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Analysis of Interleukin-8 Gene Variants Reveals Their Relative Importance as Genetic Susceptibility Factors for Chronic Periodontitis in the Han Population

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Abstract

Interleukin (IL)-8, an important chemokine that regulates the inflammatory response, plays an important role in periodontitis. Previous studies indicate that certain IL-8 gene polymorphisms are associated with periodontitis susceptibility in some populations. However, the literature is somewhat contradictory, and not all IL-8 polymorphisms have been examined, particularly in Han Chinese individuals. The aim of this study was to investigate the association of every IL-8 SNP with chronic periodontitis in Han Chinese individuals. We analyzed 23 SNPs with minor allele frequency (MAF) ≥ 0.01 , which were selected from 219 SNPs in the NCBI dbSNP and preliminary HapMap data analyses from a cohort of 400 cases and 750 controls from genetically independent Han Chinese individuals. Single SNP, haplotype and gender-specific associations were performed. We found that rs4073 and rs2227307 were significantly associated with chronic periodontitis. Further haplotype analysis indicated that a haplotype block (rs4073-rs2227307-rs2227306) that spans the promoter and exon1 of IL-8 was highly associated with chronic periodontitis. Additionally, the ATC haplotype in this block was increased 1.5-fold in these cases. However, when analyzing the samples by gender, no significant gender-specific associations in IL-8 were observed, similar to the results of haplotype association analyses in female and male subgroups. Our results provide further evidence that IL-8 is associated with chronic periodontitis in Han Chinese individuals. Furthermore, our results confirm previous reports suggesting the intriguing possibilities that IL-8 plays a role in the pathogenesis of chronic periodontitis and that this gene may be involved in the etiology of this condition.

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Introduction

Oral Gram-negative bacteria trigger periodontitis, which produces an inflammatory response in a susceptible host [1]. Individual variations in the host's immune response, which are influenced by environmental and genetic characteristics, account for the predisposition, initiation and progression of periodontitis [2]. Environmental factors, such as hormones, diabetes and drugs, modify preexisting periodontal conditions [3]. Among these factors, tobacco use is one of the main risk factors for periodontitis, because tobacco affects the oral environment, gingival vasculature, inflammatory and immunological responses, as well as the healing potential of periodontal connective tissues [4–6]. Furthermore, increasing evidence suggests that genetic factors are important risk factors for predicting the susceptibility to chronic periodontitis [7].

Several studies have investigated the role of genes and their variants (polymorphisms) in host responses to chronic periodontitis and the progression of this disease [8–11]. In some situations, genetic polymorphisms may change protein's function or expression, altering innate and adaptive immunity, which might affect disease outcomes [12]. Genetic polymorphisms may also be protective against this disease [13]. Furthermore, the immune response of patients affected by periodontitis has been widely investigated, focusing on cytokine production and its association with chronic periodontitis [14–16]. Genetic susceptibility to chronic periodontitis has also been studied, focusing on immune system genes, such as interleukin-8 (IL-8). The IL-8 gene, which is located on chromosome 4q12-q13, encodes the IL-8 protein. This protein is the most potent chemokine studied to date, and it is responsible for inducing chemotaxis, which is the directed migration of cells to a site of inflammation [17]. This chemokine

Table 1. Demographic characteristics and clinical parameters of chronic periodontitis and controls.

	Chronic periodontitis (n = 400)	Controls (n = 750)
Gender (male/female)	200/200	375/375
Range of age (years)	28–62	27–60
Mean age (years)	50.46±9.14	50.32±8.27
PD (mm)	5.52±0.26	NA
CAL (mm)	5.58±0.45	NA
BOP (%)	48.06±13.43	NA

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is important for regulation of the inflammatory response and for its ability to recruit and activate acute inflammatory cells. IL-8 is also mediates the activation and migration of neutrophils, which are the first line of defense against periodontopathic bacteria, which migrate from the peripheral blood into the tissues [18]. Moreover, IL-8 is produced early in the inflammatory response, and its presence persists for a long period of time [19].

While association studies provide a promising approach for studying the genetics of complex diseases, such as schizophrenia [20–23], identifying individual candidate genes/variants for disease risk is also important. Previous studies indicate the association between IL-8 SNPs and periodontitis compared with healthy controls. Some studies suggest that SNPs within IL-8 are associated with susceptibility to periodontitis [24–28], whereas other studies failed to demonstrate associations between periodontitis and IL-8 polymorphisms [29,30]. The association between IL-8 and chronic periodontitis has not been systematically investigated in the Han Chinese population. Despite evidence of a significant association within some populations, the contribution of IL-8 to periodontitis and its potential mechanisms of action in periodontitis remain to be elucidated. Therefore, an exploration of possible association between IL-8 polymorphisms and periodontitis is necessary among genetically independent populations. To confirm the association of IL-8 with chronic periodontitis, we performed an association study for IL-8 with chronic periodontitis in Han Chinese individuals. Additionally, this study provides further information regarding the use of IL-8 polymorphisms as markers of susceptibility to periodontal disease.

