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REVIEW ARTICLE

Julie R. Ingelfinger, M.D., *Editor*

Kidney Transplantation in Children

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and William E. Harmon, M.D.

SINCE THE FIRST SUCCESSFUL KIDNEY TRANSPLANTATION IN 1954,¹ KIDNEY transplantation has become the best treatment for adult patients with kidney failure. However, early pediatric kidney transplantation was complicated by technical, immunologic, and logistic problems, all leading to worse patient and graft survival among children than had been observed among adults. Over the past 15 years, a number of advances have greatly improved patient and graft survival among children with kidney transplants.^{2,3}

Some aspects of clinical kidney transplantation are similar in children and adults. The immunosuppressive medications and regimens used are similar, creatinine is the major serum biomarker, acute rejection is determined primarily by means of biopsy with the use of the Banff criteria for the classification of rejection (Table S1 in the Supplementary Appendix, available with the full text of this article at NEJM.org), and the rejection mechanisms of the kidney graft are generally similar.⁴⁻⁷ However, many other aspects differ between children and adults — immunologic factors, the primary kidney diseases leading to kidney failure, often with associated urologic issues, and the immunizations that are required before transplantation. Allocation policies regarding kidneys from deceased donors, surgical techniques in small children, and drug metabolism have distinctive aspects in children. The frequency of primary viral infection after transplantation is higher for children than for adults. Furthermore, children are developing so their linear-height growth needs to be optimized and their neurocognitive development fostered. Ultimately, the child with a transplant must be readied for the transition to adult care. This review considers the differences between children and adults undergoing kidney transplantation that necessitate alternative approaches in children and have resulted in innovations and important advances.

THE IMMUNE SYSTEM IN CHILDHOOD

The immune system undergoes profound modification from birth to adulthood, during which period the absolute counts and percentages of circulating T-cell and B-cell subtypes and other immunologic features (Fig. 1)^{8,9} slowly yet continuously evolve, accompanied by an increase in alloreactivity. Thymic output is robust during childhood but diminishes during adulthood.¹⁰ In adults, thymic atrophy and continuous antigen exposure shift the composition of the T-cell pool from naive to memory T cells.¹⁰

As compared with adults, children have an alloimmune response characterized by low expression of the costimulatory ligand CD40L on T cells,¹¹ fewer antigen-specific T-cell precursors,¹² type 2 helper T-cell (Th2)–skewed immunity with higher peripheral levels of Th2 and lower levels of the type 1 helper T-cell (Th1) cytokines,¹³ reduced T-cell effector function,¹⁴ an overall higher percentage of the tolerogenic dendritic-cell subset,¹⁵ and lower titers of anti-HLA antibodies before transplantation¹⁶ (Fig. 1). Younger recipients have better outcomes after transplantation, perhaps suggesting that the more naive immune system provides an advantage (Fig. 2A).

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Figure 1 (facing page). Immunologic Mechanisms, Observed in Pediatric Kidney-Transplant Recipients, That May Be Beneficial for Children.

Important features of the naive pediatric immune system can be found in T cells (e.g., reduced CD40L expression, a type 2 helper T-cell-skewed profile, and reduced effector markers; Panel A), macrophages (e.g., reduced HLA-DR expression and reduced costimulatory molecule expression; Panel B), B cells (e.g., fewer mature B cells and reduced anti-HLA antibody production; Panel C), and dendritic cells (e.g., fewer mature dendritic cells; Panel D). T cells in the recipient may be primed by donor-derived antigen-presenting cells, thus initiating rejection of the kidney allograft. Primed T cells release proinflammatory cytokines and facilitate B-cell activation in the recipient. Dendritic cells in the recipient may also activate T cells by means of indirect allorecognition, leading to chronic allograft rejection. Activated macrophages in the recipient sustain inflammation through the release of proinflammatory cytokines. TCR denotes T-cell receptor, TLR4 toll-like receptor 4, and TNF- α tumor necrosis factor α .

**CONSIDERATIONS BEFORE
TRANSPLANTATION**

CAUSES OF KIDNEY FAILURE

The most common primary causes of kidney failure are congenital or inherited disorders such as renal dysplasia, obstructive uropathies, or reflux nephropathy in young children¹⁹ and acquired glomerular diseases such as focal segmental glomerulosclerosis and lupus nephritis in older children. In contrast, the most common primary renal diseases that lead to end-stage kidney disease in adults are diabetic nephropathy, hypertension, and autosomal dominant polycystic kidney disease, which rarely cause end-stage kidney disease in children.

UROLOGIC ISSUES

Abnormal bladder function may accompany kidney failure in children. In patients with a posterior urethral valve, an open vesicostomy may need to be performed early in life to decompress a dysfunctional bladder. An open vesicostomy may be kept in place for many months after transplantation.²⁰ Children with small bladder capacity may benefit from a bladder augmentation in which segments of ileum, stomach, or appendix are used to create a permanent cutaneous conduit that enables the child to be continent and to have clean, intermittent catheterization.²⁰ Children with obstructive uropathy have a higher rate of urinary tract infection after transplantation than children with other causes of end-stage kidney disease,²¹ possibly necessitating lifelong antimicrobial prophylaxis.

