2018

Mutations in the PH Domain of DNM1 are associated with a nonepileptic phenotype characterized by developmental delay and neurobehavioral abnormalities

Emily Brereton  
*Washington University School of Medicine in St. Louis*

Emily Fassi  
*Washington University School of Medicine in St. Louis*

Gabriel C. Araujo  
*St. Louis Children's Hospital*

Jonathan Dodd  
*St. Louis Children's Hospital*

Aida Telegrafi  
*GeneDx*

See next page for additional authors

Follow this and additional works at: https://digitalcommons.wustl.edu/open_access_pubs

Recommended Citation  
Brereton, Emily; Fassi, Emily; Araujo, Gabriel C.; Dodd, Jonathan; Telegrafi, Aida; Pathak, Sheel J.; and Shinawi, Marwan, "Mutations in the PH Domain of DNM1 are associated with a nonepileptic phenotype characterized by developmental delay and neurobehavioral abnormalities." Molecular Genetics and Genomic Medicine.6,2. 294-300. (2018).  
https://digitalcommons.wustl.edu/open_access_pubs/6856

This Open Access Publication is brought to you for free and open access by Digital Commons@Becker. It has been accepted for inclusion in Open Access Publications by an authorized administrator of Digital Commons@Becker. For more information, please contact engeszer@wustl.edu.
Mutations in the PH Domain of *DNM1* are associated with a nonepileptic phenotype characterized by developmental delay and neurobehavioral abnormalities

Emily Brereton¹ | Emily Fassi² | Gabriel C. Araujo³ | Jonathan Dodd³ | Aida Telegafi⁴ | Sheel J. Pathak⁵ | Marwan Shinawi¹,²

¹Washington University School of Medicine, St. Louis, MO, USA
²Division of Genetics and Genomic Medicine, Department of Pediatrics, Washington University School of Medicine, St. Louis, MO, USA
³Department of Psychology, St Louis Children’s Hospital, St. Louis, MO, USA
⁴GeneDx, Gaithersburg, MD, USA
⁵Department of Neurology, Washington University School of Medicine, St. Louis, MO, USA

**Abstract**

**Background:** Dynamin 1 is a protein involved in the synaptic vesicle cycle, which facilitates the exocytosis of neurotransmitters necessary for normal signaling and development in the central nervous system. Pathogenic variants in *DNM1* have been implicated in global developmental delay (DD), severe intellectual disability (ID), and notably, epileptic encephalopathy. All previously reported *DNM1* pathogenic variants causing this severe phenotype occur in the GTPase and Middle domains of the dynamin 1 protein.

**Methods:** We used whole-exome sequencing to characterize the molecular basis of DD and autistic symptoms in two identical siblings.

**Results:** The twin siblings exhibit mild to moderate ID and autistic symptoms but no epileptic encephalopathy. Exome sequencing revealed a genetic variant, c.1603A>G (p.Lys535Glu), in the PH domain of dynamin 1. Previous in vitro studies showed that mutations at Lys535 inhibit endocytosis and impair PH loop binding to PIP2.

**Conclusions:** Our data suggest a previously undescribed milder phenotype associated with a missense genetic variant in the PH domain of dynamin 1.

**KEYWORDS**
developmental delay, domain, dynamin 1, epileptic encephalopathy, intellectual disability, synaptic vesicle

---

**1 | BACKGROUND**

Neurotransmission in the central nervous system depends on synaptic vesicle transport. Neurotransmitters are taken up by exocytotic vesicles which fuse with the plasma membrane, releasing neurotransmitters into the synapse (Sudhof, 2004). Neurotransmitters in the synapse are degraded or taken up into the presynaptic axon. Vesicle endocytosis, when vesicles bud off from the plasma membrane into the presynaptic axon, is mediated by proteins including amphipathin, endophilin, clathrin, and dynamin (Rizzoli, 2014). Newly synthesized or retrieved neurotransmitters enter these new vesicles and restart the cycle (Takei, 1996).

The Dynamin family of proteins interact with clathrin in the vesicle budding cycle and participate in the scission between the cell membrane and vesicle wall. After the invagination of the lipid membrane, dynamin binds GTP and its alpha helixes wrap around and pinch the budding
vesicle. Next, dynamin uses the energy from GTP hydrolysis to sever the membrane tubules and complete vesicle fission (Antonny et al., 2016). This process is critical for the synaptic vesicle cycle (McNiven, 2000). Dynamin proteins have five domains; the GTPase domain is the largest and best understood, followed by a middle domain, pleckstrin homology (PH) domain, coiled coil (CC) domain, and a proline-rich domain (PRD) (McNiven, 2000). The PH domain is thought to interact directly with the lipid bilayer (Vallis, Wigge, Marks, Evans, & McMahon, 1999).

