pathogenic PrP ORD insertions could maintain prion properties in chimeras. Chimeras containing 8 and 11 PrP repeats were created and assayed. These chimeras also behaved phenotypically, biochemically, and genetically as yeast prions (data not shown). Since all of the chimeras were able to maintain the prion state, we analyzed SP5 and SP14 further to determine the effects of repeat expansions on prion properties.

The [SP14⁺] cells displayed more robust growth on SD-ade medium and a lighter pink colony color on YPD in comparison to [SP5⁺] cells (Fig. 1C). This suggests that the repeat-expanded SP14 protein maintains a stronger prion than SP5. Interestingly, cells with wild-type SUP35 can harbor strain variants of the [PSI⁺] prion that display heritable differences in the strength of nonsense suppression without any alteration in amino acid sequence (21). These changes in nonsense suppression can be phenotypically distinguished in yeast cells harboring the ade1-14 allele as different shades of pink on YPD and different growth rates on SD-ade medium (Fig. 1C) (50). The differences in nonsense suppression between [SP5⁺] and [SP14⁺] suggest that the chimeras are propagating structurally distinct prion strain variants.

The aggregation and resulting insolubility of Sup35p are defining characteristics of the [PSI⁺] prion state (48). To examine if the chimeric [SP5⁺] and [SP14⁺] prion phenotypes are associated with insoluble protein aggregates, we subjectcd lysates from [SP5⁺], [sp5⁻], [SP14⁺], and [sp14⁻] cells to ultracentrifugation to determine the fractionation pattern of the chimeric proteins. [SP5⁺] cells contained chimeric protein in both the supernatant and pellet fractions, while [sp5⁻] cells had most the protein in the supernatant (Fig. 1D). The presence of soluble protein in the [SP5⁺] prion cell was not unexpected, given the weak nonsense suppression phenotype observed (Fig. 1C). Lysate from [SP14⁺] cells, however, showed all of the chimeric protein in the pellet fraction, consistent with the stronger nonsense suppression phenotype, and lysate from [sp14⁻] cells displayed all of the protein in the supernatant (Fig. 1D).

To further investigate the aggregation of the chimeric prion proteins, the cellular distribution of the chimeric proteins was monitored by GFP fluorescence. An inducible PFD-GFP construct specific for each repeat region (SP5-PFD-GFP or SP14-PFD-GFP) was expressed in the corresponding [PRION⁺] and isogenic [prion−] cells. Cells containing [SP5⁺] prions displayed aggregates and diffuse background fluorescence, whereas [sp5⁻] cells showed only diffuse fluorescence (Fig. 1E). [SP14⁺] cells showed primarily a punctate fluorescence pattern, whereas [sp14⁻] cells displayed diffuse fluorescence. Taken together, these data demonstrate that the chimeric proteins assume an aggregated state in cells displaying a nonsense suppressor phenotype and both traits are reversed by curing.

In order to further demonstrate the prion nature of the SP5 and SP14 chimeras, we conducted protein transformations (25, 43). Protein harvested from [SP5⁺] and [SP14⁺] strains was transformed into [sp5⁻] and [sp14⁻] cells, respectively. As expected, protein obtained from [PRION⁺] cells, but not [prion−] cells, was able to convert [prion−] cells to [PSI−/H11001] (Fig. 1F). Together, these results demonstrate that the SP5 and SP14 chimeras behave as yeast prions.

**Chimeric prions display enhanced instability.** To determine the mitotic stability of the chimeric prions, [SP5⁺], [SP14⁺], and [PSI⁺] cells were plated on YPD and the colonies were scored as solid red, pink, or sectoring. Wild-type strong (s) [PSI⁺] cells never displayed sectoring colonies, and less than 0.02% of the colonies spontaneously appeared red (Table 1). In contrast, approximately 85% of both [SP5⁺] and [SP14⁺] colonies sectorcd. This suggests that the heritable stability of the chimeric prions is much lower than that observed with [PSI⁺] cells. Since the ORD of PrP is known to bind copper (33), we tested whether the addition of copper to the medium influenced mitotic stability and found no change (data not shown). A striking difference between [SP5⁺] and [SP14⁺] cells was noted when the nonsectoring populations of cells were analyzed (Table 1). All nonsectoring [SP5⁺] colonies were red, indicating loss of the prion state. In contrast, most of the nonsectoring population of [SP14⁺] colonies was light pink, indicating maintenance of the prion state. Thus, although the SP5 protein propagates as a prion, the maintenance of the prion state is enhanced with the repeat expansion.

