2019

Association between the HLA-DRB1*03:01-DQB1*02:01 haplotype and PF4/heparin antibodies

Ray Zhang
Brian F. Duffy
Vinzenz Lange
Charles S. Eby
Chang Liu

Follow this and additional works at: https://digitalcommons.wustl.edu/open_access_pubs
Association between the HLA-DRB1*03:01-DQB1*02:01 haplotype and PF4/heparin antibodies

Ray Zhang,1 Brian F. Duffy,2 Vinzenz Lange,3 Charles S. Eby,1,4 and Chang Liu1

1Division of Laboratory and Genomic Medicine, Department of Pathology and Immunology, School of Medicine, Washington University in St. Louis, St. Louis, MO; 2HLA Laboratory, Barnes-Jewish Hospital, St. Louis, MO; 3DKMS Life Science Laboratory GmbH, Dresden, Germany; and 4Division of Hematology, Department of Medicine, School of Medicine, Washington University in St. Louis, St. Louis, MO

Key Points

• HLA-DRB1*03:01-DQB1*02:01 haplotype patients develop higher anti-platelet factor 4/heparin (PF4/H) levels following IV heparin exposure.
• HLA-restricted antigen presentation may play a role in the formation of anti-PF4/H antibodies.

Introduction

Heparin-induced thrombocytopenia (HIT) is a life-threatening complication of heparin exposure with estimated incidences of 1 in 5000 in general inpatients and 1% to 2% in inpatients following cardiac surgery.1,2 Antibodies against platelet factor 4 (PF4)/heparin (PF4/H) complexes trigger platelet activation, leading to consumptive thrombocytopenia and acquired thrombophilia capable of precipitating severe venous and arterial thrombosis.3 Enzyme-linked immunosorbent assays (ELISAs) are commonly performed to measure anti-PF4/H antibodies in patients with suspected HIT. High optical density (OD) values by ELISA correlate with increased platelet activation in serotonin release assays,4,5 and with the presence of clinical HIT.6-9

The mechanism for developing an anti-PF4/H response is poorly understood, as neither heparin nor PF4 is a foreign antigen in humans. Exogenous heparin may alter intracellular PF4 processing within antigen-presenting cells, causing presentation of aberrant PF4-derived peptides (neoantigens) to T cells.10,11 Although T cells are necessary for the development of anti-PF4/H in murine models,12 proof for aberrant antigen presentation is lacking.

Human PF4 neoantigen presentation may be restricted to specific HLA molecules. We hypothesized that patients having certain class II HLA alleles are predisposed to increased anti-PF4/H antibodies after exposure to heparin. Here, we present a cohort study of 412 heparin-exposed inpatients with paired anti-PF4/H and high-resolution HLA-typing results. We found that high anti-PF4/H antibody levels were associated with a specific class II HLA haplotype widely implicated in autoimmunity.

Methods

Study design

We conducted a single-center cohort study of inpatients who received IV unfractionated heparin for anticoagulation and were subsequently tested for anti-PF4/H due to concerns for HIT (Figure 1). Eligible patients were identified and their demographic information collected from the electronic medical record system. Anti-PF4/H in peripheral blood was measured using a commercial, poly-specific enzyme immunoassay (Asserachrom HPIA-ELISA; Diagnostic Stago). DNA was extracted from residual blood specimens and sequenced (Illumina) for high-resolution 2-field typing of the HLA-DRB1 and -DQB1 loci.13,14 Study approval was obtained from the Human Research Protection Office of Washington University in St. Louis.