Materials and Methods

Patients and controls

This study was designed as a case-controlled study. The study group was composed of 1150 individuals, ranging from 27–62 years of age. It took almost one year (June 2012 to April 2013) to complete sample collection. All of the subjects were unrelated Han Chinese individuals randomly selected from the Shaanxi Province, with no migration history within the previous 3 generations. Additionally, all participants were of a similar socio-economic level, which is important because there is a strong association between low socio-economic status and a higher risk of periodontal disease [31]. All enrolled individuals answered a questionnaire to obtain information regarding dental history, family history of periodontal disease, cigarette smoking habits and general health. All of the subjects were required to have at least 10 teeth, be in good general health and be free of oral soft tissue abnormalities or severe dental caries, except for the presence of chronic periodontitis. Patients who reported the following characteristics were excluded from the study: use of orthodontic appliances, chronic anti-inflammatory drugs or immunosuppressive chemotherapy, antibiotics within the previous 3 months, chronic inflammatory diseases, a history of diabetes mellitus, hepatitis, HIV infection, nephritis, bleeding or autoimmune disorders, diseases with severe commitment of the immune function, current pregnancy or breastfeeding. Other authors have previously used these exclusion criteria [32–35]. Because tobacco is associated with increased clinical attachment loss (CAL) and supports alveolar bone loss, it represents an important risk factor for the initiation and progression of periodontal disease [3,36–38]. The individuals enrolled in this study were all nonsmokers (never smoked before).

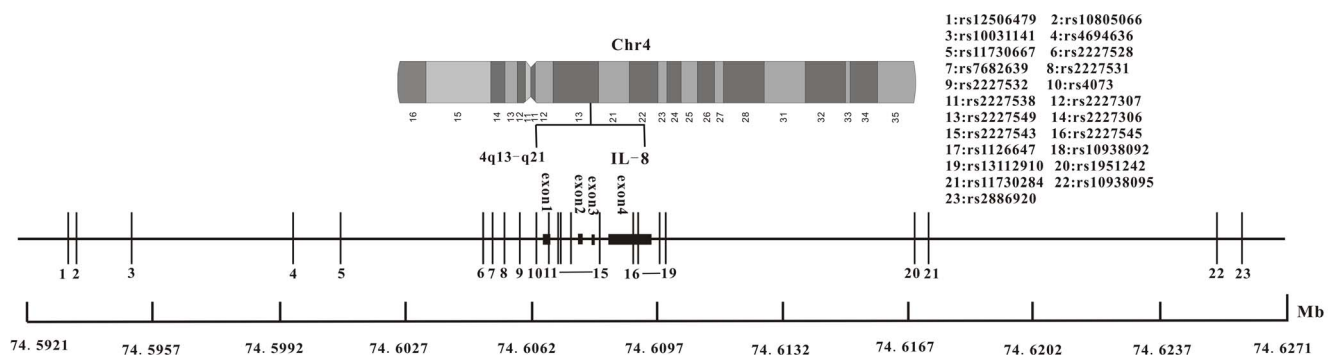


Figure 1. Distribution of the 23 SNPs across the IL-8 gene selected for the association analysis and their relationship with gene exons.

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Individuals were categorized into the control and chronic periodontitis groups.

The healthy control group did not have signs of any periodontal disease at the time of sample collection and did not have a history of previous periodontal disease as determined by a lack of sites with probing depth (PD) >3 mm and the absence of gingival recession, CAL, and bleeding on probing (BOP). The 750 blood samples from periodontally healthy Han Chinese subjects were obtained randomly, which represented the controlled population (375 males and 375 females, aged 27 to 60 years with a mean age of 50.32±8.27 years; Table 1).

Patients with chronic periodontitis often presented an amount of destruction consistent with the amount of microbial deposits, presence of subgingival calculus, probable association with local predisposing factors and a slow to moderate rate of progression. All of the chronic periodontitis subjects were previously diagnosed with moderate or severe chronic periodontal disease. Diagnosis of chronic periodontitis (CP) was established clinically and by X-ray verification, according to the criteria of the American Academy of Periodontal Disease (AAP, 1999) [39], presence of chronic gingivorrhagia, bleeding on probing, clinical attachment loss, and horizontal or vertical loss of alveolar bone. In all patients, the degree of clinical attachment loss was defined using confirmed periodontal probe. Patients with probing depths greater than 5 mm, CAL greater than 4 mm, and some degree of gingival recession and tooth mobility were chosen. This clinical form is most prevalent in adults, but its occurrence may be present in younger individuals [40]. This patient group was composed of 400 subjects (200 males and 200 females, aged 28 to 62 years with a mean age of 50.46±9.14 years; Table 1) that were recruited from the inpatient and outpatient clinical services at the First Affiliated Hospital of Xi'an Jiaotong University, the second Affiliated Hospital of Xi'an Jiaotong University and the Stomatology Hospital of Xi'an Jiaotong University.

This study was approved by the Xi'an Jiaotong University Ethics Committee. All participants completed written informed consent forms. Data related to the participants are described in Table 1.

SNP selection and genotyping

IL-8 polymorphisms were identified using the National Center for Biotechnology Information single nucleotide polymorphism database, and 209 SNPs were identified. For the first screen of the most common SNPs in Han Chinese chronic periodontitis patients, a MAF≥0.01 was used as the cut-off. Based on these criteria, we selected 14 SNPs in IL-8 (rs2227528, rs7682639, rs2227531, rs2227532, rs4073, rs2227538, rs2227307, rs2227549, rs2227306, rs2227543, rs2227545, rs1126647, rs10938092 and rs13112910). Next, we then searched for all SNPs with minor allele frequencies (MAF)≥0.01 between 20 kb upstream and 20 kb downstream of IL-8 in the HapMap HCB database using Haploview [41], which identified 9 SNPs (rs12506479, rs10805066, rs10031141, rs46946336, rs11730667, rs1951242, rs11730284, rs10938095 and rs2886920). Therefore, we selected 23 SNPs in the 45 kb region containing IL-8 (Fig. 1).