<table>
<thead>
<tr>
<th>Cell type</th>
<th>Avg % of sectoring colonies ± SD</th>
<th>Avg % of nonsectoring colonies ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>s [PSI⁺]</td>
<td>0</td>
<td>&lt;0.02</td>
</tr>
<tr>
<td>[SP5⁺]</td>
<td>85.7 ± 3.8</td>
<td>14.2 ± 3.8</td>
</tr>
<tr>
<td>[SP14⁺]</td>
<td>84.1 ± 4.8</td>
<td>1.8 ± 3.1</td>
</tr>
</tbody>
</table>

**TABLE 1. Mitotic stability of the [PRION⁺] state in the chimeras**
Strain variants of [SP14+] show a distinct change in mitotic stability. Weak [PSI+] strain variants are mitotically unstable in comparison to strong [PSI+] strain variants (21, 26). To explore the possibility that [SP14+] meiotic progeny shared this characteristic of prion strain variants, we examined the mitotic stability of the chimeric prions in the SP14 progeny. SP14-containing progeny displaying a strong and a weak nonsense suppression phenotype, in addition to the original [SP14+] isolate, were analyzed for prion loss (the appearance of red colonies). Cells were plated on YPD, and the number of solid red colonies was scored as a percentage of the total. The weak [SP14+] strain variant had almost a 20-fold higher prion loss compared to the strong strain variant (Fig. 2C). These data show that the weak [SP14+] strain variant displaying less nonsense suppression is mitotically less stable than the strong [SP14+] strain variant.

Since strain variants of [PSI+] and PrPSc are due to structurally distinct aggregates (15, 28, 44), we hypothesized that weak and strong yeast prion strain variants might show differential transmission of their respective structures onto soluble proteins with different amino acid sequences. Therefore, we tested whether each isolated strain variant could transmit its prion state to soluble wild-type Sup35p or soluble SP14 proteins. Strong and weak [PSI+] cells were mated to [psi−] and [sp14−] cells. Diploids were obtained, and tetrads were dissected. As expected, analysis of the tetrads showed that both strong (s) and weak (w) [PSI+] could transmit to [psi−] and convert Sup35p to [PSI+] (Table 2). However, only strong [PSI+] could transmit prion properties to [sp14−] to generate [SP14+] cells. The capacity of strong [PSI+] to transmit prion to [sp14−] cells is lower than the strong [SP14+] strain variant. In contrast, two independent weak [SP14+] strain variants could transmit the prion state to [sp14−] cells but could not transmit the prion

in the progeny. The tetrads were also spotted onto selective medium to follow the 2:2 segregation of the sup35 deletion and the presence of SP14. Spores expressing SP14 protein displayed phenotypic variability, as shown in the example tetrad (Fig. 2B): one light pink colony and one dark pink colony (YPD). The light pink SP14 colony grew better on SD-ade medium than the dark pink colony. Spores containing wild-type SUP35 also inherited the prion phenotype but showed a high degree of instability, as both pink and red colonies were seen on YPD. The loss of nonsense suppression in the progeny expressing wild-type Sup35p suggests inefficient templating of the [SP14+] prion structure onto the Sup35 protein. However, once a single pink colony containing wild-type Sup35p was purified by restreaking, a stable [PSI+]–dependent nonsense suppression phenotype was established (data not shown). Different strengths of nonsense suppression in [PSI+] cells are observed in prion strain variants (Fig. 1C, top two rows). The SP14-containing progeny displayed differential nonsense suppression phenotypes, suggesting that the SP14 protein propagates distinct strain variants in [SP14+] cells.