IMMUNIZATIONS

Children require multiple vaccinations during early childhood to protect them from preventable infectious diseases. However, vaccines may not be effective if administered to an immunocom-

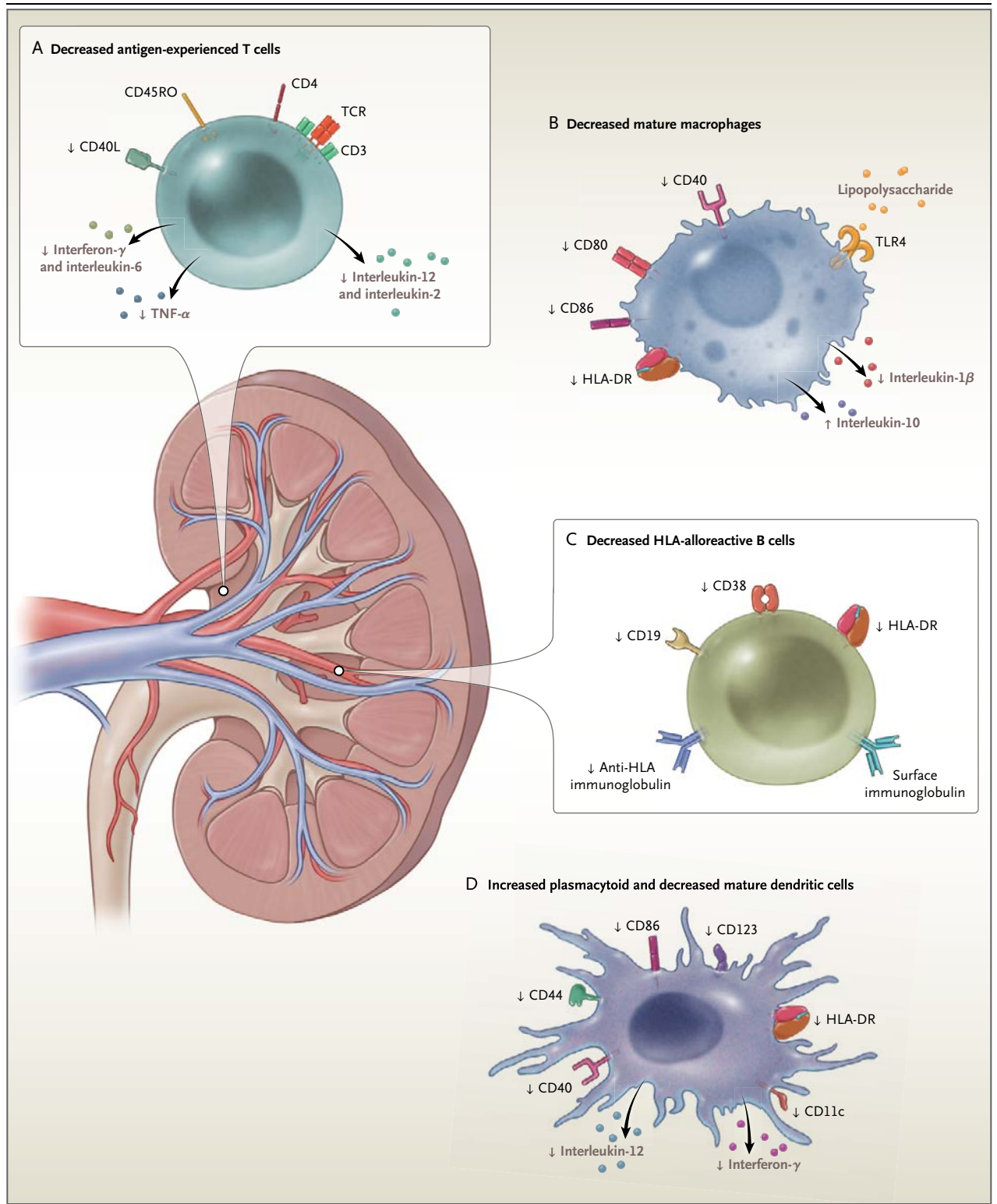
promised patient. Therefore, a vigorous effort to immunize children completely before transplantation is critical. Because children with end-stage kidney disease often have a suboptimal immune response and reduced duration of immunity, higher initial doses, extra doses, and antibody titer monitoring with booster doses of vaccines may be needed. In the period after transplantation, the administration of live vaccines is generally avoided,²² but other immunizations may be given after immunosuppressive medications have reached low maintenance levels, typically at 6 to 12 months after transplantation. Injectable influenza vaccine should be given annually.

PRIORITIZED ALLOCATION OF KIDNEYS TO CHILDREN

The allocation of kidneys from deceased donors involves a complex algorithm that includes the degree of anti-HLA sensitization, the need for multiple donor organs, the blood-group match, the relative HLA match, and the waiting time accrued by the candidate recipient. Children make up a small fraction of persons awaiting kidney transplantation, and they have been afforded exceptional societal benefits in many countries. Thus, allocation policies regarding organ transplants have preferentially allocated higher-quality kidneys from deceased donors to children in relatively prompt fashion,²³ with resultant mean waiting times as short as 3 months in some regions.²⁴ However, such policies have led to a decline in the donation of kidneys from living donors and to a greater proportion of poorly HLA-matched kidney transplants from deceased donors in children.²⁵

**SURGICAL ISSUES AT
TRANSPLANTATION**

Unlike heart and liver allografts, the kidney allograft is placed in a different location from the failed native organ, and the native organ is often



left in place. Thus, size and age matching is generally not required in kidney transplantation. In fact, matching very young donors to very young

recipients was associated previously with a very high rate of graft loss, often due to thrombosis.²⁶ On the basis of those adverse results, pediatric

Figure 2 (facing page). Graft and Patient Survival and Rates of Rejection and Post-Transplantation Lymphoproliferative Disorder (PTLD).