Patients and controls were mixed on the same plates using a double-blind procedure. Plasma samples were stored at -20°C. Genomic DNA was isolated from peripheral blood leukocytes according to the manufacturer's protocol (Genomic DNA kit, Axygen Scientific Inc., California, USA), and DNA samples were stored at -20°C for SNP analysis. SNP genotyping was performed using the Sequenom MassARRAY platform with iPLEX GOLD chemistry (Sequenom, San Diego, CA, USA), according to the manufacturer's protocol. Polymerase chain reaction (PCR) prim-

Table 2. Allele and genotype frequency of single SNP association analysis.

SNP Makers	Allele Freq. (%)		Genotype Freq. (%)	p-value ¹	H-W E P value	OR ² 95%CI	p-value	
	A	T					Dominant	Recessive
rs4073								
Case	318(39.8)	482(60.2)	AA 71(17.7)	0.028251	0.121	1.207	0.702	0.547
Control	668(44.5)	833(55.5)	AA 140(18.7)		0.222	(1.014-1.437)		
rs2227307								
Case	282(35.2)	518(64.8)	GG 44(11.1)	0.003455	0.258	1.312	0.007	0.035
Control	623(41.5)	877(58.5)	GG 127(16.9)		0.716	(1.098-1.567)		
rs2227306								
Case	574(71.8)	226(28.2)	CC 209(52.3)	0.063353	0.460	1.195	0.538	0.977
Control	1020(68.0)	480(32.0)	CC 344(45.9)		0.669	(0.990-1.443)		

OR: odds ratio; CI: confidence interval.
¹p values of the normal chi-square statistics from Monte Carlo stimulation using CLUMP (T1), and significant p values are in italic bold.
²OR refers to risk allele odds ratio in cases and controls.
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Table 3. Results of logistic regression analysis for susceptibility to chronic periodontitis.

Characteristics of subjects		Case	Control	OR	95%CI	p-value
Age				1.292	1.168–1.435	0.002
Gender	Male	200(34.78%)	375(65.22%)	Reference		
	Female	200(34.78%)	375(65.22%)	1.003	0.832–1.375	0.974
rs4073 TT carriage	Non-carriers	248(32.00%)	527(68.00%)	Reference		
	Carriers	152(40.53%)	223(59.47%)	1.451	1.196–1.628	0.017
rs2227307 T allele carriage	Non-carriers	44(25.73%)	127(74.27%)	Reference		
	Carriers	356(36.36%)	623(63.64%)	1.597	1.206–1.985	0.029
rs2227307 TT carriage	Non-carriers	237(32.33%)	496(67.67%)	Reference		
	Carriers	163(39.09%)	254(60.91%)	1.328	0.863–1.794	0.081
rs2227307 GT carriage	Non-carriers	207(35.20%)	381(64.80%)	Reference		
	Carriers	193(34.34%)	369(65.66%)	0.959	0.721–1.288	0.163

OR: odds ratio; CI: confidence interval.
Significant p values are in italic bold.
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ers and locus-specific extension primers were designed using MassARRAY Assay Design software package (v. 3.1). 50 nanograms of DNA template were used in each multiplexed PCR well. PCR products were treated with shrimp alkaline phosphatase (USB, Cleveland, OH, USA) prior to use in the iPLEX GOLD primer extension reaction. Single base extension products were desalted with SpectroCLEAN resin (Sequenom), and 10 nL of the desalted product was spotted onto a 384-format SpectroCHIP using the MassARRAY Nanodispenser. Mass determination was performed with a MALDI-TOF mass spectrometer, and MassARRAY Typer 4.0 software was employed for data acquisition. The final genotype call rate of each SNP was greater than 96% and the overall genotyping call rate was 98.1%, confirming the reliability of further statistical analyses.

Statistical analysis and power analysis

Hardy–Weinberg equilibrium (HWE) for each SNP was assessed using GENEPOP v4.0. Allelic and genotypic association tests were performed using CLUMP v2.4 with 10,000 simulations, and this program employed an empirical Monte Carlo test of significance through simulation. To control for possible confounding effects, age and gender were used as independent variables in a multiple logistic regression analysis for adjustment by commercially available software (Statistical Package for Social Sciences, version 16.0 for windows, SPSS Inc., Chicago, IL, USA). The D' values for each pair of markers were calculated using the software program 2LD [42]. Haplotype frequencies were estimated using GENECOUNTING v2.2, which computes maximum-likelihood

estimates of haplotype frequencies from unknown phase data using an expectation–maximization algorithm [43]. The significance of a haplotypic association with chronic periodontitis was evaluated using a likelihood ratio test, followed by permutation testing that compared the estimated haplotype frequencies in patients and controls [43,44]. Differences were considered significant when the p value was less than 0.05. For haplotype analyses, the global p value was based on a comparison of the frequency distribution of all possible combinations of haplotypes among patients and controls. Furthermore, we performed power calculations for case–control genetic association analyses using PGA v2.0 [45]. Our sample size can detect SNP and haplotype associations with 91% and 85% power, respectively, at a false positive rate of 5%, and a presumed minimum odds ratio (OR) of 1.5.

