Neutrophils promote VLA-4–dependent B cell antigen presentation and accumulation within the meninges during neuroinflammation

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The success of B cell depletion therapies and identification of leptomeningeal ectopic lymphoid tissue (ELT) in patients with multiple sclerosis (MS) has renewed interest in the antibody-independent pathogenic functions of B cells during neuroinflammation. The timing and location of B cell antigen presentation during MS and its animal model experimental autoimmune encephalomyelitis (EAE) remain undefined. Using a new EAE system that incorporates temporal regulation of MHCI expression by myelin-specific B cells, we observed the rapid formation of large B cell clusters in the spinal cord subarachnoid space. Neutrophils preceded the accumulation of meningeal B cell clusters, and inhibition of CXCR2-mediated granulocyte trafficking to the central nervous system reduced pathogenic B cell clusters and disease severity. Further, B cell-restricted very late antigen-4 (VLA-4) deficiency abrogated EAE dependent on B cell antigen presentation. Together, our findings demonstrate that neutrophils coordinate VLA-4–dependent B cell accumulation within the meninges during neuroinflammation, a key early step in the formation of ELT observed in MS.

**Significance**

A distinct murine model of multiple sclerosis used to examine factors involved in ectopic lymphoid tissue formation during central nervous system autoimmunity reveals that infiltration and aggregation of B cells within the leptomeninges is dependent upon B cell expression of VLA-4 and is preceded by neutrophil migration. This finding establishes the early mechanistic involvement in the establishment of chronic inflammatory changes within the meninges during EAE and is preceded by neutrophil migration. This finding establishes the early mechanistic involvement in the establishment of chronic inflammatory changes within the meninges during autoimmune inflammation that promote the formation of ectopic lymphoid tissue associated with disease progression and disability in multiple sclerosis.


The authors declare no competing interest.

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fickling and retention, critical for ELT organization within the specialized immune target of the CNS, have yet to be explored.

To examine the unique contributions of B cell antigen-presenting cell (APC) function to CD4 T cell encephalitogenicity, we developed an EAE model allowing for timed onset of disease mediated solely by B cell antigen presentation (6). In conjunction with rapid disease onset, we now describe the formation of large clusters of B cells within the spinal cord subarachnoid space resembling immature ELT. The accumulation of B cells in the spinal cord meninges was preceded by an increase in CD45hi, CD11bhi, Ly6C−, Gr-1− myeloid cells. These inflammatory myeloid cells promoted B cell trafficking to the CNS compartment and influenced the formation of pathogenic B cell clusters. Additionally, B cell expression of VLA-4 was necessary for B cell accumulation and the development of passive EAE. These data suggest that neutrophils enhance recruitment or retention of B cells in an anatomic compartment that facilitates B cell access to both antigens and autoreactive T cells.

Results

Meningeal Space Is a Niche for B Cells during EAE. To assess the location where B cells may participate as APCs within the CNS during EAE, we used flow cytometry to assess the frequency of B cells in various CNS tissues. Following immunization of B6 mice with MOG35-55, B cells were found in low frequency and detected primarily in the meninges rather than the CNS tissue parenchyma (Fig. 1A). A similar frequency and distribution of B cells was observed in the B cell-dependent EAE model involving active immunization of B6 mice with hMOG (Fig. 1B), which is unlike an initial assessment of B cell numbers in these models (19). In contrast, passive transfer of MOG-specific encephalitogenic CD4 T cells into B6 mice or mice with selective major histocompatibility complex class II (MHCII) expression by B cells with an elevated precursor frequency of MOG-specific B cells led to a significantly greater frequency of B cells in the CNS. In these passive EAE models, B cells were found in the spinal cord meninges in preponderance to the CNS tissue parenchyma (Fig. 1C and D). These results indicate that the meningeal compartment is the primary location where B cells collect during EAE and suggest that, given the proximity to antigenic debris and cognate CD4 T cells, B cells could serve as potent APCs within the meninges.

We previously reported a novel EAE model, in which MHCII expression is induced in a cell-specific as well as temporal manner. B cells are capable of serving all antigen presentation functions during passive EAE as long as the repertoire of B cells is maintained.