strains (data not shown). These results demonstrate that [SP14+] does indeed behave as a prion genetically. To determine if [SP14+] could transmit the prion phenotype to wild-type Sup35p, [SP14+] cells were crossed to [psi−] cells. The resulting diploid and meiotic progeny were analyzed for the prion phenotype. [SP14+] cells, [psi−] cells and SP14/SUP35 diploid cells were spotted onto YPD and SD-ade media to assess nonsense suppression (Fig. 2B). The SP14/SUP35 [PRION+] diploid grew on SD-ade medium but displayed both pink and red colonies on YPD, suggesting inefficient inheritance of the nonsense suppression phenotype. Tetrads from the SP14/SUP35 [PRION+] diploid were spotted onto YPD and SD-ade media to assess inheritance of the prion phenotype

FIG. 2. [SP14+] prions display non-Mendelian inheritance and mitotic instability. (A) A representative tetrad from a cross of [SP14+] to [sp14−] illustrated epigenetic inheritance of the prion phenotype. (B) [SP14+], weak (w) [PSI+], [psi−], an SP14/SUP35 [PRION+] diploid (containing one copy of SP14 and one copy of SUP35), and a tetrad from the SP14/SUP35 [PRION+] diploid were spotted onto YPD and SD-ade media. The relative strength of nonsense suppression in the progeny is described as strong (s), medium (m), or weak (w). The SUP35 allele expressed in the progeny is indicated parenthetically. (C) The original isolate of [SP14+] and phenotypically weak and strong [SP14+] variants were plated on YPD and analyzed for prion loss (red colonies). Prion loss was calculated as the change compared to the most stable [SP14+] variant.
TABLE 2. Meiotic transmission of [SP14+] strain variants

<table>
<thead>
<tr>
<th>Cell type</th>
<th>Growth on SD-adea</th>
<th>Transmission to [psi+]</th>
<th>Transmission to [sp14+]</th>
</tr>
</thead>
<tbody>
<tr>
<td>s [PSI+]</td>
<td>++++</td>
<td>Yes</td>
<td>Yesa</td>
</tr>
<tr>
<td>w [PSI+]</td>
<td>+</td>
<td>Yes</td>
<td>No</td>
</tr>
<tr>
<td>s [SP14+] v1</td>
<td>++++</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>w [SP14+] v1</td>
<td>++</td>
<td>No</td>
<td>Yes</td>
</tr>
<tr>
<td>s [SP14+] v2</td>
<td>++++</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>w [SP14+] v2</td>
<td>++</td>
<td>Rare</td>
<td>Yes</td>
</tr>
</tbody>
</table>

a Weak [PSI+] and strong [PSI+] also show this difference in transmission to [sp14+].

b ++++, very good; ++, good; +, fair.

Hence, in contrast to its curing effect on wild-type [PSI+] cells, Hsp104p overexpression stabilized the [SP14+] prion phenotype. Therefore, we asked if the [SP14+] aggregates were dramatically altered by the overexpression of Hsp104p. Protein from [SP14+] cells subjected to overexpression of Hsp104p was analyzed by SDD-AGE and Western blotting (Fig. 3C). Neither the size nor the range of the aggregates from [SP14+] cells changed following the overexpression of Hsp104p. However, the amount of monomeric protein was decreased in the presence of excess Hsp104p. Taken together, these data suggest that the overexpression of Hsp104p stabilizes the [SP14+] phenotype without changing the size of the observable protein aggregates. Given the broad range of aggregate sizes and the ability of SP14 to maintain multiple independent structures in a cell, we hypothesize that the stabilization of the [SP14+] prion could occur because Hsp104p is disaggregating the more unstable aggregates in the [SP14+] prion population (47). Indeed, it remains unclear how these aggregates observed by SDD-AGE relate to the prion phenotype.