Results

In total, 23 SNPs in the 45 kb region containing IL-8 were genotyped in 400 cases and 750 controls. The allele and genotype frequencies of all in case and control SNPs, including the results of the HWE test, are shown in Table 2 and Table S1. All SNPs were highly polymorphic in both cases and controls, with the exception of 7 SNPs (rs2227528, rs7682639, rs2227531, rs2227532, rs2227538, rs2227549 and rs2227545) in IL-8, and all SNPs were in HWE in both groups. First, we conducted a single SNP association analysis. When all of the samples were considered, we observed a significant association for rs4073 ($p = 0.028251$; OR = 1.207; 95% CI 1.014–1.437), and rs2227307

Table 4. Estimation of LD between each pair of loci within IL-8.

	rs4073	rs2227307	rs2227306	rs2227543	rs1126647
rs4073	-	0.743	0.322	0.028	0.015
rs2227307	0.916	-	0.321	0.039	0.022
rs2227306	0.924	0.981	-	0.097	0.098
rs2227543	0.257	0.322	0.331	-	0.099
rs1126647	0.222	0.284	0.347	0.369	-

D' -value are shown below the subtraction sign, and r^2 -value are shown above the subtraction sign.
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Table 5. Haplotypes frequency and association analysis.

Haplotype				Genecounting (frequency %)				
ID	SNP6	SNP7	SNP8	Case	Control	p-value ¹	correction ²	Global p ³
HAP1	T	T	C	40.3	39.4	0.692		<0.001
<i>HAP2⁴</i>	A	T	C	20.3	15.6	0.005	0.039	
<i>HAP3⁴</i>	A	G	T	10.6	18.2	<0.001	<0.001	
HAP4	T	G	T	13.4	10.3	0.028	0.227	
HAP5	A	G	C	7.02	8.75	0.147		
HAP6	T	G	C	4.15	4.28	0.872		

Significant p-values are in italic bold. Common Haplotypes are shown, if frequency more than 2.5%.

¹Based on 10000 permutations.

²Corrected by Bonferroni.

³Based on comparison of frequency distribution of all haplotypes for the combination of SNPs.

⁴Haplotypes in italics are the significant ones in the study.

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($p = 0.003455$; OR = 1.312; 95% CI 1.098–1.567). Genotype association analyses for the two SNPs suggested a similar pattern with a significant p value ($p = 0.014604$, $p = 0.008132$, respectively). Because we observed significant associations for rs4073 in the recessive model ($p = 0.004$) and rs2227307 in the dominant, recessive and co-dominant models ($p = 0.007$, 0.021 and 0.035, respectively) (Table 2), a multiple logistic regression analysis was used for rs4073 in the recessive model and used for rs2227307 in the dominant, recessive and co-dominant models to evaluate the associations of alleles or genotypes with chronic periodontitis susceptibility, while adjusting for modifying factors, such as subject age and gender, to control for possible confounding effects (Table 3). A significant association was observed between age and chronic periodontitis ($p = 0.002$), while gender was not associated with chronic periodontitis (Table 3). The rs4073 TT genotype and rs2227307 T allele were identified as significant risk factors for chronic periodontitis after adjustment for age and gender (OR = 1.451, 95% CI = 1.196–1.628, $P = 0.017$; OR = 1.597, 95% CI = 1.206–1.985, $P = 0.029$; respectively) (Table 3).

Table 4 presents the results of LD tests (noted as D' and r^2) between pairs of SNP markers within IL-8 for the respective control groups. According to these results, LD ($D' > 0.8$) was observed in the five-SNP linkage disequilibrium estimation. When combining the allele frequency data with the LD, the associated SNPs, rs4073 and rs2227307, were detected in the same LD block as rs2227306 ($D' > 0.8$ between them, Table 4). Next, we performed the haplotypic association analysis of the LD block mentioned above (Table 5). Tests of the common three-marker haplotype (frequency > 0.025) association for rs4073, rs2227307, and rs2227306 indicated a significant association between these SNPs and chronic periodontitis ($p < 0.001$, global permutation). Some haplotypes of these three SNPs were significantly associated with chronic periodontitis. For example, HAP2 (ATC) was significantly associated with chronic periodontitis, and its frequency increased nearly 1.5-fold in cases (corrected $p = 0.039$). Due to higher frequencies in the controls, HAP3 (AGT) may provide a protective effect with nearly a 1.8-fold prevalence in controls (corrected $p < 0.001$) (Table 5).

To examine whether gender would play a key role in the association, we analyzed our data by separating females and males according to the above results. Neither rs4073 nor rs2227307 displayed gender-specific associations with chronic periodontitis (Table 6). Moreover, haplotype association analyses were similar

to the single-SNP analysis results, revealing no gender-specific association in females or males (Table 7).

Discussion

IL-8 is a chemokine related to the initiation and amplification of acute inflammatory responses and the chronic inflammatory process [46]. The purpose of this study was to explore the relationship between all SNPs within the IL-8 gene and chronic periodontitis in Han Chinese individuals. In this study, we present evidence for the association of markers that are mapped to the 45 kb region of IL-8 gene with chronic periodontitis. We identified a significant association in the region between rs4073 and rs2227306, and several lines of evidence suggest that the observed association is unlikely to be an artifact. First, both the single SNP and the haplotype-based association analyses support the association. Second, population stratification is an unlikely explanation because all of our samples are from the same geographical region. Lastly, similar results were obtained from other genetically independent populations in previous studies [25,26,28,47], reaffirming the observed association.