![Diagram](https://example.com/diagram.png)

**Fig. 1.** The meningeal space is a niche for B cells during EAE. The mean ± SEM of B cells in different CNS tissues analyzed by flow cytometry. WT mice immunized with (A) MOG35-55 (n = 7, pooled from 2 different experiments) and (B) hMOG harvested 3 d post active EAE (n = 6, pooled from 2 different experiments). (C) WT mice (n = 15, pooled from 7 different experiments) and (D) CD19-BMHCIIxIgHMOG mice (n = 12, pooled from 4 experiments) harvested 3 d post passive EAE onset. (E) Tam-WT APC (n = 12, pooled from 5 different experiments) and (F) Tam-BAPC mice (n = 13 mice, pooled from 6 different experiments) treated with Tam 3 wk after CD4 T cell transfer and harvested 3 d post EAE onset. (G) Disease severity for samples from A–F at 3 d postonset. Br, Brain; Br-M, Brain meninges; Sc, Spinal cord; Sc-M, Spinal cord meninges. (H) B220 (pink), CD3 (green), and DAPI (blue) immunofluorescence staining of spinal cords carefully excised from recipients of encephalitogenic CD4 T cells. (Left) Spinal cord from Tam-BAPC mouse harvested 3 wk after T cell transfer without Tam treatment. Spinal cords harvested 3 d post EAE onset from Tam-WT APC (Middle) and Tam-BAPC (Right) mice treated with Tam 3 wk after CD4 T cell transfer. Images are taken at 4× magnification and are representative of n = 7 to 8 mice per genotype, pooled from 4 separate experiments. (Scale bars, 100 μm.)
is narrowed on the T cell cognate antigen (MOG) resulting from IgHMOG expression (20). Using this system to examine the critical timing of cognate interactions during neuroinflammation, the onset of disease between Tam-WTAPC and Tam-BAPC mice can be synchronized by inducing MHCII expression upon oral gavage with Tam after encephalitogenic CD4 T cell transfer (20). We showed that Tam-WTAPC mice develop passive EAE similar to WT mice, with or without IgHMOG transgene expression, while Tam-BAPC mice can exhibit accelerated disease onset dependent on the timing of MHCII expression relative to encephalitogenic T cell transfer (20). As with other models of EAE, flow cytometric assessment of Tam-WTAPC and Tam-BAPC mice with EAE revealed infiltration of B cells predominantly within the spinal cord meninges (Fig. 1E and F). No significant difference in disease severity was observed between different models at day 3 postonset (Fig. 1G). Examination of spinal cord tissue for the presence of B cells in Tam-WTAPC and Tam-BAPC mice failed to identify evidence of inflammation prior to Tam treatment of mice that had received encephalitogenic CD4 T cells 3 wk prior to harvest. However, B220+ B cells and CD3+ T cells were evident in the spinal cords of Tam-WTAPC mice and Tam-BAPC mice harvested 3 d post EAE onset, indicating that MHCII+ expression is associated with the induction of lymphocyte trafficking to the CNS (Fig. 1E). Interestingly, B cells appeared organized in dense clusters within the meninges in the spinal cord sections from Tam-BAPC mice, and this pattern was less common in Tam-WTAPC mice (Fig. 1E, arrowheads).

B Cell Clusters with Features of Rudimentary Ectopic Lymphoid Follicles Form in the Spinal Cord of Mice with B Cell-Dependent EAE. To investigate differences in meningeal B cell clusters in Tam-BAPC and Tam-WTAPC mice, we quantified the volume and number of B cell clusters. Spinal cords left intact within the spinal column were harvested from mice 3 d after EAE onset, optically cleared, and stained for B cells. Based on this more exhaustive imaging method, B cell staining within the parenchyma of the spinal cord again was rare in both genotypes. Within the meninges, however, small clusters of B cells were observed proximal to the spinal cord of Tam-WTAPC mice by confocal imaging (Fig. 2A). In contrast, Tam-BAPC mice displayed extensive clusters of B220+ B cells that extended several millimeters in length (Fig. 2B). Qualitatively, B cell clusters in the meninges of Tam-BAPC mice appeared denser than the diffuse B cell infiltration seen in Tam-WTAPC mice (Movie S1). Quantifying the area and number of distinct areas of B cell staining in cleared spinal cord specimens, a statistically significant difference in mean voxel area per cluster between the spinal cords of Tam-WTAPC (6857 ± 1236) and Tam-BAPC mice (9675 ± 757) was observed (P = 0.05). Our findings suggest that this model of EAE offers an opportunity to explore the earliest steps involved in B cell organization leading to ELT formation.