The rates of graft survival at 10 years after receipt of a transplant from a living donor are lowest among recipients older than 65 years of age and adolescents and are highest among the youngest recipients (Panel A). The data are from the Scientific Registry of Transplant Recipients 2012 annual report.¹⁷ Graft survival has improved for transplants from living donors and deceased donors, but a gap between the two sources remains (Panel B). The data, which are based on 11,603 renal transplantations in 10,632 children, are from the North American Pediatric Renal Trials and Collaborative Studies (NAPRTCS).¹⁸ Rates of acute rejection within the first 6 months have decreased in each more-recent cohort, but the slope of the rate of acute rejection at later time points has remained the same (Panel C). The graph for transplants from deceased donors (data not shown) is similar to the graph shown for transplants from living donors. The data are from NAPRTCS.¹⁸ As compared with survival among adult recipients of kidney transplants, patient survival among children is very high, regardless of whether the transplant was from a living or deceased donor (Panel D). The data are from NAPRTCS.¹⁸ Rates of PTLD rose dramatically in the mid-1990s and early 2000s, primarily driven by a rise in early PTLD that may have been related to the effects of more-potent immunosuppressive medications (Panel E) (Martz K, EMMES: personal communication).

programs now transplant adult kidneys into small children once the recipient has reached a sufficient size, typically 6.5 to 10.0 kg of body weight. An infant's peritoneal cavity has enough space to accommodate an adult kidney without compressing the allograft.^{27,28} However, the youngest pediatric recipients have an allograft-size mismatch that leads to a high glomerular filtration rate and makes interpretation of serum creatinine results more difficult, since acute rejection may initially occur without an elevation of the serum creatinine level. Kidneys from deceased donors who were very small children are no longer allocated to small children but are, in fact, now transplanted en bloc (both kidneys together, attached to a single segment of the aorta and vena cava) into adults with excellent results.²⁹

The surgical procedure for a kidney transplantation in a child with a body weight of more than 30 kg is identical to that in an adult. However, in a child with a body weight of less than 10 kg, a midline longitudinal abdominal incision is needed. Space between the peritoneum and subcutaneous fascia is limited, so the kidney is placed intraperitoneally, with a small risk that it will migrate to another part of the abdominal cavity. Blood vessels from the donor are connected to the recipient's aorta and inferior vena cava. In children with a body weight of 10 to 30 kg, surgeons individualize the incision and allograft sites and blood-vessel anastomoses on the basis of the child's anatomy. In small children, strict attention must be paid to maintenance of the intravascular volume during the operation and in the early postoperative period.^{30,31} Some children may require native nephrectomies, either to prevent blood-flow steal by the native kidneys or to eliminate excess urine volume or

protein losses. Additional surgical details are described elsewhere.³⁰

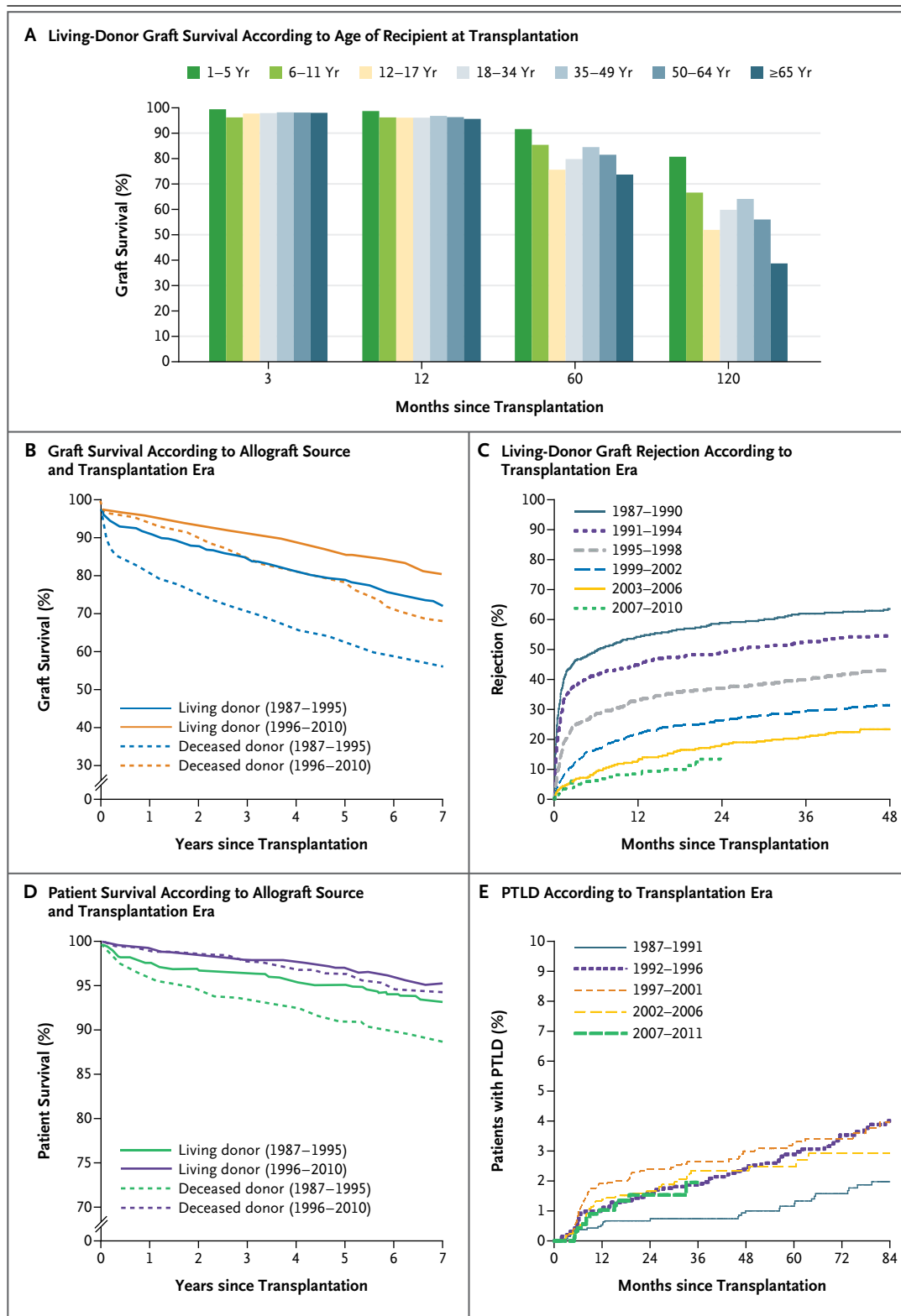
Temporary ureteral stenting is commonly used in adult kidney-transplant recipients,³² but whether stenting is truly helpful in children is not known. Recommendations to stent became common before the association of ureteral stenting with BK virus nephropathy was known.^{33,34} Some pediatric kidney-transplant recipients require specialized ureteral reimplantation, bladder augmentation, or urinary diversion procedures.³⁵