Data analysis

HLA genotypes were examined at both the allele and the amino acid residue levels. Alleles exceeding 5% frequency in our study population were analyzed. Significance for differences in OD values between groups was evaluated using the Welch 2-sided unequal variances Student t test. Significance for associations between OD values (categorized as either high or low based on a moving OD cutoff) and

Submitted 16 April 2019; accepted 13 September 2019. DOI 10.1182/bloodadvances.2019000311. © 2019 by The American Society of Hematology
Adult inpatients who
1. Received intravenous unfractionated heparin for anticoagulation
AND
2. Were tested for anti-PF4/H antibodies
(n=437)

Excluded due to lack of specimen
(n=17)

Residual blood specimen retrieved
(n=420)

Excluded due to failed HLA typing
(n=8)

Clinical data
1. Demographics
   Age
   Gender
   Race
2. Laboratory
   Anti-PF4/H OD

HLA typing
1. High-resolution sequencing
   HLA-DRB1
   HLA-DQB1
(n=412)

Analysis
1. Differences in OD distribution (Welch t-test)
   HLA allele absent vs present (Fig. 2A top row)
   HLA amino acid absent vs present (Fig. 2B top row)
2. Association (Chi-squared test)
   HLA allele vs OD (Fig. 2A bottom row)
   HLA amino acid vs OD (Fig. 2B bottom row)

HLA protein visualization (PyMOL)

Figure 1.
either allele or amino acid residue (categorized as present or absent) was evaluated using the \( \chi^2 \) test of independence. The false discovery rate was controlled at 5\% using the Benjamini-Hochberg procedure.

**Results and discussion**

**Patients**

Between September 2016 and November 2017, 437 patients were evaluated, of whom 420 with blood specimens available were enrolled. The final analysis included 412 patients (176 females [42.7\%]) who were successfully typed at both HLA loci (Figure 1A). Our study population contained 337 white, 73 black, and 2 Asian patients. Ages ranged from 21 to 87 years (median, 62 years; interquartile range, 50-69 years). Antibody OD values ranged from 0.01 to 3.00 (median, 0.20; interquartile range, 0.09-0.53). There was no significant difference in OD values between patients based on sex, race, or age. Altogether, 43 DRB1 and 18 DQB1 alleles exceeded 5\% frequency in our study population (Figure 1B). Genetic linkage was evident between several DRB1-DQB1 allele pairs (Figure 1C).

**Anti-PF4/H vs HLA allele**

Patients with the DRB1*03:01 and/or DQB1*02:01 allele(s) had higher OD values than those without (\( P = .004-.007 \); Figure 2A top row), even after controlling for sex, race, and age. Using a moving OD cutoff to classify anti-PF4/H levels as either high or low, the presence of these alleles was associated with high anti-PF4/H over a broad range of OD value cutoffs (Figure 2A middle and bottom rows). Notably, patients possessing both DRB1*03:01 and DQB1*02:01 had 4 times the odds of other patients of having an OD value exceeding 1.5. Patients having both alleles constituted 44.7\% of all patients whose OD values exceeded 1.5 (18 of 38); the baseline frequency was 22.3\% (92 of 412).

**Anti-PF4/H vs HLA amino acid residue**

The top amino acid residue candidates for driving increased anti-PF4/H were R74, N77, and Y26, all within the peptide-binding groove of DRB1 (Figure 2B-C). Patients having these residues had higher OD values than those who did not (\( P = .015-.027 \); Figure 2B top row). These patients had 3 times the odds of other patients of having an OD value exceeding 1.5 (Figure 2B middle and bottom rows). Patients having all 3 residues constituted 47.4\% of all patients whose OD values exceeded 1.5 (18 of 38); the baseline frequency was 26.4\% (109 of 412).

**Discussion**

We investigated immunogenetic risk for elevated anti-PF4/H, which ultimately causes clinical HIT in a subset of patients, by the hypothesis that HLA-restricted aberrant antigen presentation precedes production of anti-PF4/H. Our end point was ELISA anti-PF4/H level. Other end points such as serotonin release assay or clinical HIT with thrombosis, despite their importance, require additional elements (eg, platelet-related factors) to manifest. Due to a lack of uniformly accepted OD value cutoff for positivity,\(^5\) we characterized the complete OD value distribution and examined all possible cutoffs. We found that DRB1*03:01 and DQB1*02:01 were significant risk factors for increased anti-PF4/H antibodies following heparin exposure among inpatients.