The \textit{DNM1} gene (MIM 602377) codes for dynamin 1 and is mainly expressed in the central nervous system (Romeu & Arola, 2014). Pathogenic \textit{DNM1} variants affect brain development and function and cause epileptic encephalopathy associated with global developmental delay (DD) (Allen et al., 2016; Deng et al., 2016; Epi4K Consortium; Epilepsy Phenome/Genome Project, 2013; Euro E-RESC, EpilepsyPhenome/Genome P, Epi4K Consortium, 2014; Nakashima et al., 2016). Previously reported patients with \textit{DNM1}-related disorder carry pathogenic variants in the GTPase or middle domains, but never in the PH domain (Allen et al., 2016; Deng et al., 2016; Epi4K Consortium; Epilepsy Phenome/Genome Project, 2013; Euro E-RESC, EpilepsyPhenome/Genome P, Epi4K Consortium, 2014; Nakashima et al., 2016).

Here, we report 8-year-old identical twin sisters with a de novo p.Lys535Glu likely pathogenic novel variant in exon 15 of \textit{DNM1} (NM_004408.3), the first documented pathogenic variant in the PH domain. Although all previously reported patients with \textit{DNM1} pathogenic variants exhibited epileptic encephalopathy and severe neurological phenotype, the two probands in this report had no seizures and exhibited mild to moderate DD/intellectual disability (ID) and autism spectrum disorder. Our data suggest an emerging and previously undescribed phenotype associated with a specific \textit{DNM1} missense genetic variant in the PH domain.

2 | CLINICAL REPORT

Patients 1 and 2 are monozygotic triplet sisters who presented for evaluation of DD, autism spectrum disorder, some dysmorphic features, and hypotonia. They were born at 36 weeks gestation, which was conceived naturally, via an uncomplicated cesarian section secondary to triplet pregnancy. There were no complications after delivery. They have a healthy triplet brother. Patient 1 had a birth weight of 2.8 kg (>97th percentile for triplet gestation), a birth length of 49 cm (>97th percentile), and an occipitofrontal circumference (OFC) of 33.8 cm (80th percentile). Patient 2 had a birth weight of 2.55 kg (50th–90th percentile for triplet gestation), a birth length of 48.2 cm (>97th centile), and OFC of 33.1 cm (65th percentile). Their triplet brother had a birth weight of 2.38 kg (50th–90th percentile for triplet gestation), a birth length of 46.9 cm (97th percentile), and OFC of 32 cm (40th percentile).

Developmental concerns were first noted at 4 months of age when both sisters were noted to lack visual tracking, but this spontaneously resolved at 7 months of age. Patient 1 and 2 started receiving physical, occupational, and speech therapies at 1 year of age due to delayed speech and motor milestones. Both sisters started sitting at 7–8 months, crawling at 11 months, and walking at 18 months. At 18 months, Patient 1 could say 5–10 words, but lost most of this vocabulary by age 2. Patient 2 had delayed language acquisition but no language loss. Their triplet brother started sitting at 6 months, crawling at 9 months and walking at 15 months. He required speech therapy for 9 months during third year of life for mild speech delay. The brother is in mainstream classes, never required any resources at school, and does not exhibit neurobehavioral abnormalities. The sisters were diagnosed with autism at 3 years of age and have been receiving applied behavior analysis (ABA) therapy since. At the age of 5, Patient 2 had one episode of febrile seizure. She had a normal brain MRI after this event. She also had a few staring episodes, but her EEG at 7 years of age was normal.

Developmental evaluation of both sisters at age 8 years revealed that Patient 1 can say 15 words but rarely speaks. She can add and subtract double digits, recognize sight words, and use a few signs. She sometimes becomes agitated and may bite or hit herself or others. Patient 2 can use 50 words, combine 2–3 words, use many signs, write her name, recognize some sight words, and complete some basic addition and subtraction. She occasionally becomes agitated, may hit herself or others, and refuses to cooperate with therapy or schoolwork. Patient 2 receives Ritalin and Clonidine for behavioral problems. A formal neuropsychological assessment was performed at 8 years of age for both sisters and the detailed results are provided in Appendix S1.