In association studies, it is critical to identify common risk variants in different populations. To examine whether common risk variants exist in genetically independent populations, we compared our results with those of previous studies. Individual differences in interleukins levels can be attributed to gene polymorphisms, especially when these polymorphisms are located within exons or promoters. The common rs4073 A/T polymorphism in the IL-8 promoter influences the production and expression of IL-8. The rs4073 A allele increases IL-8 production both in vitro and in vivo [48,49]. Lee et al. [49] reported that the presence of the rs4073 T allele of IL-8 exerts a 2–5-fold higher transcriptional activity than the rs4073 A allele. Two additional studies evaluated the association of the IL-8 rs4073 A/T polymorphism and periodontitis, but contradictory results were obtained [25,50]. Kim et al. [29] found no association between the genotype distribution and allele frequency of this gene and chronic periodontitis in the Brazilian population. In contrast, Andia et al. [25] discovered a significant association between the IL-8 rs4073 A/T polymorphism and chronic periodontitis in Brazilian nonsmokers, with a higher frequency of the A allele in the disease group compared to the control group. The results of our study do not agree with these studies. We found that the A allele of rs4073 displayed a tendency to be lower in cases compared to controls in

Table 6. Allele and genotype association analysis in female and male.

Marks	Allele frequency (%)		p value ¹	Genotype frequency (%)			H-W E P value	p value ¹	OR ² , 95%CI	p value			
	A	T		AA	AT	TT				Dominant	Recessive	Co-dominant	
rs4073													
Female	Case	157(39.3)	243(60.7)	0.110589	34(17.1)	89(44.4)	77(38.5)	0.327	0.103541	1.223	0.735	0.066	0.711
	Control	331(44.1)	419(55.9)		68(18.1)	195(52.0)	112(29.9)	0.290		(0.955–1.565)			
Male	Case	161(40.3)	239(59.7)	0.126923	37(18.3)	88(44.0)	75(37.7)	0.226	0.139648	1.192	0.838	0.054	0.633
	Control	337(44.9)	413(55.1)		72(19.3)	192(51.2)	111(29.5)	0.501		(0.932–1.525)			
rs2227307													
Female	Case	142(35.5)	258(64.5)	0.035592	25(12.5)	92(46.0)	83(41.5)	0.950	0.111624	1.309	0.045	0.881	0.238
	Control	314(41.9)	436(58.1)		67(17.9)	180(48.0)	128(34.1)	0.784		(1.018–1.682)			
Male	Case	140(34.9)	260(65.1)	0.044555	19(9.7)	101(50.4)	80(39.9)	0.123	0.063942	1.316	0.031	0.127	0.070
	Control	308(41.1)	442(58.9)		60(15.9)	189(50.4)	126(33.7)	0.427		(1.023–1.693)			
rs2227306													
Female	Case	290(72.5)	110(27.5)	0.191898	105(52.5)	80(40.0)	15(7.5)	0.965	0.398158	1.196	0.522	0.183	0.794
	Control	516(68.8)	234(31.2)		175(46.7)	166(44.2)	34(9.1)	0.567		(0.914–1.564)			
Male	Case	284(71.1)	116(28.9)	0.186387	104(52.1)	76(38.0)	20(9.9)	0.287	0.273542	1.195	0.803	0.113	0.774
	Control	504(67.2)	246(32.8)		169 (45.1)	166(44.2)	40(10.7)	0.959		(0.917–1.557)			

OR: odds ratio; CI: confidence interval.

Significant p values are in italic bold. CI: confidence interval; OR: odds ratio.

¹p values of the normal chi-square statistics from Monte Carlo stimulation using CLUMP (T1), and significant p values are in italic bold.

²OR refers to risk allele odds ratio in cases and controls.

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Table 7. Haplotypes frequency and association analysis in female and male.

Haplotype	Female		Male		p value ¹	
	Case	Control	Case	Control	Female	Male
rs4073- rs2227307- rs2227306						
ATC	21.2	16.4	19.4	14.8	<i>0.042</i>	<i>0.041</i>
AGT	11.3	18.1	9.65	18.5	<i><0.001</i>	<i><0.001</i>
	Global-p value <i><0.001</i>					<i>0.039</i>
						<i><0.001</i>

¹Based on 10000 permutations, and significant p values are in italic bold.
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the Han Chinese population. Collectively, the consistency between these studies in different ethnic populations provides strong evidence that the rs4073 polymorphism in the IL-8 gene may be involved in chronic periodontitis susceptibility. Additionally, we also observed other differences among these studies. Rs2227307 had a significant allelic and genotypic association with chronic periodontitis in our analysis; however, only genotypic association data were similar to that reported by Scarel-Caminaga et al. [28]. To evaluate potential confounding effects that could cause bias in this association study of chronic periodontitis, important factors known to influence the pathogenesis of chronic periodontitis were assessed by multiple logistic regression analysis. Multiple logistic regression analysis revealed that age was associated with chronic periodontitis (Table 3). Regarding age, an important risk factor for periodontitis, Grossi et al. (1995) observed an OR = 2.6 (95% CI: 1.75–3.83) for the age group 35–44 years and an OR = 24.08 (95% CI: 15.93–36.29) for the age group 65–74 years, which are similar to the results of our study (Table 3) [51]. There is evidence that both the prevalence and severity of chronic periodontitis increase with increasing age [52]. However, the age effect could conceivably represent the cumulative effect of prolonged exposure to other risk factors [53].

The ability to draw conclusions regarding associations based on the analysis of individual SNPs is limited [54]. Therefore, to obtain stronger statistical evidence, we performed a haplotype analysis. Haplotype analysis uses additional information regarding the linkage between typed markers. The results of haplotype frequency estimation for three-SNP combinations (rs4073-rs2227307-rs2227306) indicated significant associations with chronic periodontitis (p<0.001, global permutation). The frequency of the ATC haplotype was 20.3% in cases and 15.6% in controls, demonstrating that individuals with ATC were almost 1.5 times more likely to develop chronic periodontitis than individuals carrying other haplotypes. This result was consistent with that reported by Scarel-Caminaga et al. [28]. However, a protective haplotype, AGT, which was almost 1.8 times more prevalent in controls than in cases, displayed a significant positive association with chronic periodontitis in our studies. Additionally, no gender-specific associations were observed in single SNP or haplotype analyses.