CONSIDERATIONS AFTER TRANSPLANTATION

GRAFT SURVIVAL

Kidney-allograft survival has improved tremendously over time in successive cohorts of pediatric recipients,¹⁸ regardless of whether the transplant was from a living or deceased donor (Fig. 2B). Such progress can be attributed to multiple factors — refinements in pretransplantation preparation, enhanced surgical techniques, better choice of donors, more potent immunosuppressive medications, greater understanding of pediatric-specific pharmacokinetics, and use of evidence-based medication protocols. In addition, overall rates of acute rejection among children have declined; the acute-rejection rate at 1 year among recipients of allografts from living donors decreased from 55% in the late 1980s to 10 to 15% in the most recent cohorts (Fig. 2C).¹⁸ Although developing countries have lower rates of transplantation than developed countries, in addition to limited resources for acquiring the newer, more expensive immunosuppressive agents, they have had similar improvements.³⁶

Kidneys transplanted into children 5 years of



age or younger have shown the most dramatic improvement (Fig. 2A). Unfortunately, adolescents now have the worst long-term graft survival

among all pediatric-recipient age groups and represent the highest-risk recipients. Many reasons are postulated for this outcome, of which

poor adherence to medication therapy is believed to be a major factor.^{37,38} The early mortality among pediatric kidney-transplant recipients is very low (Fig. 2D), and death results mostly from infection or cancer,¹⁸ whereas mortality after transplantation is much higher among adults, and deaths are largely due to cardiovascular disease.

VIRAL INFECTIONS

Opportunistic viruses have emerged as great challenges to clinical management after kidney transplantation, probably related to the immunosuppressive regimens used currently, which are more potent than those used in the past. Since the mid-1990s, the incidence of the Epstein-Barr virus (EBV)-driven cancer known as post-transplantation lymphoproliferative disorder (PTLD) has dramatically increased (Fig. 2E), and BK virus has emerged as a new cause of infection. These two viruses typically infect people early in life, when they are immunocompetent, and cause mild disease but leave behind a pool of latent virus in the reticulothelium or urothelium. Since kidneys transplanted in children are usually from adult donors, there is an increased chance that a kidney from a seropositive donor (with latent virus) will be transplanted into a seronegative recipient. Thus, as compared with adults, children are at higher relative risk for severe disease from cytomegalovirus, EBV, or BK virus,³⁹ with higher rates of complications, graft loss, and death.^{40,41}

Transplantation physicians typically reduce immunosuppression as a first response to each infection, with varied results. Ganciclovir is generally effective both as prophylaxis against and as treatment for cytomegalovirus infection, and antiviral prophylaxis has been associated with reduced rates of PTLD.^{42,43} For BK virus infection, no antiviral treatment strategies have been validated, although cidofovir and leflunomide have been used in both adults and children.^{41,44} Many pediatric kidney-transplantation centers perform serial monitoring for viruses with the use of a polymerase-chain-reaction (PCR) assay in the first 12 months after transplantation, in order to detect infections early.⁴⁵

GROWTH CONCERNS

Children are in a state of active growth. Chronic kidney failure can lead to severe growth failure, often with associated loss of self-esteem.⁴⁶ Children with kidney failure were once approximately

2.5 SD below the expected height for their age at the time of transplantation. Improved nutrition before transplantation and aggressive use of recombinant human growth hormone have reduced, although not eliminated, this height deficit.⁴⁶ Renal transplantation generally improves linear growth but does not completely restore it.⁴⁷ The greatest recovery in growth is seen in the youngest children, and the least is seen in adolescents. The use of glucocorticoid withdrawal or avoidance protocols and the administration of growth hormone after transplantation may further improve growth recovery.⁴⁸

TRANSITION OF CARE

Adolescents must eventually graduate to adult care — a transition that can be stressful for both the patient and the caregiver. A gradual transition rather than an abrupt transfer, early preparation of the patient and family, and the use of checklists to document maturing transition skills in the patient have been recommended to ensure a successful transition.⁴⁹

PEDIATRIC IMMUNOSUPPRESSION — LESSONS FROM CLINICAL TRIALS

An important advance in pediatric kidney transplantation over the past two decades has been the emergence of prospective trial groups in the United States, Europe, Australia, and Asia. With relatively low patient volumes (10 to 30 transplantations per year) at most centers, meaningful results can be obtained through multicenter collaboration. Retrospective data registries such as the North American Pediatric Renal Trials and Collaborative Studies, combined adult and pediatric registries such as the United Network for Organ Sharing, the United States Renal Data System, the Collaborative Transplant Study, the Australia and New Zealand Dialysis and Transplant Registry, and the newly formed Cooperative European Pediatric Renal Transplant Initiative Registry in Europe⁵⁰ have provided important, but limited, information (Table S2 in the Supplementary Appendix). The results of prospective, randomized, multicenter clinical trials conducted in the United States or Europe are summarized in Table S3 in the Supplementary Appendix, and the results of nonrandomized, multicenter trials are summarized in Table S4 in the Supplementary Appendix. These trials have shown that high doses of immunosuppressive drugs to compensate for gluco-

corticoid withdrawal can lead to unacceptable rates of PTLD, that glucocorticoid avoidance is not immunologically detrimental,⁵¹ although it does not ameliorate chronic histologic damage,⁵² and that tacrolimus is associated with a significantly lower rate of acute rejection at 6 months than is cyclosporine.⁵³