On physical exam at 8 years, Patient 1 and 2 had no major dysmorphic features but were each noted to have a broad forehead, smooth philtrum, widely spaced teeth, high-arched palate, and mild retrognathia (Figure 1a,b). Their neurological exam revealed mild axial and appendicular hypotonia and drooling. Patient 1’s weight was 24 kg (24th centile), height was 126.6 cm (28th centile) and OFC was 53.3 cm (85th centile). Patient 2’s weight was 24.7 kg (30th centile), height was 125.4 cm (22nd centile) and OFC was 54.3 cm (95th centile).

The genetic workup of the Patient 1 and 2 started at 3 years of age and included normal karyotype, chromosomal microarray analysis, and fragile X testing. Because of muscle weakness and elevated blood lactate, the two sisters...
were diagnosed with a nonspecific mitochondrial disease. However, mtDNA sequencing and deletion/duplication studies and a mitochondrial nuclear gene panel were normal except for a variant of unknown significance, p.Asp85Asn, in KCNE1. This finding prompted a cardiology consultation and both sisters had a normal echocardiogram and EKG. The exome results (see below) prompted an electroencephalogram which was normal in both sisters.

3 | MATERIALS AND METHODS

The Institutional Review Board of Washington University School of Medicine approved our study protocol and informed consent was obtained from the parents for publication of genetic results and photographs. Clinical whole-exome sequencing was performed using a trio design with one of the identical twin sisters submitted as a proband and the second sibling and their parents used for segregation analysis. For more details see Appendix S2.

4 | MOLECULAR RESULTS

Exome sequencing of the sisters identified a de novo, heterozygous, likely pathogenic variant designated as c.1603A>G (p.Lys535Glu) in DNM1. This variant has not been observed in approximately 6,500 individuals that were sequenced as part of the National Heart, Lung, and Blood Institute (NHLBI) exome sequence project or in ExAC database (http://exac.broadinstitute.org/). Both parents tested negative for this variant.

5 | DISCUSSION

Pathogenic variants in DNM1 have been reported in association with early-onset epileptic encephalopathy (including West and Lennox-Gastaut syndromes), intractable seizures starting within the first year of life, motor impairments, and severe to profound ID (Allen et al., 2016; Deng et al., 2016; Epi4K Consortium; Epilepsy Phenome/Genome Project, 2013; Euro E-RESC, EpilepsyPhenome/Genome P, Epi4K Consortium, 2014; Nakashima et al., 2016) (Table 1). Other rare clinical features reported in some individuals include hypotonia, developmental regression, movement disorder, autism, cortical visual impairment, behavioral concerns, and microcephaly (Allen et al., 2016; Deng et al., 2016; Epi4K Consortium; Epilepsy Phenome/Genome Project, 2013; Euro E-RESC, EpilepsyPhenome/Genome P, Epi4K Consortium, 2014; Nakashima et al., 2016) (Table 1). The two patients described here exhibit a milder phenotype with mild to moderate ID, autistic symptoms, and other than a single febrile seizure in Patient 2, no history of seizures.

The twin sisters studied here share the de novo variant, c.1603A>G (p.Lys535Glu) in exon 15 of DNM1, classified as likely pathogenic. This variant has not previously been reported in patients or databases of healthy controls. It is a nonconservative amino acid substitution that occurs at an evolutionary conserved position (Figure S1). Previous in vitro studies presented below further support the variant’s pathogenicity.

Eight previously reported patients with DNM1-related disorder carry pathogenic variants in the GTPase domain and two carry variants in the middle domain (Figure 2a).