To gain insight into the resemblance of B cell clusters in the spinal cord meninges to ELT, we first isolated the spinal cord meninges of Tam-BAPC mice at 3 d post EAE onset and used flow cytometry to analyze the composition of leukocytes. Splenocytes harvested from an hMOG-immunized mouse were used to develop a gating strategy to detect the expression of GC markers and determine B cell populations present in the meningeal clusters (Fig. 3A). Based on expression of IgM and IgD we found that the majority of B cells in the spinal cord meninges included IgM+ IgD− B cells and IgM− IgD− naive follicular B cells (Fig. 3B). While relatively few B cells existed in the CNS parenchyma, most expressed MHCII. In the spinal cord meninges, where B cells are plentiful, nearly all were MHCII+, indicating a capacity for cognate interactions with CD4 T cells (Fig. 3C). CD138+ plasmablasts and plasma cells were very rare in all tissues analyzed but were most frequent in the brain compared to the spinal cord and spinal cord meninges (Fig. 3C). The mean frequency of GL7+ GC B cells in the spinal cord meninges was low, although elevated compared to brain and spinal cord parenchyma, indicating that most B cells in the CNS were not undergoing GC-like reactions at this early stage of the disease (Fig. 3C). Spinal cord specimens stained with both B220 and the proliferation marker Ki-67 revealed the absence of co-staining (Fig. 3D), demonstrating the formation of B cell clusters just 3 d following the onset of EAE is likely due to a large infiltration of B cells rather than proliferation in situ. Some B cell clusters in the spinal cord meninges were heavily infiltrated by CD3+ T cells (Fig. 3E, i, arrows), while T cells were sparse in other clusters (Fig. 3E, ii, arrows). In summary, the spinal cord meninges of Tam-BAPC mice with EAE rapidly developed dense clusters of activated, class-switched, MHCII+ B cells, suggesting that this is a critical location for cognate interactions during EAE.

To define the specific anatomical site of B cell infiltration within the meninges during EAE, we analyzed decalcified spinal cords harvested from Tam-BAPC mice 3 d after EAE onset using immunofluorescent staining. We detected B cell extravasation from the cerebrovasculature which formed clusters surrounding vascular cell adhesion molecule-1 (VCAM-1)+ activated endothelial cells (Fig. 4A and B). B cell clusters were excluded from the spinal cord parenchyma, identified by glial fibrillary acidic protein (GFAP) staining (Fig. 4B), and pia mater, identified by laminin (Fig. 4C). Rather, B cell clusters were associated with the arachnoid mater as seen by Podoplanin-1 staining (Fig. 4B and C). Overlays of these immunofluorescent stains revealed that B cell clusters were specifically in the subarachnoid space, an anatomical compartment with access to both antigens and cognate CD4+ T cells.