The children enrolled in these trials had a high rate of adherence to the trial requirements, including protocol-specified graft biopsies. However, pilot trials of immunosuppression in children should be conducted only if there are at least preliminary safety and perhaps efficacy data from studies involving adults, unless the studies apply uniquely to children (e.g., studies of growth strategies). Because only 700 to 800 kidney transplantations are performed in children in the United States annually, cooperation among pediatric research consortia is critical for full enrollment and study completion.

Pharmacokinetic and mechanistic studies have been coupled to many of these prospective studies. Pharmacokinetic studies showed that cyclosporine has a shorter half-life in children than in adults and requires dosing three times daily.⁵⁴ Similarly, sirolimus has a shorter half-life in children than in adults and often requires twice-daily dosing.^{55,56} The area under the curve of dose-normalized mycophenolic acid is higher in children than has been commonly observed in adults.⁵⁶ These differences in metabolism are believed to be due to developmental changes in biliary transporters and metabolic enzymes such as cytochrome P-450 and glucuronosyltransferases.⁵⁶ In glucocorticoid-free protocols as compared with protocols that include glucocorticoids, the use of mycophenolate mofetil is associated with more frequent and severe leukopenia, anemia, and gastrointestinal disturbances.^{57,58}

Some of the most important results from these trials were obtained from mechanistic studies, particularly those that may account for the replenishment of the immune system after lymphocyte depletion — homeostatic proliferation of peripheral memory T cells in adult recipients and thymopoiesis in pediatric recipients.⁵⁹ In children, CD8+ naive cells were shown to be more resistant to alemtuzumab-mediated depletion, and their recovery, albeit greater than that of CD4+ cells, remained low at 24 months,⁶⁰ whereas CD8+ cells in adults were fully recovered at 6 months after transplantation.^{61,62} The prolonged depletion obtained with alemtuzumab (a potent

monoclonal anti-CD52 antibody) in children is in conflict with the general theory that depletion would be more transient in children, owing to more active lymphopoiesis, than in adults.⁶³ Induction therapy with rabbit antithymocyte globulin (ATG), a polyclonal depleting agent, promotes long-term graft survival in pediatric kidney-transplant recipients,⁶⁴ with depletion of naive T cells and central memory T cells and little effect on effector memory T cells.⁶⁵ In addition, regulatory T cells (Tregs) were spared with alemtuzumab and expanded with ATG,⁶⁵ conferring an additional immunologic benefit by helping to accommodate the graft.

B-cell depletion with the lytic chimeric mouse-human anti-CD20 antibody rituximab is used increasingly in pediatric kidney-transplant recipients.⁶⁶ Data have shown full recovery of the B-cell pool 15 months after rituximab treatment in children, whereas recovery began only at 24 months in adults and was never complete.⁶⁷ Reemerging B cells appeared to be naive, immature, and more regulatory in children than in adults.⁶⁶ By 3 to 5 years after transplantation, new antibodies directed against HLA antigens developed in 25% of children,⁶⁸ as compared with only 10% of adults.⁶⁹

Identification of relatively noninvasive biomarkers of acute rejection that might replace transplant biopsy is important. PCR-based panels of candidate markers such as interferon-inducible protein 10 and forkhead box P3 from urine have been tested.^{70,71} Recently, several of these markers have been validated for acute rejection in adult kidney-transplant recipients, although BK virus infection also elevates these markers.⁷⁰ A PCR-based five-gene panel was tested in pediatric kidney-transplant recipients and validated as highly sensitive for the prediction of acute rejection.⁷² The panels in these two studies^{70,72} have no markers in common with each other or with the AlloMap panel⁷³ used to predict acute rejection in cardiac-transplant recipients, raising the question of whether the mechanisms of acute rejection are fundamentally different in different organ systems or patient populations or are influenced by differences in immunosuppression protocols.

FUTURE THERAPEUTIC DEVELOPMENTS

Inducing tolerance to a kidney allograft (tolerogenic strategies) holds the promise of immunosuppressive-free management. Different tolerogenic strategies have been explored primarily in the

preclinical setting but could have important benefits for children. For example, a phase 3 study of belatacept-based immunosuppression regimens versus cyclosporine in renal-transplant recipients (Belatacept Evaluation of Nephroprotection and Efficacy as First-line Immunosuppression Trial [BENEFIT]) showed a better average glomerular filtration rate at month 60 in the belatacept group than in the cyclosporine group.⁷⁴ Belatacept, a selective T-cell (lymphocyte) costimulation blocker, is being studied cautiously in children owing to concern about the risk of PTLD. Abatacept, another selective costimulatory agent, may have positive effects beyond the potential for tolerogenicity, because it has decreased proteinuria in kidney-transplant recipients with recurrent focal segmental glomerulosclerosis.⁷⁵

Treatment with stem cells and Tregs, which is being studied in adult kidney-transplant recipients, may become feasible for pediatric recipients as well.⁷⁶ Mesenchymal stem cells are poorly immunogenic bone marrow-derived stem cells that inhibit T-cell proliferation and induce Tregs.⁷⁷ A recent study evaluated mesenchymal stem cells as an alternative to antibody induction therapy

in adult kidney-transplant recipients.⁷⁸ However, mesenchymal stem cells have triggered concerns, given their oncogenic potential.⁷⁹

SUMMARY

Transplantation in children with kidney failure once presented many technical, immunologic, and logistic problems that led to worse patient and allograft survival, as compared with adults. Advances in all these areas and the development of pediatric-trial groups have resulted in dramatic improvements, such that young children now have the best long-term graft survival among all age groups, including adults.