To our knowledge, the monozygotic twin sisters in this
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Domain involved</td>
<td>GTPase</td>
<td>GTPase</td>
<td>GTPase</td>
<td>GTPase</td>
<td>GTPase</td>
<td>GTPase</td>
<td>Middle</td>
<td>Middle</td>
<td>PH</td>
<td>PH</td>
</tr>
<tr>
<td>Gender, age (years)</td>
<td>M, 15</td>
<td>M, 6</td>
<td>M, 5</td>
<td>M, 15</td>
<td>M, 8</td>
<td>M, 6</td>
<td>M, N/A</td>
<td>M, 6</td>
<td>M, N/A</td>
<td>F, 8</td>
</tr>
<tr>
<td>Diagnosis</td>
<td>LGS</td>
<td>Infantile spasms</td>
<td>LGS</td>
<td>Infantile spasms</td>
<td>Infantile spasms</td>
<td>Infantile spasms</td>
<td>West syndrome</td>
<td>Infantile spasms</td>
<td>LGS</td>
<td>Infantile spasms</td>
</tr>
<tr>
<td>Development prior to seizure onset</td>
<td>Global DD</td>
<td>Global DD</td>
<td>Some DD</td>
<td>Probably normal</td>
<td>Some DD noted</td>
<td>Global DD</td>
<td>Global DD</td>
<td>Normal</td>
<td>N/A</td>
<td>Autism, DD</td>
</tr>
<tr>
<td>Seizure onset (months)</td>
<td>11</td>
<td>13</td>
<td>7</td>
<td>7</td>
<td>6</td>
<td>12</td>
<td>10</td>
<td>2</td>
<td>2</td>
<td>Infancy</td>
</tr>
<tr>
<td>Seizure type at onset</td>
<td>Neck anteflexion, rolling of the eyes and elevating upper limbs</td>
<td>Epileptic spasms</td>
<td>Epileptic spasms</td>
<td>Epileptic spasms</td>
<td>Epileptic spasms</td>
<td>Epileptic spasms</td>
<td>Atonic type with head dropping</td>
<td>Nodded, slightly ocular myoclonus</td>
<td>Epileptic spasms</td>
<td>N/A</td>
</tr>
<tr>
<td>Other seizure types/course of seizures</td>
<td>Atonic and tonic seizures, myoclonic seizure</td>
<td>Atypical absences, tonic, focal dyscognitive seizures, obtundation status</td>
<td>Atonic and tonic seizures</td>
<td>Myoclonic, atypical absences, focal dyscognitive seizures, generalized tonic clonic seizures</td>
<td>Absences</td>
<td>N/A</td>
<td>Seizure-free since age 3.5 years</td>
<td>N/A</td>
<td>N/A</td>
<td>Age 5, one episode of unresponsiveness and eye deviation to the left side, characterized by some back arching and leaning over to the left side</td>
</tr>
<tr>
<td>EEG at onset of seizures</td>
<td>Partial hypsarrhythmia</td>
<td>Hypsarrhythmia</td>
<td>Sharp wave, slow wave</td>
<td>Slow background, multifocal discharges</td>
<td>Hypsarrhythmia</td>
<td>Modified hypsarrhythmia</td>
<td>Multiple epileptic activity with disorganization of basal activity</td>
<td>High voltage bilateral slow spike-wave discharges</td>
<td>Normal</td>
<td>Normal</td>
</tr>
<tr>
<td>Neurological Findings</td>
<td>N/A</td>
<td>Axial hypotonia</td>
<td>Axial hypotonia</td>
<td>Mild diffuse hypotonia, mild ataxia with wide based gait, mild tremor</td>
<td>General hypotonia</td>
<td>Axial hypotonia, secondary microcephaly</td>
<td>General hypotonia</td>
<td>Defomity, movement disorders</td>
<td>General hypotonia</td>
<td>Hypotonia</td>
</tr>
</tbody>
</table>

(Continues)
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Brain Imaging/MRI</td>
<td>Normal with generalized cerebral atrophy</td>
<td>Left ventricle slightly widened</td>
<td>Normal</td>
<td>Normal</td>
<td>Arachnoid cyst in right temporal lobe</td>
<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
<td>Proband 1 Proband 2</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Response to treatment/antiepileptic drugs</td>
<td>Intractable; seizure free from 3 to 5 years old with VPA and CLB</td>
<td>Intractable; seizure free from 4 years old to 5 years old with VPA</td>
<td>Intractable; seizure free from 3 to 8 years old with VPA</td>
<td>Intractable; some response to ketogenic diet</td>
<td>Intractable; seizure free on ketogenic diet</td>
<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
<td></td>
</tr>
<tr>
<td>Intellectual Development</td>
<td>Severe</td>
<td>Profound</td>
<td>Severe</td>
<td>Severe</td>
<td>Severe</td>
<td>Profound</td>
<td>Severe</td>
<td>Severe</td>
<td>Severe</td>
<td>N/A</td>
</tr>
<tr>
<td></td>
<td>Development delay</td>
<td>Normal</td>
<td>Profound</td>
<td>Severe</td>
<td>Severe</td>
<td>Profound</td>
<td>Severe</td>
<td>Severe</td>
<td>Severe</td>
<td>N/A</td>
</tr>
</tbody>
</table>
| CLB, clobazam; DD, developmental delay; F, female; LGS, Lennox Gastaut syndrome; M, male; N/A, not available or applicable; VGB, vigabatrin; VPA, valproic acid.
study have the first reported PH likely pathogenic variant. Our report indicates this variant in the PH domain causes less functional impairments and leads to a less severe phenotype than variants in the GTPase or middle domains.