Inflammatory Myeloid Cells Facilitate B Cell Cluster Formation in the Subarachnoid Space. The mechanisms of B cell localization to the meninges remain unclear. To understand the kinetics of immune cell trafficking and B cell cluster formation, the spinal cord meninges of Tam-WTAPC and Tam-BAPC mice harvested at various time points throughout the course of EAE were examined by flow cytometry. The baseline composition of immune cells was measured in Tam-WTAPC and Tam-BAPC animals harvested 3 wk after receiving encephalitogenic CD4+ T cells but prior to Tam
Fig. 3. B cell clusters in the spinal cord meninges exhibit features of rudimentary ectopic lymphoid follicles after EAE onset. (A) Example flow cytometry gating strategy generated from hMOG-immunized WT splenocytes harvested 3 d post active EAE onset to detect and quantify the data shown in B and C. Histogram of B cell MHCII staining is overlaid with shaded histogram of CD11b+ cells. (B) Frequency of IgM− IgD+ B cells, IgM+ IgD+, IgM− IgD−, and IgM+ IgD− (naive mature follicular B cells) B cells, as a percent of total B cells in the spinal cord meninges of Tam-BAPC mice treated with Tam at week 3 post CD4 T cell transfer and harvested around 3 d post EAE onset. Data generated from n = 11 mice, pooled from 4 separate experiments. (C) Mean frequency with 95% confidence interval of MHCII+ B cells (Left) as a percent of total B cells and of CD138+ B220+ plasma cells (Middle) and IgD− GL7+ GC B cells (Right), out of the total frequency of CNS mononuclear cell singlets in the brain (squares), spinal cord (triangles), and spinal cord meninges (circles) of Tam-BAPC mice treated with Tam at week 3 post CD4 T cell transfer and harvested 3 d post EAE onset. Data generated from n = 11 mice, pooled from 4 separate experiments. (D) B220 (pink), Ki-67 (green), and DAPI (blue) immunofluorescence staining of decalcified spinal cord in the vertebral of Tam-BAPC mice treated with Tam at week 3 post CD4 T cell transfer and harvested 3 d post EAE onset. Images are representative of n = 7 mice, pooled from at least 4 separate experiments. (Scale bars, 100 μm.) (E) B220 (red/ pink), CD3 (green), and DAPI (blue) immunofluorescence staining of decalcified spinal cord in the vertebral of Tam-BAPC mice treated with Tam at week 3 post CD4 T cell transfer and harvested 3 d post EAE onset. (i and ii, Insets) Single-color images from boxed regions of interest imaged at 10x magnification. Arrows indicate B and T cell colocalization. Images are representative of n = 7 mice, pooled from at least 4 separate experiments. (Scale bars, 100 μm.)
Fig. 4. B cell clusters form in the subarachnoid space adjacent to activated endothelial vasculature. Tam-BAPC mice were treated with Tam at week 3 post CD4 T cell transfer and killed 3 d post EAE onset. (A, Left) Decalcified spinal cords were stained for B220 (pink), CD3 (green), and DAPI (blue) and imaged at 4× magnification. (A, Right) Boxed insets are regions of interest (from left) stained for activated endothelial cells (VCAM-1, red), astrocytes (GFAP, green), and DAPI (blue) and imaged at 10× magnification. (B, Left, Top, and Bottom) Overlays of single-color images displayed at Right. (i) GFAP (green), (ii) VCAM-1 (red), (iii) DAPI (blue), and (iv) podoplanin-1 (white). (C) Serial sections of the spinal cord shown in B. (Left, Top, and Bottom) Overlay images of single-color images displayed at Right. (i) Laminin (green), (ii) B220 (red), (iii) DAPI (blue), and (iv) podoplanin-1 (white). All images are representative of n = 7 mice, pooled from at least 4 separate experiments. (Scale bars, 100 μm.)

Treatment. At 24 and 72 h after induction of MHCII expression, B cell counts were low in the spinal cord meninges of Tam-WTAPC mice (Fig. 5A) and Tam-BAPC mice (Fig. 5B). However, by 3 d post EAE onset, B cells were the most numerous cell type analyzed (Fig. 5 A and B). Donor T cells were virtually undetectable in the spinal cord meninges until the onset of EAE symptoms for Tam-WTAPC mice (Fig. 5A) and Tam-BAPC mice (Fig. 5B). Strikingly, we observed early and sustained infiltration of CD45hiCD11bhiGr-1hi inflammatory cells in Tam-BAPC mice but not Tam-WTAPC mice, a difference that was statistically significant at 24 h post Tam, 72 h post Tam, and 3 d post EAE onset (Fig. 5C). We reasoned that these CD45hiCD11bhiGr-1hi inflammatory cells were most likely polymorphonuclear neutrophils (PMNs), as this cell type is rapidly recruited to sites of inflammation and infection and has been detected during initial phases of EAE (21, 22). Imaging of decalcified spinal cords from Tam-BAPC mice with EAE revealed the proximity of B cell clusters with Ly6G+ PMNs (SI Appendix, Fig. S1), consistent with a coordinated influx of PMNs promoting an environment conducive to B cell antigen presentation and the rapid establishment of B cell clusters following EAE onset.

Immune Cell Trafficking to the CNS to Induce EAE Is Directed by Distinct Sequences of Chemokine Regulation in WT and B Cell–Dependent EAE. We reasoned that identification and quantification of the chemokines present in the CSF during the early stages of EAE induction would provide insight into the previously observed differences in the coordinated trafficking of immune cells to the CNS in B cell–dependent EAE. CSF was collected from naive WT mice and pre-Tam Tam-BAPC mice, as well as from Tam-WTAPC and Tam-BAPC mice at 24 and 72 h post Tam, the day of EAE onset, 3 d post EAE onset, and 30 d post Tam to determine the dynamic expression of inflammatory cytokines and chemokines.