FIG. 3. [SP14+] contains large aggregates which are unaffected by prion stabilization. (A) Protein aggregates and monomeric Sup35p from wild-type [PSI+] (strong [s] and weak [w]) and [SP5+] and [SP14+] (original isolate, weak and strong) strains were separated by SDD-AGE and detected by Western blotting. (B) Wild-type [PSI+] and [SP14+] cells with and without the overexpression (OE) of Hsp104p plated on YPD. (C) Protein aggregates and soluble protein (monomer) from lysates of [SP14+] cells with (+) or without (−) Hsp104p overexpression were separated by SDD-AGE and detected by Western blotting.
[\textit{psi}] strains have been characterized in the context of the prion state, but the role of the repeat expansion in altering prion properties is not well understood.

\textbf{Expanded PrP repeats can replace the Sup35p PFD.} We observed that the expanded repeat domain in \textit{Sup35p} enables the protein to initiate and propagate many structural conformations, thereby suggesting that the repeat expansion confers enhanced structural flexibility to PrP. Both the PFD of \textit{Sup35p} and the N terminus of PrP, including the ORD, are highly conserved and are important for the maintenance of prion properties. Strong [\textit{psi}'] prions are associated with the ORD, while weak [\textit{psi}'] prions are not.

\textbf{Expanded \textit{PSI}^+ strain variants interconvert readily.} Our genetic analysis suggests that strain variants of the chimeric [\textit{SPI}^+] strain (data not shown). We isolated [\textit{SP5}], which is enhanced by [\textit{RNQ}'], and [\textit{SP14}] variants expressing different levels of \textit{PSI}^+ and \textit{RNQ}'. However, the spontaneous conversion of [\textit{SP5}] and [\textit{SP14}] was not enhanced in comparison to the [\textit{RNQ}'] prion state. Intriguingly, this result differed from that of the wild-type \textit{Sup35p} repeat expansion with a dramatic increase in frequency of prion conversion (31).

Given this difference, we set out to explore extragenic factors that may play a role in conversion efficiency. We found that the spontaneous conversion of [\textit{RNQ}'] in the prion state (18, 20, 37) is not necessarily associated with continued propagation of [\textit{PSI}^+] (20). We determined that the [\textit{RNQ}'] prion was not required for continued propagation of the chimeric prions [\textit{SP5}] and [\textit{SP14}] by expressing the chimeras in a \textit{Δrnq1} strain (data not shown).

Since we initially assessed spontaneous conversion in cells that were [\textit{rnq}'] (Fig. 4B), we examined the impact of the [\textit{RNQ}'] prion on the spontaneous conversion of the chimeric proteins. Unlike the induction of [\textit{PSI}^+], which is enhanced by [\textit{RNQ}'], we found that the spontaneous conversion of [\textit{psi}'] to [\textit{PSI}^+] is not affected by [\textit{RNQ}'] (~1 in 10^6 cells in both [\textit{rnq}'] and [\textit{RNQ}'] cells). However, the spontaneous conversion of [\textit{sp5}] and [\textit{sp14}] both [\textit{RNQ}'] cells to the [\textit{PRION}'] state was markedly enhanced (Fig. 4C). This suggests that the repeat expansion does influence the spontaneous prion conversion of the chimeric protein, but only in the presence of another aggregate, [\textit{RNQ}'].
P14MC protein in the prion state, [P14MC⁺⁺], and nonprion state, [p14MC⁻⁻], were grown on YPD, SD-ade, and medium containing GdHCl to assess nonsense suppression and curing (Fig. 5A). [P14MC⁺⁺] cells were pink on YPD and grew well on SD-ade medium, suggesting that the protein maintains the nonsense suppression phenotype in the prion state. Conversely, [p14MC⁻⁻] cells were red on YPD and could not grow on SD-ade medium, suggesting that the protein is functional in the nonprion state.