Genetic backgrounds vary among ethnic populations; therefore, differences in the results among these studies might be the result of ethnic differences and the genetic heterogeneity that existing in the IL-8 gene, despite some similarities in general association patterns. Nevertheless, our results should be validated in other populations or with a larger sample size in this population. This validation is required because the statistically significant results could occur by chance, and because the associations were not significant after an adjustment for multiple testing, despite remaining significant after 10,000 permutation tests. In addition, SNPs and haplotype structures can vary amongst different ethnic groups. Therefore, our data should be interpreted with caution, as the combination of these polymorphisms with others in different genes may also be important to define the role of these polymorphisms in the pathogenesis of chronic periodontitis.

A major limitation of the current study was that we did not perform further replication analyses for the possible risk of the SNPs identified in the study due to the lack of another cohort of patients and controls. However, as a replication study to confirm the previous studies and providing further evidence for the association of IL-8 with chronic periodontitis, our study did not appear extremely heterogeneous, and it could be considered reasonable and reliable. Additionally, although we selected subjects with no migration history within the previous three

generations to control population stratification caused by genetic factors, we did not focus on other possible factors leading to population stratification; thus, we cannot completely rule out hidden stratification interference. Therefore, our findings should be considered preliminary. Given that the pathomechanism of chronic periodontitis includes further cytokines, chemokines, and pattern-associated molecules, additional follow-up studies are required to find possible causal variants. Ideally, these studies will involve the use of more sophisticated techniques of genetic investigation, such as DNA microarrays, to better understand the involvement of genetic factors in chronic periodontitis. Additionally, it is important to investigate the interaction of host genetics with clinical parameters and the immune response because chronic periodontitis is a complex disease, and the interaction of different factors has been insufficiently evaluated.

In recent years, particular interest has been given to investigating functional polymorphisms in candidate genes for disease association. The expression levels of a protein may be modulated by genetic polymorphisms in regulatory regions of the gene, mainly the promoter region [55]. Considering that some IL-8 polymorphisms were previously reported to be associated with higher IL-8 production and that a significantly higher level of this protein was found in the gingival crevicular fluid from patients with periodontitis [56], we hypothesize that the genetic differences between individuals in relation to IL-8 production may somehow predispose them to periodontal disease. The significantly associated SNPs identified in our study are not randomly distributed over the gene. Rather, these SNPs in the promoter and intron 1 are in the same LD block spanning the promoter and exon 1 of IL-8. Therefore, the significant associations in our study may be of interest for future studies. First, there is a cluster of significantly associated SNPs that span the promoter and exon 1 of IL-8, indicating a region of interest that might harbor functionally relevant variants. Second, we provide additional data in agreement with the previously reported IL-8 polymorphisms that are associated with chronic periodontitis susceptibility. To remedy the

mentioned shortcomings, we will try to observe the effects of these polymorphisms on periodontal systemic therapy in future studies.

In summary, our work provides supportive evidence for the association of IL-8 with chronic periodontitis. To our knowledge, this is the first study to demonstrate all SNPs between 20 kb upstream and 20 kb downstream of the IL-8 gene with chronic periodontitis in the Han Chinese population. Moreover, as an intriguing new insight into the pathogenesis of chronic periodontitis, we also confirmed previous reports suggesting that this gene plays an important role in the etiology of this condition. Because chronic periodontitis is multifactorial in nature, involving interactions between genes, the environment and lifestyle, genetic periodontal risk assessments may be valuable in developing preventive, diagnostic and therapeutic strategies against the incidence and progression of this condition. Given the complex patterns of findings from association studies focusing on chronic periodontitis and its underlying genetic heterogeneity, further inquiries and wider replications are required, particularly within different ethnic samples.

Supporting Information

Table S1 Allele and genotype frequency of other 13 SNPs association analyses.

(DOC)

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Author Contributions

Conceived and designed the experiments: NZ. Performed the experiments: NZ DXZ HJY. Analyzed the data: YHX TXZ Bo Zhang. Contributed reagents/materials/analysis tools: Bao Zhang ZFF. Wrote the paper: NZ.