The mean concentrations of analytes observed in naive WT mice were similar to the levels detected in CSF collected from Tam-BAPC mice harvested 3 wk post CD4 T cell transfer but prior to Tam treatment (Fig. 6 A–E), indicating that encephalitogenic CD4 T cells did not induce inflammatory responses in the absence of MHCII expression. Tam-WTAPC (red squares) exhibited higher concentration of IFNγ at 24 h post Tam compared to Tam-BAPC mice (P = 0.006). At the day of EAE onset, the concentration in Tam-WTAPC mice was still significantly up-regulated compared to Tam-BAPC mice (P < 0.0001) (Fig. 6A). Other mediators of inflammation, including CC chemokine ligand (CCL)5 and CCL19 within the CSF CCL5 were significantly different between Tam-WTAPC and Tam-BAPC mice (Fig. 6 B and C). Drastically different concentrations of the CX3C glutamate-2-leucine-arginine (ELR)+ chemokines CXCL5, CXCL2, and CXCL1 were identified in the CSF of Tam-WTAPC and Tam-BAPC mice at the same stages of disease. Tam-WTAPC mice tended to rapidly develop much higher concentrations of these chemokines than Tam-BAPC mice (Fig. 6D).

Given the significance of CXCL13 for B cell trafficking and GC reactions, differences in the concentration of this chemokine between Tam-WTAPC and Tam-BAPC mice could provide insight into the progression of B cell cluster formation. Although a substantial increase in concentration was observed in both groups, the only difference was identified, at 3 d post EAE onset, with Tam-WTAPC mice exhibiting significantly lower CXCL13 concentrations compared to Tam-BAPC mice (Fig. 6E).

The variability in chemokine concentrations in the CSF between Tam-WTAPC mice and Tam-BAPC mice, especially at 72 h post Tam, could explain our observation that these 2 genotypes differ in...
CD45b<sup>hi</sup>CD11b<sup>hi</sup>Gr-1<sup>hi</sup> inflammatory PMN invasion of the spinal cord meninges (Fig. 5C). These ELR<sup>+</sup> CXC chemokines all share the receptor CXC chemokine receptor (CXCR2), which is predominantly expressed by PMNs. In Tam-B<sup>APC</sup> mice 72 h post Tam, CXCR2 expression was detected in splenic CD45b<sup>hi</sup>CD11b<sup>hi</sup>Gr-1<sup>hi</sup> inflammatory PMN cells, but not on PMNs isolated from the meninges (Fig. 6F). However, 3 d post EAE onset, PMN expression of CXCR2 in the spinal cord meninges was notably higher (Fig. 6F). B cells did not express CXCR2 in any tissue at any time point examined (Fig. 6F). Further flow cytometric analysis of the phenotype of CD45b<sup>hi</sup>CD11b<sup>hi</sup>Gr-1<sup>hi</sup> inflammatory PMNs in the spinal cord meninges of Tam-B<sup>APC</sup> mice confirmed minimal contamination with Ly6C<sup>+</sup> monocytes (SI Appendix, Fig. S2). Between 72 h post Tam and 3 d post EAE onset, PMNs isolated from the spinal cord meninges exhibited signs of activation with a reduction in surface expression of CD62L coinciding with increased CXCR2 (SI Appendix, Fig. S2). These data are corroborated by reports that transmigration of the blood–brain barrier (BBB) induces enhanced inflammatory phenotypes in PMNs (23).

Overall, the data indicate that B cell-dependent EAE promotes a unique inflammatory signaling cascade resulting in the CXCR2-directed recruitment of PMNs at temporally regulated stages of disease.