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Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

REFERENCES

- Murray JE. The first successful organ transplants in man. *J Am Coll Surg* 2005; 200:5-9.
- Horslen S, Barr ML, Christensen LL, Ettenger R, Magee JC. Pediatric transplantation in the United States, 1996-2005. *Am J Transplant* 2007;7:1339-58.
- Harmon WE. Pediatric kidney transplantation. In: Avner ED, Harmon WE, Niaudet P, Yoshikawa N, eds. *Pediatric nephrology*. 6th ed. Berlin: Springer-Verlag, 2009:1867-902.
- Solez K, Colvin RB, Racusen LC, et al. Banff 07 classification of renal allograft pathology: updates and future directions. *Am J Transplant* 2008;8:753-60.
- Harmon WE. Pediatric renal transplantation. In: Himmelfarb J, Sayegh MH, eds. *Chronic kidney disease, dialysis and transplantation*. Philadelphia: Elsevier, 2010:591-608.
- Nankivell BJ, Alexander SI. Rejection of the kidney allograft. *N Engl J Med* 2010; 363:1451-62.
- Haas M, Sis B, Racusen LC, et al. Banff 2013 meeting report: inclusion of c4d-negative antibody-mediated rejection and antibody-associated arterial lesions. *Am J Transplant* 2014;14:272-83.
- Shearer WT, Rosenblatt HM, Gelman RS, et al. Lymphocyte subsets in healthy children from birth through 18 years of age: the Pediatric AIDS Clinical Trials Group P1009 study. *J Allergy Clin Immunol* 2003;112:973-80.
- Härtel C, Adam N, Strunk T, Temming P, Müller-Steinhardt M, Schultz C. Cytokine responses correlate differentially with age in infancy and early childhood. *Clin Exp Immunol* 2005;142:446-53.
- McFarland RD, Douek DC, Koup RA, Picker LJ. Identification of a human recent thymic emigrant phenotype. *Proc Natl Acad Sci U S A* 2000;97:4215-20.
- Brugnoni D, Airo P, Graf D, et al. Ontogeny of CD40L [corrected] expression by activated peripheral blood lymphocytes in humans. *Immunol Lett* 1996;49:27-30. [Erratum, *Immunol Lett* 1996;52:61.]
- Hassan J, Reen DJ. Reduced primary antigen-specific T-cell precursor frequencies in neonates is associated with deficient interleukin-2 production. *Immunology* 1996;87:604-8.
- Hanna-Wakim R, Yasukawa LL, Sung P, et al. Age-related increase in the frequency of CD4(+) T cells that produce interferon-gamma in response to staphylococcal enterotoxin B during childhood. *J Infect Dis* 2009;200:1921-7.
- Chiba Y, Higashidate Y, Suga K, Honjo K, Tsutsumi H, Ogra PL. Development of cell-mediated cytotoxic immunity to respiratory syncytial virus in human infants following naturally acquired infection. *J Med Virol* 1989;28:133-9.
- Jyonouchi H, Cui C, Geng L, Yin Z, Fitzgerald-Bocarsly P. Age-dependent changes in peripheral blood dendritic cell subsets in normal children and children with specific polysaccharide antibody deficiency (SPAD). *Eur J Pediatr* 2010;169:1233-9.
- Gupta A, Iveson V, Varagunam M, Bodger S, Sinnott P, Thuraisingham RC. Pretransplant donor-specific antibodies in cytotoxic negative crossmatch kidney transplants: are they relevant? *Transplantation* 2008;85:1200-4.
- Scientific Registry of Transplant Recipients. 2012 Annual data report (http://srtr.transplant.hrsa.gov/annual_reports/2012/Default.aspx).
- North American Pediatric Renal Trials and Collaborative Studies. 2010 Annual transplant report (https://web.emmes.com/study/ped/annlrept/2010_Report.pdf).
- McEnery PT, Stablein DM, Arbus G, Tejani A. Renal transplantation in children — a report of the North American Pediatric Renal Transplant Cooperative Study. *N Engl J Med* 1992;326:1727-32.
- Mitchell ME, Balcom AH. Bladder dysfunction in children. In: Avner ED, Harmon WE, Niaudet P, Yoshikawa N, eds. *Pediatric nephrology*. 6th ed. Berlin: Springer-Verlag, 2009:1379-404.
- Dharnidharka VR, Agodoa LY, Abbott KC. Effects of urinary tract infection on