We next analyzed the biochemical properties of the P14MC protein. Cell lysates were subjected to ultracentrifugation to determine the aggregation state of the protein in [P14MC⁺⁺] and [p14MC⁻⁻] cells. Most of the protein from [P14MC⁺⁺] cells was present in the pellet fraction, whereas that from [p14MC⁻⁻] cells was in the supernatant (Fig. 5B). Thus, the nonsense suppression phenotype observed in the [P14MC⁺⁺] cells correlates with the aggregation of the P14MC protein. In addition, curing [P14MC⁺⁺] cells concomitantly resolubilized the protein and reversed the nonsense suppression phenotype (Fig. 5A and B). In agreement with the sedimentation results, expression of SP14-PFD-GFP showed a distinct punctuate fluorescence pattern in [P14MC⁺⁺] cells and diffuse fluorescence in [p14MC⁻⁻] cells (Fig. 5C). To further examine the aggregates, chimeric Sup35p from [P14MC⁺⁺] was analyzed by SDD-AGE and Western blotting. Protein aggregates from [P14MC⁺⁺] cells were larger than those from wild-type strong or weak [P5⁺] cells (Fig. 5D) and resembled aggregates from [SP5⁺⁺] and [SP14⁺⁺] cells (Fig. 3A). Furthermore, this assay demonstrated that [p14MC⁻⁻] cells contain only monomeric P14MC protein. These results suggest that the P14MC protein biochemically behaves as a prion.

Next, we determined if the [P14MC⁺⁺] prion aggregates could be inherited in an epigenetic manner, as is characteristic of yeast prions. We mated [P14MC⁺⁺] to [psi⁻⁻] (data not shown), [p14MC⁺⁺], [sp14⁺⁺], or [sp5⁻⁻] cells, obtained diploids, and dissected tetrads. The meiotic progeny all showed a 4:0 nonsense suppression phenotype, indicating that the [P14MC⁺⁺] prion was transmitted in an epigenetic fashion to the chimeras (Fig. 5E) but displayed a slightly weaker nonsense suppression phenotype than [SP14⁺⁺] (Fig. 5E; also compare SD-ade panels in Fig. 1C and 5A). In addition, protein obtained from [P14MC⁺⁺] cells converted [p14MC⁻⁻] cells to [P14MC⁺⁺] after protein transformation (data not shown). We also examined if the lack of the Q/N region in the P14MC protein affected the stability of the [P14MC⁺⁺] prion in comparison to [SP14⁺⁺] cells (Fig. 5F). [P14MC⁺⁺] cells plated on YPD displayed both red and sectoring colonies, suggesting that the change in the context of the 14-PrP repeats did not alter the stability of the chimeric prion, as [SP14⁺⁺] cells also showed red and sectoring colonies (Table 1). Taken together, these results indicate that 14-PrP repeats can replace both the N-terminal Q/N region and the ORD of Sup35p and still maintain prion competence. While the PFD of wild-type Sup35p has a striking 44% Q/N content, this prion-competent P14MC PFD contains only 18% Q/N. This suggests that the structural requirements for a yeast prion protein are not limited by a critical percentage of Q/N residues and that there may be other ways to achieve the structural flexibility required for yeast prion propagation.

**DISCUSSION**

Here, we describe a novel model system using a chimeric yeast-mammalian prion to evaluate the functionality of the PrP
repeats in the context of the yeast prion protein Sup35p. Precise replacement of the entire repeat domain of Sup35p with the PrP ORD resulted in a tractable system to evaluate the effect of the PrP repeat expansion. The wild-type number of PrP repeats and all ORD expansions in the Sup35p-PrP chimeras behaved phenotypically, biochemically, and genetically as prions, although the PrP repeats did confer unique characteristics on the resulting prions. SP14 displayed a stronger prion phenotype in comparison to SP5, suggesting that the addition of octapeptide repeats allowed for a more stable self-replicating structure. This resembles what is seen in human patients with PrP repeat expansions, considering that the repeat expansion is associated with aggregation and spontaneous development of prion disease (16). The 14-PrP repeats also facilitated the establishment and propagation of prion strain variants, as indicated by differences in phenotype, mitotic stability, and prion transmission. Furthermore, in the presence of [RNQ] prion, SP14 converted into the prion form more readily than SP5. Strikingly, the 14-PrP repeats replaced the Q/N-rich region and the repeat region of Sup35p and maintained prion properties. Our data suggest that the PrP repeat expansion can influence prion conversion and enhance the formation of multiple aggregate structures.