**Early PMN Recruitment to the CNS Is Required for the Formation of Meningeal B Cell Clusters and Development of EAE.** Modulating CXCR2 signaling with anti-CXCR2 antibodies or small-molecule competitive antagonism has been shown to alter the site of neuroinflammation and the composition of inflammatory infiltrates during EAE and can even modulate the expression of disease symptoms to protect mice from atypical EAE induced by Th17 cells (24, 25). To test whether the early, coordinated recruitment of inflammatory myeloid cells precedes B cell infiltration in B cell-dependent EAE. (A–C) Box and whiskers plots showing mean and 10 to 90th percentile data points generated from the frequency (Left) and absolute number (Right) of B cells (maroon circles), Gr-1<sup>+</sup> PMNs (black squares), and donor CD4 T cells (green triangles) in the spinal cord meninges of Tam-B<sup>APC</sup> mice harvested at 3 wk post T cell transfer and before Tam treatment or from Cre-littermates of Tam-B<sup>APC</sup> mice harvested at 3 wk post T cell transfer. Data generated from n = 15 mice, pooled from at least 4 separate experiments. (A) Box and whiskers plots showing mean and min/max data points generated from the frequency (Left) and absolute number (Right) of B cells (maroon circles), Gr-1<sup>+</sup> PMNs (black squares), and donor CD4 T cells (green triangles) in the spinal cord meninges of Tam-B<sup>APC</sup> mice harvested at various time points in disease. Data are generated from n = 8 to 12 mice per time point, pooled from at least 10 separate experiments. (B) Box and whiskers plots showing mean and 10 to 90th percentile data points generated from the frequency (Left) and absolute number (Right) of B cells (maroon circles), Gr-1<sup>+</sup> PMNs (black squares), and donor CD4 T cells (green triangles) in the spinal cord meninges of Tam-B<sup>APC</sup> mice harvested at various time points in disease. Data generated from n = 11 to 16 mice, pooled from at least 10 different experiments. (C) Box and whiskers plots showing mean and all data points for frequency (Left) and absolute number (Right) of Gr-1<sup>+</sup> PMNs harvested from the spinal cord meninges of Tam-WT<sup>APC</sup> mice (red) and Tam-B<sup>APC</sup> mice (blue) killed at various time points in disease course. Statistical significance was determined by 2-tailed Mann–Whitney U tests. Exact P values calculated, *P < 0.05, **P < 0.01.

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of PMNs to the CNS was functionally relevant to EAE dependent on B cell antigen presentation, we neutralized CXCR2 in Tam-B^APC^ mice. Anti-CXCR2 serum was administered after encephalitogenic CD4 T cells were transferred into Tam-B^APC^ mice but prior to MHCII induction 3 wk later. The disease severity as represented by total area under the curve of mean disease scores was similar for mice receiving control serum and untreated control mice (Fig. 7A). Mice treated with anti-CXCR2 blocking antibody did not develop EAE as quickly or as severely as the control groups (Fig. 7B). Examination of immunofluorescence images indicates that B cell clusters were present in the spinal cord meninges of both control groups, but were not detected in mice treated with anti-CXCR2 serum (Fig. 7B), suggesting that meningeal B cell cluster formation is pathogenic for EAE dependent on B cell antigen presentation. By flow cytometry, donor T cells in the spinal cord meninges of anti-CXCR2 mice (Fig. 7C) and spinal cord meninges (Right) of Tam-B^APC^ mice collected at 72 h post Tam (Left two panels) and 3 d post EAE onset (Right two panels). Graphs are representative of n = 4 to 5 mice, pooled from 2 separate experiments.

**suggest that interrupting CXCR2 signaling by PMNs diminished the ability of B cells to access the CNS, a step that may be required for B cell APCs to support EAE.**

**B Cell Access to the Subarachnoid Space Is Necessary for B Cell Antigen Presentation to Support Passive EAE.** Based on the observation that EAE was ameliorated and B cell clusters were absent in anti-CXCR2–treated mice, we hypothesized that entry into the CNS compartment was required for B cell antigen presentation to induce neuroinflammation. To test this, CD19-B^MHCIIxIgHMOG^ xVLA-4f/f mice and treated them with Tam at 3 wk post-αencoding the 4 integrin of the VLA-4 molecule is flanked by LoxP (f/f) sites (VLA-4f/f mice). Protein expression changes were confirmed in peripheral blood B cells from Tam-B^APC^ mice prior to Tam treatment and 72 h post Tam treatment (Fig. 7C). WT and Cre^VLA-4f/f^ mice with WT VLA-4 expression levels were both susceptible to EAE (SI Appendix, Table S1) with a similar day of onset and disease severity (Fig. 7D, Top). CD19-B^MHCIIxIgHMOG^ and CD19-B^MHCIIxIgHMOG^ xVLA-4f/f^ mice were susceptible to EAE, but CD19-B^MHCIIxIgHMOG^ xVLA-4f/f^ mice were completely protected (Fig. 7D, Center). We repeated the passive EAE experiment with Tam-B^APC^, Tam-B^APC^ xVLA-4f/f^, and Tam-B^APC^ xVLA-4f/f^ mice and treated them with Tam at 3 wk post-encephalitogenic CD4 T cell transfer. As expected, mice with WT