Earlier studies generated inconsistent findings of association between HLA and anti-PF4/H.\(^6\)-\(^8\) These studies were limited by the typing methodology at the time, providing low-resolution\(^6\),\(^7\) or inferred\(^8\) typing only. One study also used healthy individuals without known heparin exposure as controls.\(^6\) Our results corroborate the finding of DR3-HIT antibody association from Paparella et al,\(^7\) extending their conclusions from cardiovascular surgical patients to general inpatients. Notably, our study...
used high-resolution HLA typing to enable prediction of risk-carrying residues within the peptide-binding groove of DRB1, providing a basis for future investigations into which peptides may cause disease.

The DRB1*03:01-DQB1*02:01 haplotype has been associated with multiple autoimmune disorders, including celiac disease, type 1 diabetes, and systemic lupus erythematosus. HIT may share similar pathogenesis with these disorders via aberrant antigen presentation by DRB1*03:01 and/or DQB1*02:01. One of the DRB1 amino acid residues associated with increased anti-PF4/H has been implicated in type 1 diabetes and autoimmune thyroid disease. In patients with celiac disease, altered gut microbiota or defective intestinal barriers may also prime an anti-PF4/H response given the observed connection between bacteria and HIT antibodies. Presence of a shared susceptibility to these autoimmune disorders and elevated anti-PF4/H antibodies could signal an increased pretest probability for HIT in thrombocytopenic heparin-exposed patients having a known history of autoimmunity.

Our study is limited by sampling from a single institution without a replication cohort. Dosages, durations, and indications for heparin, as well as patient histories including the presence of autoimmune disease, could not be reliably ascertained for the study. We also examined only DRB1 and DQB1 in order to balance investigating the most functionally relevant class II loci with minimizing redundant or multiple testing (for example, DRB3/4/5 were omitted due to well-established tight genetic linkage with DRB1-DQB1), and therefore cannot rule out the contribution of other HLA loci or other genetically linked genes in the HLA region that may affect anti-PF4/H production. Finally, an immunoglobulin G–specific ELISA may have been better suited for this study compared with the poly-specific assay available at our institution, although high values from either assay correlate well with HIT. Future multidisciplinary studies are warranted.

Figure 2. Association between HLA and anti-PF4/H antibody levels. (A) HLA allele associations: top row, distribution of OD values; middle row, odds ratio for having an OD value above vs below a given cutoff; and bottom row, significance of association across all possible cutoffs (95% confidence intervals in pink). (B) HLA amino acid residue associations: top row, distribution of OD values; middle row, odds ratio for having an OD value above vs below a given cutoff; and bottom row, significance of association across all possible cutoffs (95% confidence intervals in pink). (C) Two views of candidate amino acid residues within DRB1 driving risk for anti-PF4/H antibody development (Protein Data Bank code: 1A6A). *Significance within a 5% false discovery rate.
Figure 2. (Continued).
to further understand the functional relationship between HLA and anti-PF4/H antibodies.

Acknowledgments

The authors thank DKMS Life Science Laboratory GmbH for performing Illumina-based HLA typing, and Rhonda Porche-Sorbet for technical assistance.

This work was partly supported by the Washington University in St. Louis Hematology Scholars K12 award (K12-HL087107-07 from the National Institutes of Health, National Heart, Lung, and Blood Institute) (C.L.).

Authorship

Contribution: R.Z. analyzed data and composed the manuscript; B.F.D. handled specimens and extracted DNA; V.L. performed HLA typing; C.S.E. designed the study and composed the manuscript; and C.L. designed the study, procured specimens, analyzed data, and composed the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: R.Z., 0000-0001-9668-3659; V.L., 0000-0002-6442-9573.

Correspondence: Charles S. Eby, Division of Laboratory and Genomic Medicine, Department of Pathology and Immunology, and Division of Hematology, Department of Medicine, School of Medicine, Washington University in St. Louis, 660 South Euclid Ave, Campus Box 8118, St. Louis, MO 63110; e-mail: eby@wustl.edu; and Chang Liu, Division of Laboratory and Genomic Medicine, Department of Pathology and Immunology, School of Medicine, Washington University in St. Louis, 660 South Euclid Ave, Campus Box 8118, St. Louis, MO 63110; e-mail: cliu32@wustl.edu.

References