- outcomes after renal transplantation in children. *Clin J Am Soc Nephrol* 2007;2:100-6.
22. Neu AM, Fivush BA. Recommended immunization practices for pediatric renal transplant recipients. *Pediatr Transplant* 1998;2:263-9.
23. Organ Procurement and Transplantation Network. Allocation of deceased kidneys. Washington, DC: Department of Health and Human Services, 2010 (http://optn.transplant.hrsa.gov/PoliciesandBylaws2/policies/pdfs/policy_7.pdf).
24. Reese PP, Hwang H, Potluri V, Abt PL, Shults J, Amaral S. Geographic determinants of access to pediatric deceased donor kidney transplantation. *J Am Soc Nephrol* 2014;25:827-35.
25. Moudgil A, Dharnidharka VR, Lamb KE, Meier-Kriesche HU. Best allograft survival from share-35 kidney donors occurs in middle-aged adults and young children — an analysis of OPTN data. *Transplantation* 2013;95:319-25.
26. Harmon WE, Alexander SR, Tejani A, Stablein D. The effect of donor age on graft survival in pediatric cadaver renal transplant recipients — a report of the North American Pediatric Renal Transplant Cooperative Study. *Transplantation* 1992;54:232-7.
27. Li L, Chang A, Naesens M, et al. Steroid-free immunosuppression since 1999: 129 pediatric renal transplants with sustained graft and patient benefits. *Am J Transplant* 2009;9:1362-72.
28. Sarwal MM, Cecka JM, Millan MT, Salvatierra O Jr. Adult-size kidneys without acute tubular necrosis provide exceedingly superior long-term graft outcomes for infants and small children: a single center and UNOS analysis. *Transplantation* 2000;70:1728-36.
29. Dharnidharka VR, Stevens G, Howard RJ. En-bloc kidney transplantation in the United States: an analysis of United Network of Organ Sharing (UNOS) data from 1987 to 2003. *Am J Transplant* 2005;5:1513-7.
30. Salvatierra O Jr, Millan M, Concepcion W. Pediatric renal transplantation with considerations for successful outcomes. *Semin Pediatr Surg* 2006;15:208-17.
31. Shapiro R, Sarwal MM. Pediatric kidney transplantation. *Pediatr Clin North Am* 2010;57:393-400.
32. Wilson CH, Bhatti AA, Rix DA, Manas DM. Routine intraoperative stenting for renal transplant recipients. *Transplantation* 2005;80:877-82.
33. Thomas A, Dropic LK, Rahman MH, Geetha D. Ureteral stents: a novel risk factor for polyomavirus nephropathy. *Transplantation* 2007;84:433-6.
34. Brennan DC, Agha I, Bohl DL, et al. Incidence of BK with tacrolimus versus cyclosporine and impact of preemptive immunosuppression reduction. *Am J Transplant* 2005;5:582-94. [Erratum, *Am J Transplant* 2005;5:839.]
35. Riley P, Marks SD, Desai DY, Mushtaq I, Koffman G, Mamode N. Challenges facing renal transplantation in pediatric patients with lower urinary tract dysfunction. *Transplantation* 2010;89:1299-307.
36. Rizvi SA, Sultan S, Zafar MN, et al. Pediatric kidney transplantation in the developing world: challenges and solutions. *Am J Transplant* 2013;13:2441-9.
37. Dobbels F, Ruppert T, De Geest S, Decorte A, Van Damme-Lombaerts R, Fine RN. Adherence to the immunosuppressive regimen in pediatric kidney transplant recipients: a systematic review. *Pediatr Transplant* 2010;14:603-13.
38. Hsu DT. Biological and psychological differences in the child and adolescent transplant recipient. *Pediatr Transplant* 2005;9:416-21.
39. Dharnidharka VR, Cherikh WS, Abbott KC. An OPTN analysis of national registry data on treatment of BK virus allograft nephropathy in the United States. *Transplantation* 2009;87:1019-26.
40. Dharnidharka VR, Martz KL, Stablein DM, Benfield MR. Improved survival with recent post-transplant lymphoproliferative disorder (PTLD) in children with kidney transplants. *Am J Transplant* 2011;11:751-8.
41. Dharnidharka VR, Abdunour HA, Araya CE. The BK virus in renal transplant recipients — review of pathogenesis, diagnosis, and treatment. *Pediatr Nephrol* 2011;26:1763-74.
42. Funch DP, Walker AM, Schneider G, Ziyadeh NJ, Pescovitz MD. Ganciclovir and acyclovir reduce the risk of post-transplant lymphoproliferative disorder in renal transplant recipients. *Am J Transplant* 2005;5:2894-900.
43. Marks WH, Ilsley JN, Dharnidharka VR. Posttransplantation lymphoproliferative disorder in kidney and heart transplant recipients receiving thymoglobulin: a systematic review. *Transplant Proc* 2011;43:1395-404.
44. Dharnidharka VR, Araya CE, Wadsworth CS, McKinney MC, Howard RJ. Assessing the value of ureteral stent placement in pediatric kidney transplant recipients. *Transplantation* 2008;85:986-91.
45. Al Khasawneh E, Araya CE, Dharnidharka VR. Missed viral surveillance testing visits associate with full blown viral diseases in children receiving kidney transplants. *Pediatr Transplant* 2013;17:129-32.
46. Fine RN. Etiology and treatment of growth retardation in children with chronic kidney disease and end-stage renal disease: a historical perspective. *Pediatr Nephrol* 2010;25:725-32.
47. Pape L, Ehrlich JH, Zivicnjak M, Offner G. Growth in children after kidney transplantation with living related donor graft or cadaveric graft. *Lancet* 2005;366:151-3.
48. Fine RN, Stablein D, Cohen AH, Tejani A, Kohaut E. Recombinant human growth hormone post-renal transplantation in children: a randomized controlled study of the NAPRTCS. *Kidney Int* 2002;62:688-96.
49. Bell LE, Bartosh SM, Davis CL, et al. Adolescent transition to adult care in solid organ transplantation: a consensus conference report. *Am J Transplant* 2008;8:2230-42.
50. Plotnicki L, Kohl CD, Höcker B, et al. The CERTAIN Registry: a novel, Web-based registry and research platform for pediatric renal transplantation in Europe. *Transplant Proc* 2013;45:1414-7.
51. Sarwal MM, Ettenger RB, Dharnidharka V, et al. Complete steroid avoidance is effective and safe in children with renal transplants: a multicenter randomized trial with three-year follow-up. *Am J Transplant* 2012;12:2719-29.
52. Naesens M, Salvatierra O, Benfield M, et al. Subclinical inflammation and chronic renal allograft injury in a randomized trial on steroid avoidance in pediatric kidney transplantation. *Am J Transplant* 2012;12:2730-43.
53. Trompeter R, Filler G, Webb NJ, et al. Randomized trial of tacrolimus versus cyclosporin microemulsion in renal transplantation. *Pediatr Nephrol* 2002;17:141-9.
54. Cooney GF, Habucky K, Hoppu K. Cyclosporin pharmacokinetics in paediatric transplant recipients. *Clin Pharmacokinet* 1997;32:481-95.
55. Schachter AD, Benfield MR, Wyatt RJ, et al. Sirolimus pharmacokinetics in pediatric renal transplant recipients receiving calcineurin inhibitor co-therapy. *Pediatr Transplant* 2006;10:914-9.
56. Filler G, Bendrick-Pearl J, Christians U. Pharmacokinetics of mycophenolate mofetil and sirolimus in children. *Ther Drug Monit* 2008;30:138-42.
57. Höcker B, Weber LT, Bunchman T, Rashford M, Tönshoff B. Mycophenolate mofetil suspension in pediatric renal transplantation: three-year data from the Tricontinental Trial. *Pediatr Transplant* 2005;9:504-11.
58. Sarwal MM, Yorgin PD, Alexander S, et al. Promising early outcomes with a novel, complete steroid avoidance immunosuppression protocol in pediatric renal transplantation. *Transplantation* 2001;72:13-21.
59. Mackall CL, Gress RE. Thymic aging and T-cell regeneration. *Immunol Rev* 1997;160:91-102.
60. De Serres SA, Mfarrej BG, Magee CN, et al. Immune profile of pediatric renal transplant recipients following alemtuzumab induction. *J Am Soc Nephrol* 2012;23:174-82.
61. Trzonkowski P, Zilveti M, Chapman S, et al. Homeostatic repopulation by CD28-CD8+ T cells in alemtuzumab-depleted kidney transplant recipients treated with reduced immunosuppression. *Am J Transplant* 2008;8:338-47.
62. Knechtle SJ, Pascual J, Bloom DD, et al. Early and limited use of tacrolimus to avoid rejection in an alemtuzumab and

