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Draft Genome Sequence of the *bla*_{OXA-436}⁻ and *bla*_{NDM-1}⁻-Harboring *Shewanella putrefaciens* SA70 Isolate

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ABSTRACT We sequenced a carbapenem-resistant *Shewanella putrefaciens* isolate cultured from the sink handle of a Pakistan hospital room. Assembly annotation indicates that the isolate has a chromosomal *bla*_{OXA-436} carbapenemase and a plasmid-borne *bla*_{NDM-1} gene. To our knowledge, this is the first report of a *Shewanella* species harboring *bla*_{NDM-1}.

Shewanella putrefaciens is an infrequent human pathogen associated with opportunistic infections (1–3). We isolated *S. putrefaciens* from the sink handle of an intensive care unit room from a tertiary care hospital in Pakistan using the Eswab collection device and transport system (Becton, Dickinson, and Company). The isolate was recovered on MacConkey agar with 5 μg/mL cefotaxime (Hardy Diagnostics) and identified with a confidence value of 99.9% using VITEK matrix-assisted laser desorption ionization–time of flight mass spectrometry (MALDI-TOF MS) with library version v2.3.3 (bioMérieux, Durham, NC). Antimicrobial susceptibility testing using Kirby-Bauer disk diffusion per Clinical and Laboratory Standards Institute M2 guidelines demonstrated an aztreonam zone size of 30 mm, ceftazidime zone size of 6 mm, meropenem zone size of 6 mm, and imipenem zone size of 6 mm. Using *Enterobacteriaceae*, *Pseudomonas aeruginosa*, or *Aeromonas* sp. guidelines, the isolate is considered aztreonam susceptible but ceftazidime, meropenem, and imipenem resistant, a resistance profile suggestive of a Class B carbapenemase. The isolate was also positive for phenotypic production of a carbapenemase via the carbapenem inactivation method (4). To identify known carbapenemase genes in *S. putrefaciens*, we extracted genomic DNA with the Bacteremia kit (MoBio, Carlsbad, CA), used 0.5 ng of DNA to make a Nextera XT Illumina sequencing library (5), and performed whole-genome sequencing on an Illumina NextSeq high-output sequencer to obtain 1,454,810 2 × 150 bp reads.

We processed the sequence data on the High Throughput Computing Facility 2250 processor computing cluster at Washington University, St. Louis School of Medicine. Nextera indexes and sequencing primers were trimmed from forward and reverse reads using Trimmomatic, and human genome sequence reads, which may bleed through from libraries on the same lane, were removed using DeconSeq (6, 7). The resulting 1,452,994 paired-end reads were assembled with SPAdes (8). QUAST determined that the assembly contained 302 contigs (354,418 bp largest contig) and 70,827 bp *N*₅₀ for contigs with >0 nucleotides (9). We annotated contigs with lengths >500 bp, representing 5,110,564 bp, using RAST (10). RAST identified 10 rRNA genes, 71 tRNA genes,

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and 4,522 CDS. Fifty percent of CDS have annotated functional systems. We identified antibiotic resistance genes with Resfams, finding two class D β -lactamases and two class B β -lactamases (11). BLASTp of the amino acid sequences against the NCBI nonredundant protein database (accessed 14 March 2017) confirmed these genes as *bla*_{OXA-436} (99% identity), *bla*_{OXA-10r}, a metallo-fold hydrolase, and *bla*_{NDM-1} (all 100% identity). OXA-436 (GenBank accession no. KT959103) and NDM-1 can hydrolyze carbapenems, implicating them as the mechanism of carbapenem resistance (12). BLASTn of the *bla*_{NDM-1} contig against the nonredundant nucleotide database (accessed on 14 March 2017) returned 100% identity to *bla*_{NDM-1} containing IncN type plasmids from *Escherichia coli* (KJ440076) and *Klebsiella pneumoniae* (KJ440075) (13). The *bla*_{OXA-436} carbapenemase contig had the highest BLASTn alignment score to the complete genome of *Shewanella* sp. MR-7 (CP000444).

This *Shewanella putrefaciens* genome documents the diversity of host species possessing *bla*_{NDM-1}. Identification of carbapenemases in this isolate supports the importance of antimicrobial susceptibility testing in infrequent pathogens. Importantly, *Shewanella* spp. could represent potential reservoirs for horizontally transferred antimicrobial resistance genes within the health care environment.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession no. [NGZL00000000](https://doi.org/10.1093/genomea.121357). The version described in this paper is NGZL01000000.

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REFERENCES

- Janda JM, Abbott SL. 2014. The genus *Shewanella*: from the briny depths below to human pathogen. *Crit Rev Microbiol* 40:293–312. <https://doi.org/10.3109/1040841X.2012.726209>.
- Wall JD, Krumholz LR. 2006. Uranium reduction. *Annu Rev Microbiol* 60:149–166. <https://doi.org/10.1146/annurev.micro.59.030804.121357>.
- Tsai MS, You HL, Tang YF, Liu JW. 2008. *Shewanella* soft tissue infection: case report and literature review. *Int J Infect Dis* 12:e119–e124. <https://doi.org/10.1016/j.ijid.2008.03.020>.
- McMullen AR, Yarbrough ML, Wallace MA, Shupe A, Burnham CD. 2017. Evaluation of genotypic and phenotypic methods to detect carbapenemase production in Gram-negative bacilli. *Clin Chem* 63:723–730. <https://doi.org/10.1373/clinchem.2016.264804>.
- Baym M, Kryazhimskiy S, Lieberman TD, Chung H, Desai MM, Kishony R. 2015. Inexpensive multiplexed library preparation for megabase-sized genomes. *PLoS One* 10:e0128036. <https://doi.org/10.1371/journal.pone.0128036>.
- Schmieder R, Edwards R. 2011. Fast identification and removal of sequence contamination from genomic and metagenomic datasets. *PLoS One* 6:e17288. <https://doi.org/10.1371/journal.pone.0017288>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankевич A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Gibson MK, Forsberg KJ, Dantas G. 2015. Improved annotation of antibiotic resistance determinants reveals microbial resistomes cluster by ecology. *ISME J* 9:207–216. <https://doi.org/10.1038/ismej.2014.106>.
- Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, Walsh TR. 2009. Characterization of a new metallo-beta-lactamase gene, bla(NDM-1), and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrob Agents Chemother* 53:5046–5054. <https://doi.org/10.1128/AAC.00774-09>.
- Chen CJ, Wu TL, Lu PL, Chen YT, Fung CP, Chuang YC, Lin JC, Siu LK. 2014. Closely related NDM-1-encoding plasmids from *Escherichia coli* and *Klebsiella pneumoniae* in Taiwan. *PLoS One* 9:e104899. <https://doi.org/10.1371/journal.pone.0104899>.