Dynamin is a family of proteins with GTPase activity that facilitate vesicle scission (Achiriloaie, Barylko, & Albanesi, 1999). In vitro studies showed that the expression of certain GTPase mutants decreased endocytosis activity by interfering with vesicle scission during synaptic vesicle endocytosis and a middle domain mutation disrupted higher order DNM1 oligomerization (Dhindsa et al., 2015). Experiments also showed that when residue 44 in the GTPase domain of \( DNM1 \) was mutated, the protein had reduced binding affinity for GTP and reduced fission activity (Lee, Frank, Marks, & Lemmon, 1999).

The PH domain is primarily responsible for binding PIP2, yet the PH loop has been suggested to have constrictive functioning in the oligomer helix (Achiriloaie, Barylko, & Albanesi, 1999). This structure tends to insert into the plasma membrane and induce curvature before fission. In vitro dominant negative inhibition variants of the PH domain abolish phosphoinositide binding and knock out all synaptic vesicle reuptake (Lee et al., 1999). Variants in the PH domain of dynamin 1 have been previously studied by Vallis et al. (1999) by creating a Lys535Ala mutant. The position Lys535 was chosen because it comes into proximity to the 5’phosphate of the inositol ring in inositol-1,4,5-trisphosphate (IP3). The point mutation described in that article is at the same amino acid position as both patients in our study. While Patients 1 and 2 have a Lys535Glu variant, the mutated residue in that study is a Lys535Ala substitution. Each of these amino acid substitutions are significant because they do not share the same structure or chemical properties as lysine. Multiple PH domain mutants were compared, and while others did not show changed levels of transferrin uptake, a well-established endocytosis assay, Lys535Ala showed inhibition of endocytosis and impairment of PIP2 binding (Vallis et al., 1999). These findings support the critical role of Lys535 in the coordination of phosphoinositide binding, an essential step in endocytosis (Vallis et al., 1999).

A similar study by Achiriloaie et al. (1999) compared PH domain variants K535M and K561M to a GTPase domain variant N272 (missing most of the GTPase domain) and wild-type \( DNM1 \). The GTPase domain variant and K535M both inhibited endocytosis, while the K561 mutant and wild type \( DNM1 \) showed no significant difference in endocytosis (Achiriloaie, Barylko, & Albanesi, 1999). This implies that not all amino acid substitutions in the PH domain are significant and reinforces the claim that Lys535 is essential to the PH loop binding to PIP2 and promoting endocytosis. PH domain mutations at other sites may not have a clinically discernable phenotype. We also examined whether the position of the pathogenic variants affects different isoforms of Dynamin 1, but found all variants involve all isoforms (Figure 2b).
In summary, Dynamin 1 is mainly expressed in the central nervous system, explaining the neurological phenotype in DNM1-related disorder. Previously reported patients carry pathogenic variants in the GTPase or middle domains and presented with epileptic encephalopathy and severe neurodevelopmental symptoms. Here, we report the first documented DNM1 variant in the PH domain associated with a milder phenotype without epilepsy. Our data suggest a novel, possibly unique, and less severe phenotype resulting from a variant in the PH domain of dynamin 1. Reports of additional patients are needed to better define the genotype–phenotype correlation.

ACKNOWLEDGMENTS
We thank the family of the patients for participating in this study and the Exome Aggregation Consortium (http://exac.broadinstitute.org/about).

CONFLICTS OF INTEREST
AT discloses her employment with GeneDx. EB, EF, GCA, JD, SJP, and MS declare no conflicts of interest.

ORCID
Marwan Shinawi http://orcid.org/0000-0003-1329-4100

REFERENCES

SUPPORTING INFORMATION
Additional Supporting Information may be found online in the supporting information tab for this article.

How to cite this article: Brereton E, Fassi E, Araujo GC, et al. Mutations in the PH Domain of DNM1 are associated with a nonepileptic phenotype characterized by developmental delay and neurobehavioral abnormalities. Mol Genet Genomic Med. 2018;6:294–300. https://doi.org/10.1002/mgg3.362