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**Fig. 6.** Temporal regulation of chemokine expression mediates immune cell recruitment to the CNS. (A–F) Graphs show box and whiskers plots with min-to-max error bars of concentrations for (A) IFN-γ; (B) RANTES/CCL5; (C) MIP3-β/CCL19; (D) Left) CXCL5, (Middle) CXCL2, (Right) CXCL1; and (E) CXCL13 detected in the CSF of naive WT (black), Tam-WT^APC^ (red) and Tam-B^APC^ (blue) mice harvested at various time points in EAE disease course. Statistical significance was determined by 2-tailed Mann-Whitney U tests. Exact P values calculated, *P < 0.05, **P < 0.001, ***P < 0.0001, ****P < 0.0001. (F) Flow cytometric analysis of CXCR2 expression on PMNs (gray histogram) and B cells (blue histograms) isolated from the spleen (Left) and spinal cord meninges (Right) of Tam-B^APC^ mice collected at 72 h post Tam (Left two panels) and 3 d post EAE onset (Right two panels). Graphs are representative of n = 4 to 5 mice, pooled from 2 separate experiments.
and heterozygous expression of VLA-4 in B cells were susceptible to EAE with similar severity and day of onset, while mice devoid of B cell VLA-4 expression were completely protected (Fig. 7D, Bottom). Examination of spinal cords by immunofluorescence confirmed that Tam-B^{APC} mice and Tam-B^{APC}/VLA-4^{f/+} mice with EAE both developed substantial clusters of B cells in the spinal cord meninges, while the deletion of B cell VLA-4 expression prevented B cells from entering the subarachnoid space by extravasation through VCAM-1+ blood vessels (SI Appendix, Fig. S4). These results demonstrate the requirement for B cell VLA-4 expression for access to the CNS compartment and antigen presentation function during EAE.
Discussion

While cognate interactions between B cells and CD4 T cells are known to be important for autoantibody formation in MS, the nature and location of these pathogenic interactions in EAE are unclear. Similarly, the genesis of ELT within the meninges of patients with progressive MS is thought to be clinically relevant, but the mechanisms leading to the formation of these structures remain unknown. We sought to determine the sequence of proinflammatory events that fosters the formation of antigen-specific B and T cell interactions within the CNS intrinsic to the clinico-pathologic features of MS. We have found that the subarachnoid space of the spinal cord meninges serves as a niche for B cell cluster formation during EAE. This is corroborated by other reports of ELT in EAE, including Dang et al. (26), who found nonclassically activated B cells in the spinal cord meninges of TCR\textsuperscript{MOG}\textsuperscript{+}\textsuperscript{-}IgG\textsuperscript{MOG}\textsuperscript{-} mice with spontaneous EAE. In our model, we found a high frequency of MHCII\textsuperscript{+} B cells that frequently exhibited surface markers such as IgM\textsuperscript{-} IgD\textsuperscript{-} B cells (27) and naive follicular B cells. The low frequency of CD138\textsuperscript{+} plasma cells or plasmablasts and few GL7\textsuperscript{+} GC B cells we observed in the spinal cord meninges can be accounted for by the early time points used in this study. One caveat to these results is that CD138 staining can be challenging based on the lack of stability of this surface marker during processing for flow cytometry, leading to a potential underestimation of the number of plasma cells/plasmablasts. Also, it is important to note that the encephalitogenic donor CD4 T cell lines used to induce passive EAE in our model are very strongly Th1-skewed and thus distinct from models exhibiting ELT in B6 or SJL (Swiss Jim Lambert) mice involving IL-17-skewed CD4 T cells (12, 18). This is relevant, given that prior work has shown that Th17 cells are capable of trafficking to the CNS during EAE without expression of VLA-4 (28). Additionally, donor T cells driving disease in our model may not be capable of functioning as efficient T follicular helper cells to sustain GC reactions. These explanations are not mutually exclusive and deserve further exploration to elucidate the critical mechanisms involved in transitions from immature to established ELT structures in EAE.