Strain variants of both yeast prions and PrPSc are believed to be composed of structurally unique self-propagating aggregates (15, 25, 28, 43). Although conformationally distinct variants have been associated with PrPSc strains (2, 3), they have not been associated with inherited PrP mutations. The phenotypic variability associated with some pathogenic point mutations in PrP has been attributed to changes in glycoform ratio, but this did not correlate with the repeat expansion mutations (23). We suggest that the phenotypic variability observed in inherited repeat expansion prion diseases could result, in part, from differences in the structures acquired by the mutant protein.

Our data show that the repeat-expanded chimera displays phenotypic variation reminiscent of prion strains. The phenotypically distinct [SP14] strain variants exhibited different mitotic stabilities (Fig. 2C) and differential transmission to imperfect prion protein sequences (Table 2). Taken together, these data suggest that the prions within these variants possess unique characteristics that might be structural, although the nature of the structural differences could not be elucidated. To date, the precise nature of the structural differences in mammalian and yeast prion strain variants also has not been described. A unique feature of the [SP14] strain variants was their enhanced ability to interconvert. Strong variants could spontaneously give rise to weak variants, and vice versa (Fig. 4A). This observation suggests that the SP14 protein is not entirely committed to one particular prion conformation. There are two possibilities of how the interconversion could arise. First, the [SP14] cells could maintain multiple structures and distinct phenotypes emerge when they separate and multiply (upon restreaking). The phenotype observed in one colony need not indicate structural homogeneity of the chimeric protein, but rather the colony might only show the phenotype of the dominant structure. Second, the SP14 protein may spontaneously create new prion aggregates when the prion state is lost, and these need not be the same strain variant. The influence of [RNQ] could then contribute to interconversion via reacquisition. In the absence of the [RNQ] prion, the [sp5] and the [sp14] cells did not spontaneously convert to the [PRION] state more readily than [ps5] converts to [PS1] (Fig. 4B). However, in the presence of [RNQ], the spontaneous conversion of the chimeric proteins was greatly enhanced (Fig. 4C). This result is similar to that obtained with the Sup35p repeat expansion, which also had a higher frequency of spontaneous conversion in comparison to wild-type Sup35p (31). We have found that the previously described Sup35p repeat expansion (31) also converted more frequently to [PS1] only in the presence of the [RNQ] prion (E. M. H. Tank and H. L. True, unpublished data).

In vitro studies suggest that the repeat expansion in PrP decreases the lag phase of amyloid formation (34) and increases the accessibility of the N terminus of the mutant protein (53), thereby allowing a quicker conformational change into a pathogenic state. Our in vivo model indicates that the spontaneous conversion of the repeat expansion protein to an aggregated state need not be the sole mechanism whereby these mutations affect structure and, ultimately, disease. Instead, the repertoire of conformations achieved by the 14-PrP repeat chimeric protein appeared to be increased. However, in the presence of an additional aggregate, the conversion rate of SP14 was considerably enhanced. Therefore, other misfolded proteins may act as a potential source for initiating a conformational change in the repeat-expanded PrP mutants expressed in the brain. The ORD expansion proteins may interact with other preexisting aggregates to foster a change in conformation or create a high local concentration of the mutant protein which then aggregates.

Our model provides a possible explanation for the behavior of PrP repeat expansions and their propensity not only to cause disease but to contribute to the large degree of phenotypic variability observed in patients. Our data suggest that additional repeats in PrP may allow the formation of multiple unique aggregate conformations that could propagate in different tissues at different rates. In addition, if multiple aggregate structures are present in expanded PrP repeat diseases, then this could offer an explanation as to why infectious aggregate structures may also be produced in some cases (7) but not others (8, 14, 45). When we investigated the interaction between SP5 and SP14 by analyzing strains expressing both chimeras, indeed, we did observe a wide range of phenotypes (data not shown). Previous analyses of repeat expansion mutants isolated from human brain tissue demonstrated coaggregation with wild-type PrP, suggesting that phenotypic variability might be explained by the extent of this association (9). We suggest that differential association of mutant and wild-type PrP might be a consequence of different structures of the repeat expansion-containing PrP. Thus, while other genetic modifiers may play a role in disease variability, at least some of the phenotypic variability observed with PrP repeat expansion diseases may be based on intragenic features of the mutant.

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