References

- Schenkein HA (2006) Host responses in maintaining periodontal health and determining periodontal disease. *Periodontol* 2000 40: 77–93.
- Bartold PM (2006) Periodontal tissues in health and disease: introduction. *Periodontol* 2000 40: 7–10.
- Kinane DF, Peterson M, Stathopoulou PG (2006) Environmental and other modifying factors of the periodontal diseases. *Periodontol* 2000 40: 107–119.
- Erdemir EO, Duran I, Haliloglu S (2004) Effects of smoking on clinical parameters and the gingival crevicular fluid levels of IL-6 and TNF-alpha in patients with chronic periodontitis. *J Clin Periodontol* 31: 99–104.
- Palmer RM, Wilson RF, Hasan AS, Scott DA (2005) Mechanisms of action of environmental factors—tobacco smoking. *J Clin Periodontol* 32 Suppl 6: 180–195.
- Gautam DK, Jindal V, Gupta SC, Tuli A, Kotwal B, et al. (2011) Effect of cigarette smoking on the periodontal health status: A comparative, cross sectional study. *J Indian Soc Periodontol* 15: 383–387.
- Dutra WO, Moreira PR, Souza PE, Gollob KJ, Gomez RS (2009) Implications of cytokine gene polymorphisms on the orchestration of the immune response: lessons learned from oral diseases. *Cytokine Growth Factor Rev* 20: 223–232.
- Gera I, Vari M (2009) [Genetic background of periodontitis. Part II. Genetic polymorphism in periodontal disease. A review of literature]. *Fogorv Sz* 102: 131–140.
- Laine ML, Loos BG, Crielaard W (2010) Gene polymorphisms in chronic periodontitis. *Int J Dent* 2010: 324719.
- Loos BG, John RP, Laine ML (2005) Identification of genetic risk factors for periodontitis and possible mechanisms of action. *J Clin Periodontol* 32 Suppl 6: 159–179.
- Nikolopoulos GK, Dimou NL, Hamodrakas SJ, Bagos PG (2008) Cytokine gene polymorphisms in periodontal disease: a meta-analysis of 53 studies including 4178 cases and 4590 controls. *J Clin Periodontol* 35: 754–767.
- Kinane DF, Hart TC (2003) Genes and gene polymorphisms associated with periodontal disease. *Crit Rev Oral Biol Med* 14: 430–449.
- Korhonen KS, di Giovine FS (1998) Genetic variations in cytokine expression: a risk factor for severity of adult periodontitis. *Ann Periodontol* 3: 327–338.
- Ferreira SB Jr, Trombone AP, Repeke CE, Cardoso CR, Martins W Jr, et al. (2008) An interleukin-1beta (IL-1beta) single-nucleotide polymorphism at position 3954 and red complex periodontopathogens independently and additively modulate the levels of IL-1beta in diseased periodontal tissues. *Infect Immun* 76: 3725–3734.
- Kamma JJ, Giannopoulou C, Vasdekis VG, Mombelli A (2004) Cytokine profile in gingival crevicular fluid of aggressive periodontitis: influence of smoking and stress. *J Clin Periodontol* 31: 894–902.
- Trombone AP, Cardoso CR, Repeke CE, Ferreira SB Jr, Martins W Jr, et al. (2009) Tumor necrosis factor-alpha -308G/A single nucleotide polymorphism and red-complex periodontopathogens are independently associated with increased levels of tumor necrosis factor-alpha in diseased periodontal tissues. *J Periodontol Res* 44: 598–608.
- Remick DG (2005) Interleukin-8. *Critical Care Medicine* 33(12 Suppl): S466–467.
- Marshall JC (2005) Neutrophils in the pathogenesis of sepsis. *Critical Care Medicine* 33(12 Suppl): S502–504.
- DeForge LE, Fantone JC, Kenney JS, Remick DJ (1992) Oxygen radical scavengers selectively inhibit interleukin 8 production in human whole blood. *Journal of Clinical Investigation* 90(5): 2123–2129.
- Guan FL, Zhang C, Wei SG, Zhang HB, Gong XM, et al. (2012) Association of PDE4B polymorphisms and schizophrenia in Northwestern Han Chinese. *Human Genetics* 131(7): 1047–1056.
- Guan FL, Wei SG, Feng JL, Zhang C, Xing B, et al. (2012) Association study of a new schizophrenia susceptibility locus of 10q24.32-33 in a Han Chinese population. *Schizophrenia Research*, 138(1): 63–68.
- Guan FL, Wei SG, Zhang C, Zhang HB, Zhang B, et al. (2013) A population-based association study of 2q32.3 and 8q21.3 loci with schizophrenia in Han Chinese. *Journal of Psychiatric Research* 47(6): 712–717.
- Guan FL, Zhang B, Yan TL, Li L, Liu F, et al. (2014) MIR137 gene and target gene CACNA1C of miR-137 contribute to schizophrenia susceptibility in Han Chinese. *Schizophrenia Research* 152(1): 97–104.