- sirolimus regimen for kidney transplantation: clinical results and immune monitoring. *Am J Transplant* 2009;9:1087-98.
63. Knechtle SJ. Present experience with campath-1H in organ transplantation and its potential use in pediatric recipients. *Pediatr Transplant* 2004;8:106-12.
64. Mota C, Martins L, Costa T, et al. Nineteen years of experience utilizing anti-T-lymphocyte globulin induction in pediatric kidney transplantation. *Ann Transplant* 2010;15:84-91.
65. Gurkan S, Luan Y, Dhillon N, et al. Immune reconstitution following rabbit antithymocyte globulin. *Am J Transplant* 2010;10:2132-41. [Erratum, *Am J Transplant* 2010;10:2728.]
66. Zarkhin V, Lovelace PA, Li L, Hsieh SC, Sarwal MM. Phenotypic evaluation of B-cell subsets after rituximab for treatment of acute renal allograft rejection in pediatric recipients. *Transplantation* 2011;91:1010-8.
67. Genberg H, Hansson A, Wernerson A, Wennberg L, Tydén G. Pharmacodynamics of rituximab in kidney allotransplantation. *Am J Transplant* 2006;6:2418-28.
68. Chaudhuri A, Ozawa M, Everly MJ, et al. The clinical impact of humoral immunity in pediatric renal transplantation. *J Am Soc Nephrol* 2013;24:655-64.
69. Harmon W, Guleria I, Ikle D, et al. Children develop de novo anti-HLA antibodies (Abs) following kidney transplantation at a higher incidence than adults: an analysis of the NIH CTOT/CCTPT-02 Study. *Am J Transplant* 2013;13:Suppl:36-7. abstract.
70. Suthanthiran M, Schwartz JE, Ding R, et al. Urinary-cell mRNA profile and acute cellular rejection in kidney allografts. *N Engl J Med* 2013;369:20-31.
71. Muthukumar T, Dadhania D, Ding R, et al. Messenger RNA for FOXP3 in the urine of renal-allograft recipients. *N Engl J Med* 2005;353:2342-51.
72. Li L, Khatri P, Sigdel TK, et al. A peripheral blood diagnostic test for acute rejection in renal transplantation. *Am J Transplant* 2012;12:2710-8.
73. Pham MX, Teuteberg JJ, Kfoury AG, et al. Gene-expression profiling for rejection surveillance after cardiac transplantation. *N Engl J Med* 2010;362:1890-900.
74. Rostaing L, Vincenti F, Grinyó J, et al. Long-term belatacept exposure maintains efficacy and safety at 5 years: results from the long-term extension of the BENEFIT study. *Am J Transplant* 2013;13:2875-83.
75. Yu CC, Fornoni A, Weins A, et al. Abatacept in B7-1-positive proteinuric kidney disease. *N Engl J Med* 2013;369:2416-23.
76. Fiorina P, Jurewicz M, Vergani A, et al. Targeting the CXCR4-CXCL12 axis mobilizes autologous hematopoietic stem cells and prolongs islet allograft survival via programmed death ligand 1. *J Immunol* 2011;186:121-31.
77. Fiorina P, Jurewicz M, Augello A, et al. Immunomodulatory function of bone marrow-derived mesenchymal stem cells in experimental autoimmune type 1 diabetes. *J Immunol* 2009;183:993-1004.
78. Tan J, Wu W, Xu X, et al. Induction therapy with autologous mesenchymal stem cells in living-related kidney transplants: a randomized controlled trial. *JAMA* 2012;307:1169-77.
79. Fiorina P, Voltarelli J, Zavazava N. Immunological applications of stem cells in type 1 diabetes. *Endocr Rev* 2011;32:725-54.

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