To explore the factors recruiting and retaining B cells to the subarachnoid space to the exclusion of other CNS tissues we investigated the kinetics of innate and adaptive immune cell trafficking at several stages of EAE progression. Prior to EAE onset, CD45\textsuperscript{+}CD11b\textsuperscript{+}Gr-1\textsuperscript{hi} PMNs are the most common cell type observed in the spinal cord meninges of Tam-WT\textsuperscript{APC} and Tam-B\textsuperscript{APC} mice. This proinflammatory cell type is often a major component of early inflammatory infiltrates in EAE (22, 29, 30), functioning as efficient T follicular helper cells to sustain GC reactions. Formation of these rudimentary meningeal ELT in Tam-B\textsuperscript{APC} mice is essential for B cell antigen presentation to support neuroinflammation. Prior to the onset of EAE and well before B cell cluster formation, PMNs are recruited to the CNS by ELR\textsuperscript{+} CXC chemokines. Inhibition of CXCR2-mediated PMN trafficking to the CNS reduces pathogenic B cell clusters, suggesting that PMN recruitment primes the CNS compartment for B cell cluster formation. Therapeutic targeting of CXCR2 signaling could be effective for MS, and more research is needed to investigate the possibility of direct or indirect interactions between PMNs and B cells. In accord with successful therapies that eliminate B cells (BCDTs) or prevent immune cell trafficking to the CNS (Natalizumab or Fingolimod), B cell-restricted VLA-4 deficiency inhibits the development of EAE symptoms when B cells are the only APCs. Coordinated trafficking of innate and adaptive immune cells may promote EAE by supporting VLA-4-dependent B cell accumulation within the meninges during neuroinflammation. Future experiments aimed at determining whether B cell access to the CNS is necessary for antigen acquisition or presentation to CD4 T cells will be essential in refining cellular immune therapeutic targeting with distinct CNS compartments in mind.

Materials and Methods

Mice. Mice were housed in a single pathogen-free housing facility and fed mouse chow and water ad libitum. Male and female mice between 6 and 10 wk of age were used for all experiments with the exception of the anti-CXCR2 treatment experiments that utilized female mice exclusively. Further information regarding strains and breeding is available in SI Appendix.

EAE. Passive EAE was induced as previously reported (6) and is described in more detail in SI Appendix. For induction of MHCI in vivo, Tam-WT\textsuperscript{APC}, Tam-B\textsuperscript{APC}, and Tam-B\textsuperscript{APC} xVLA-4\textsuperscript{-} or Tam-B\textsuperscript{APC} xVLA-4\textsuperscript{-} mice were treated with 5 μg Tam (Sigma-Aldrich) in 50 μL corn oil (Sigma-Aldrich) by oral gavage 3 times weekly on days –2, –0, and 2 relative to Tam treatment by oral gavage (at day 0) 3 wk after CD4 T cell transfer. Mice were scored daily for clinical disease and killed 3 d post EAE onset or 13 to 15 d post Tam.

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Flow Cytometry. Isolation and staining of single cells from various tissues for flow cytometric analysis was performed according to prior reports (6) and is detailed in SI Appendix.

CSF Collection and Analysis. Mice were avertin-anesthetized prior to CSF collection from the cisterna magna. CSF was stored in MAXYMum Recovery, low-protein-binding PCR tubes and immediately spun down and frozen. Concentration of cytokines and chemokines was assessed using the BioRad 33-plex kit analyzed on a Luminex. Concentrations of CSF were transformed logarithmically for graphing and statistical analysis purposes.

Imaging. Immunofluorescence staining for CD3 and B220 was performed as previously reported (6). Intermittent background labeling from anti-CD3 staining was noted, but consistently within the gray matter and not in a pattern confounding for cellular labeling of T cells. Background immunofluorescence was a result of anti-CD3, as control staining with secondary antibody only never resulted in excessive background signal. Staining for additional markers, including VCAM-1, Ly-6G, podoplanin, and K-67 is detailed in SI Appendix. Optical clearing of spinal cords was initiated by perfusing mice with 25 mL ice-cold phosphate buffered saline followed by 20 mL 4% paraformaldehyde (Sigma-Aldrich). CNS tissue was extracted while still encased in the skull and vertebral column, then fixed in 4% paraformaldehyde for more than 12 h. Spinal cord vertebral were decalified in 6% trichloroacetic acid (Sigma-Aldrich) for 5 d and washed with PBS prior to optical clearing using iDISCO protocol (36). Additional details including confocal imaging are provided in SI Appendix.

Data Availability. All data discussed in the paper are available via Figshare, https://doi.org/10.6084/m9.figshare.10052051.v1.

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