24. Amaya MP, Criado L, Blanco B, Gomez M, Torres O, et al. (2013) Polymorphisms of pro-inflammatory cytokine genes and the risk for acute suppurative or chronic nonsuppurative apical periodontitis in a Colombian population. *Int Endod J* 46: 71–78.
25. Andia DC, de Oliveira NF, Letra AM, Nociti FH Jr, Line SR, et al. (2011) Interleukin-8 gene promoter polymorphism (rs4073) may contribute to chronic periodontitis. *J Periodontol* 82: 893–899.
26. Kim YJ, Viana AC, Curtis KM, Orrico SR, Cirelli JA, et al. (2010) Association of haplotypes in the IL-8 gene with susceptibility to chronic periodontitis in a Brazilian population. *Clin Chim Acta* 411: 1264–1268.
27. Li G, Yue Y, Tian Y, Li JL, Wang M, et al. (2012) Association of matrix metalloproteinase (MMP)-1, 3, 9, interleukin (IL)-2, 8 and cyclooxygenase (COX)-2 gene polymorphisms with chronic periodontitis in a Chinese population. *Cytokine* 60: 552–560.
28. Scarel-Caminaga RM, Kim YJ, Viana AC, Curtis KM, Corbi SC, et al. (2011) Haplotypes in the interleukin 8 gene and their association with chronic periodontitis susceptibility. *Biochem Genet* 49: 292–302.
29. Kim YJ, Viana AC, Curtis KM, Orrico SR, Cirelli JA, et al. (2009) Lack of association of a functional polymorphism in the interleukin 8 gene with susceptibility to periodontitis. *DNA Cell Biol* 28: 185–190.
30. Andia DC, Letra A, Casarin RC, Casati MZ, Line SR, et al. (2012) Genetic analysis of the IL-8 gene polymorphism (rs4073) in generalized aggressive periodontitis. *Arch Oral Biol*.
31. Albandar JM, Rams TE (2002) Risk factors for periodontitis in children and young persons. *Periodontol* 2000 29: 207–222.
32. Fan WH, Liu DL, Xiao LM, Xie CJ, Sun SY, et al. (2011) Coronary heart disease and chronic periodontitis: is polymorphism of interleukin-6 gene the common risk factor in a Chinese population? *Oral Dis* 17: 270–276.
33. Moreira PR, de Sa AR, Xavier GM, Costa JE, Gomez RS, et al. (2005) A functional interleukin-1 beta gene polymorphism is associated with chronic periodontitis in a sample of Brazilian individuals. *J Periodontol Res* 40: 306–311.
34. Nibali L, Madden I, Franch Chillida F, Heitz-Mayfield L, Brett P, et al. (2011) IL6 –174 genotype associated with *Aggregatibacter actinomycetemcomitans* in Indians. *Oral Dis* 17: 232–237.
35. Scarel-Caminaga RM, Trevisatto PC, Souza AP, Brito RB, Camargo LE, et al. (2004) Interleukin 10 gene promoter polymorphisms are associated with chronic periodontitis. *J Clin Periodontol* 31: 443–448.
36. Apatzidou DA, Riggio MP, Kinane DF (2005) Impact of smoking on the clinical, microbiological and immunological parameters of adult patients with periodontitis. *J Clin Periodontol* 32: 973–983.
37. Bergstrom J (2004) Tobacco smoking and chronic destructive periodontal disease. *Odontology* 92: 1–8.
38. Kornman KS (2005) Diagnostic and prognostic tests for oral diseases: practical applications. *J Dent Educ* 69: 498–508.
39. Armitage GC (1999) Development of a classification system for periodontal diseases and conditions. *Ann Periodontol* 4: 1–6.
40. Highfield J (2009) Diagnosis and classification of periodontal disease. *Aust Dent J* 54 Suppl 1: S11–26.
41. Barrett JC, Fry B, Maller J, Daly MJ (2005) Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 21: 263–265.
42. Zhao JH (2004) 2LD, GENECOUNTING and HAP: Computer programs for linkage disequilibrium analysis. *Bioinformatics* 20: 1325–1326.
43. Curtis D, Knight J, Sham PC (2006) Program report: GENECOUNTING support programs. *Ann Hum Genet* 70: 277–279.
44. Zhao JH, Lissarrague S, Essioux L, Sham PC (2002) GENECOUNTING: haplotype analysis with missing genotypes. *Bioinformatics* 18: 1694–1695.
45. Menashe I, Rosenberg PS, Chen BE (2008) PGA: power calculator for case-control genetic association analyses. *BMC Genet* 9: 36.
46. Campa D, Hung RJ, Mates D, Zaridze D, Szeszenia-Dabrowska N, et al. (2005) Lack of association between –251 TNA polymorphism of IL8 and lung cancer risk. *Cancer Epidemiol Biomarkers Prev* 14: 2457–2458.
47. Corbi SC, Anovazzi G, Finoti LS, Kim YJ, Capela MV, et al. (2012) Haplotypes of susceptibility to chronic periodontitis in the Interleukin 8 gene do not influence protein level in the gingival crevicular fluid. *Arch Oral Biol* 57: 1355–1361.
48. Taguchi A, Ohmiya N, Shirai K, Mabuchi N, Itoh A, et al. (2005) Interleukin-8 promoter polymorphism increases the risk of atrophic gastritis and gastric cancer in Japan. *Cancer Epidemiol Biomarkers Prev* 14: 2487–2493.
49. Lee WP, Tai DI, Lan KH, Li AF, Hsu HC, et al. (2005) The –251T allele of the interleukin-8 promoter is associated with increased risk of gastric carcinoma featuring diffuse-type histopathology in Chinese population. *Clin Cancer Res* 11: 6431–6441.
50. Cipollone F, Toniato E, Martinotti S, Fazio M, Iezzi A, et al. (2004) A polymorphism in the cyclooxygenase 2 gene as an inherited protective factor against myocardial infarction and stroke. *JAMA* 291: 2221–2228.
51. Grossi SG, Genco RJ, Machtei EE, Ho AW, Koch G, et al. (1995) Assessment of risk for periodontal disease. II. Risk indicators for alveolar bone loss. *J Periodontol* 66: 23–29.
52. Borrell LN, Papapanou PN (2005) Analytical epidemiology of periodontitis. *J Clin Periodontol* 32 Suppl 6: 132–158.
53. Papapanou PN, Lindhe J, Sterrett JD, Eneroth L (1991) Considerations on the contribution of ageing to loss of periodontal tissue support. *J Clin Periodontol* 18: 611–615.
54. Korostishevsky M, Kaganovich M, Cholostoy A, Ashkenazi M, Ratner Y, et al. (2004) Is the G72/G30 locus associated with schizophrenia? single nucleotide polymorphisms, haplotypes, and gene expression analysis. *Biol Psychiatry* 56: 169–176.
55. Stern DL (2000) Evolutionary biology. The problem of variation. *Nature* 408: 529–531.
56. Tsai CC, Ho YP, Chen CC (1995) Levels of interleukin-1 beta and interleukin-8 in gingival crevicular fluids in adult periodontitis. *J Periodontol* 66: 852–859.